## Package 'notameStats'

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```
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cohens\_d

```
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cohens\_d Cohen's D

## Description

Computes Cohen's D for each feature. If time and ID are supplied, change between two time points is computed for each subject, and Cohen's d is computed from the changes.

## Usage

```
cohens_d(object, group, id = NULL, time = NULL, assay.type = NULL)
```

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#### **Arguments**

```
object a SummarizedExperiment object
group character, name of the group column
id character, name of the subject ID column
time character, name of the time column
assay.type character, assay to be used in case of multiple assays
```

#### Value

A data frame with Cohen's d for each feature.

#### **Examples**

```
data(toy_notame_set, package = "notame")
d_results <- cohens_d(notame::drop_qcs(toy_notame_set), group = "Group")
d_results_time <- cohens_d(notame::drop_qcs(toy_notame_set),
    group = "Group", time = "Time", id = "Subject_ID"
)</pre>
```

fit\_rf

Fit Random Forest

## Description

Fits a random forest, where given response column in pheno data is predicted using the features. Can be used both for classification and regression. For more information, see the documentation of randomForest. After fitting the random forest, use importance\_rf as a shortcut for getting the feature importance in random forest prediction.

## Usage

```
fit_rf(
  object,
  y,
  all_features = FALSE,
  covariates = NULL,
  importance = TRUE,
  assay.type = NULL,
  ...
)
```

## **Arguments**

```
object a SummarizedExperiment object

y character, column name of pheno data giving the dependent variable of the model

all_features logical, should all features be included in the model? if FALSE, flagged features are left out
```

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covariates character, column names of pheno data to use as covariates in the model, in addition to molecular features

importance Should importance of features be assessed?

character, assay to be used in case of multiple assays

other parameters passed to randomForest

#### Value

An object of class randomForest.

#### See Also

```
randomForest, importance_rf
```

## **Examples**

```
data(toy_notame_set, package = "notame")
rf <- fit_rf(toy_notame_set, y = "Group")
rf
importance_rf(rf)</pre>
```

fold\_change

Fold change

## Description

Computes fold change between each group for each feature.

## Usage

```
fold_change(object, group, assay.type = NULL)
```

## **Arguments**

object a SummarizedExperiment object group character, name of the group column

assay.type character, assay to be used in case of multiple assays

## Value

A data frame with fold changes for each feature.

```
data(toy_notame_set, package = "notame")
# Between groups
fc <- fold_change(toy_notame_set, group = "Group")
# Between time points
fc <- fold_change(toy_notame_set, group = "Time")</pre>
```

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importance\_rf

Feature importance in random forest

## **Description**

Extracts feature importance in random forest in a nice format.

## Usage

```
importance_rf(rf)
```

#### **Arguments**

rf

An object of class randomForest

#### Value

A data frame of feature importance.

## See Also

```
randomForest, fit_rf
```

## **Examples**

```
data(toy_notame_set, package = "notame")
rf <- fit_rf(toy_notame_set, y = "Group")
rf
importance_rf(rf)</pre>
```

muvr\_analysis

Multivariate modelling with minimally biased variable selection (MUVR)

## **Description**

A wrapper around MUVR2 (random forest, PLS(-DA)) and MUVR2\_EN (elastic net) functions from the MUVR2 package.

## Usage

```
muvr_analysis(
  object,
  y = NULL,
  id = NULL,
  multi_level = FALSE,
  multi_level_var = NULL,
  covariates = NULL,
  static_covariates = NULL,
```

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```
all_features = FALSE,
  nRep = 50,
  nOuter = 6,
  nInner = nOuter - 1,
  varRatio = 0.75,
  method = c("PLS", "RF"),
  assay.type = NULL,
   ...
)
```

## **Arguments**

object a SummarizedExperiment object

y character, column name in pheno data of the target variable

id character, column name in pheno data of the subject ID variable in case of re-

peated measurements

multi\_level logical, whether multi-level modeling should be applied, see Details

multi\_level\_var

character, column name in pheno data of the variable for splitting the data in

multi-level modeling

covariates, static\_covariates

character, column names of pheno data to use as covariates in the model, in addition to molecular features. static\_covariates are ignored for non-multi-level models. For multi-level models, the change in covariates is computed,

while static\_covariates are taken from the first time point.

all\_features logical, should all features be included in the model? if FALSE, flagged features

are left out

nRep Number of repetitions of double CV, parameter of MUVR
nOuter Number of outer CV loop segments, parameter of MUVR
Number of inner CV loop segments, parameter of MUVR

varRatio Ratio of variables to include in subsequent inner loop iteration, parameter of

MUVR

method Multivariate method. Supports 'PLS', 'RF' and 'EN' assay. type character, assay to be used in case of multiple assays

... other parameters to MUVR2 or MUVR2\_EN and getVar (when method == "EN")

#### **Details**

This function is now using the MUVR2 package, characterized as an upgrade extending the original MUVR package by the inclusion of elastic net regression (EN) and some functionality not covered by this wrapper. Elastic net regression supports covariate adjustment by suppressing regularization of specified features from the regularization procedure. Note that this is different from simply including covariates such as sex. EN also differs from PLS and RF in that no recursive variable elimination is performed, so an additional scheme is used to obtain the 'min', 'mid' and 'max' models using getVar.

Sex would be entered as a static covariate, since the change in sex is zero for all individuals, so computing the change and using that as a covariate does not make sense.

Note that there are several more plots available in MUVR2 for inspecting the results, notably plotMV, plotStability and plotVIRank Many of these return different plots depending on the model specification.

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#### Value

A MUVR object.

#### See Also

MUVR2 MUVR2\_EN getVar plotMV plotStability plotVIRank plotVAL

#### **Examples**

```
data(toy_notame_set, package = "notame")
ex_set <- notame::drop_qcs(toy_notame_set)[1:10, ]</pre>
ex_set$Injection_order <- as.numeric(ex_set$Injection_order)</pre>
# Simple PLS regression model
pls_model <- muvr_analysis(ex_set,</pre>
  y = "Injection_order", nRep = 2, method = "PLS")
# RF classification with covariate and repeated measures (not longitudinal)
rf_model <- muvr_analysis(ex_set, y = "Group", id = "Subject_ID",</pre>
  nRep = 2, method = "RF", covariates = "Injection_order")
# RF classification on multilevel variable comparing levels of y
rf_model_ <- muvr_analysis(ex_set,</pre>
  y = "Group", multi_level = TRUE, id = "Subject_ID",
  multi_level_var = "Time", method = "RF", nRep = 2)
# EN regression on multilevel variable with covariate and static covariate
ex_set$Group <- as.numeric(ex_set$Group)</pre>
en_model <- muvr_analysis(ex_set, id = "Subject_ID",</pre>
multi_level = TRUE, multi_level_var = "Time",
covariates = "Injection_order", static_covariates = "Group",
 method = "EN", nRep = 2)
```

perform\_auc

Area under curve

#### **Description**

Compute area under curve (AUC) for each subject and feature. Creates a pseudo SummarizedExperiment object, where the "samples" are subjects (or subject/group combinations in case the same subjects are submitted to different treatments) and the "abundances" are AUCs. This object can then be used to compute results of e.g. t-tests of AUCs between groups.

## Usage

```
perform_auc(object, time, subject, group, assay.type = NULL)
```

#### **Arguments**

```
object a SummarizedExperiment object
time, subject, group
column names of pheno data holding time, subject and group labels
assay.type character, assay to be used in case of multiple assays
```

#### Value

A pseudo SummarizedExperiment object with the AUCs.

#### See Also

auc

## **Examples**

```
perform_correlation_tests
```

Correlation test

#### **Description**

Performs a correlation test between two sets of variables. All the variables must be either feature names or column names of pheno data (sample information). There are two ways to use this function: either provide a set of variables as x, and all correlations between those variables are computed. Or provide two distinct sets of variables x, y and correlations between each x variable and each y variable are computed.

#### Usage

```
perform_correlation_tests(
  object,
    x,
    y = x,
    id = NULL,
    object2 = NULL,
    fdr = TRUE,
    all_pairs = TRUE,
    duplicates = FALSE,
    assay.type1 = NULL,
    assay.type2 = NULL,
    ...
)
```

## **Arguments**

```
object a SummarizedExperiment object
x character vector, names of variables to be correlated
y character vector, either identical to x (the default) or a distinct set of variables to be correlated against x
```

id	character, column name for subject IDs. If provided, the correlation will be computed using the rmcorr package	
object2	optional second object. If provided, x variables will be taken from object and y variables will be taken from object2. Both objects should have the same number of samples.	
fdr	logical, whether p-values from the correlation test should be adjusted with FDR correction	
all_pairs logical, whether all pairs between x and y should be tested. If FALSE, x a give the exact pairs of variables to test, and should have the same length.		
duplicates	logical, whether correlations should be duplicated. If TRUE, each correlation will be included in the results twice, where the order of the variables '(which is x and which is y) is changed. Can be useful for e.g. plotting a heatmap of the results, see examples of plot_effect_heatmap.	
assay.type1	character, assay of object(1) to be used in case of multiple assays	
assay.type2	character, assay of object2 to be used in case of multiple assays	
	other parameters passed to cor.test, such as method	

#### Value

A data frame with the results of correlation tests: the pair of variables, correlation coefficient and p-value.

## See Also

```
cor.test, rmcorr
```

```
data(toy_notame_set, package = "notame")
# Correlations between all features
correlations <- perform_correlation_tests(toy_notame_set,</pre>
  x = rownames(toy_notame_set), id = "Subject_ID")
# Spearman Correlations between features and sample information variables
\mbox{\tt\#} Drop QCs and convert time to numeric
no_qc <- notame::drop_qcs(toy_notame_set)</pre>
no_qc$Time <- as.numeric(no_qc$Time)</pre>
correlations <- perform_correlation_tests(no_qc,</pre>
 x = rownames(toy_notame_set),
 y = c("Time", "Injection_order"), method = "spearman"
# Correlations between variables from two distinct objects
cross_object_cor <-perform_correlation_tests(toy_notame_set,</pre>
  x = rownames(toy_notame_set),
  object2 = toy_notame_set,
 y = rownames(toy_notame_set),
  all_pairs = FALSE
```

## **Description**

Performs Bartlett's, Levene's and Fligner-Killeen tests for equality of variances.

## Usage

```
perform_homoscedasticity_tests(
  object,
  formula_char,
  all_features = FALSE,
  assay.type = NULL
)
```

## **Arguments**

```
object a SummarizedExperiment object

formula_char character, the formula to be used in the linear model (see Details)

all_features should all features be included in FDR correction?

assay.type character, assay to be used in case of multiple assays
```

#### **Details**

The model is fit on combined\_data(object). Thus, column names in pheno data can be specified. To make the formulas flexible, the word "Feature" must be used to signal the role of the features in the formula. "Feature" will be replaced by the actual Feature IDs during model fitting. For example, if testing for equality of variances in study groups, use "Feature ~ Group".

#### Value

A data frame with the results.

## See Also

```
bartlett.test, leveneTest, fligner.test
```

```
data(toy_notame_set, package = "notame")
perform_homoscedasticity_tests(toy_notame_set,
  formula_char = "Feature ~ Group")
```

perform\_kruskal\_wallis

```
perform_kruskal_wallis
```

Kruskal-Wallis rank-sum test

## **Description**

Performs Kruskal-Wallis rank-sum test for equality.

## Usage

```
perform_kruskal_wallis(
  object,
  formula_char,
  all_features = FALSE,
  assay.type = NULL
)
```

## **Arguments**

```
object a SummarizedExperiment object

formula_char character, the formula to be used in the linear model (see Details)

all_features should all features be included in FDR correction?

assay.type character, assay to be used in case of multiple assays
```

## **Details**

The model is fit on combined\_data(object). Thus, column names in pheno data can be specified. To make the formulas flexible, the word "Feature" must be used to signal the role of the features in the formula. "Feature" will be replaced by the actual Feature IDs during model fitting. For example, if testing for equality of means in study groups, use "Feature ~ Group".

#### Value

A data frame with the results.

#### See Also

```
kruskal.test
```

```
data(toy_notame_set, package = "notame")
perform_kruskal_wallis(toy_notame_set, formula_char = "Feature ~ Group")
```

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ar models
-----------

## **Description**

Fits a linear model separately for each feature. Returns all relevant statistics.

## Usage

```
perform_lm(object, formula_char, all_features = FALSE, assay.type = NULL, ...)
```

#### **Arguments**

```
object a SummarizedExperiment object

formula_char character, the formula to be used in the linear model (see Details)

all_features should all features be included in FDR correction?

assay.type character, assay to be used in case of multiple assays

additional parameters passed to lm
```

#### **Details**

The linear model is fit on combined\_data(object). Thus, column names in pheno data can be specified. To make the formulas flexible, the word "Feature" must be used to signal the role of the features in the formula. "Feature" will be replaced by the actual Feature IDs during model fitting, see the example.

## Value

A data frame with one row per feature, with all the relevant statistics of the linear model as columns.

#### See Also

1m

```
data(toy_notame_set, package = "notame")
# A simple example without QC samples
# Features predicted by Group and Time
lm_results <- perform_lm(notame::drop_qcs(toy_notame_set),
    formula_char = "Feature ~ Group + Time")</pre>
```

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perform_lmer	Linear mixed models
--------------	---------------------

## **Description**

Fits a linear mixed model separately for each feature. Returns all relevant statistics.

#### Usage

```
perform_lmer(
  object,
  formula_char,
  all_features = FALSE,
  ci_method = c("Wald", "profile", "boot"),
  test_random = FALSE,
  assay.type = NULL,
  ...
)
```

#### **Arguments**

```
object a SummarizedExperiment object

formula_char character, the formula to be used in the linear model (see Details)

all_features should all features be included in FDR correction?

ci_method The method for calculating the confidence intervals as in confint

test_random logical, whether tests for the significance of the random effects should be performed

assay.type character, assay to be used in case of multiple assays

... additional parameters passed to lmer
```

#### **Details**

The model is fit on combined\_data(object). Thus, column names in pheno data can be specified. To make the formulas flexible, the word "Feature" must be used to signal the role of the features in the formula. "Feature" will be replaced by the actual Feature IDs during model fitting, see the example. With bootstrap ("boot") confidence intervals, the results are reproducible if RNGseed is set for the BiocParallel backend.

#### Value

A data frame with one row per feature, with all the relevant statistics of the linear mixed model as columns.

## See Also

lmer for model specification

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#### **Examples**

```
data(toy_notame_set, package = "notame")
# A simple example without QC samples
# Features predicted by Group and Time as fixed effects with Subject ID as a
# random effect
lmer_results <- perform_lmer(notame::drop_qcs(toy_notame_set),
    formula_char = "Feature ~ Group + Time + (1 | Subject_ID)",
    ci_method = "Wald"
)</pre>
```

perform\_lm\_anova

Linear models ANOVA table

## **Description**

Fits a linear model separately for each feature and compute an ANOVA table. Returns all relevant statistics.

#### Usage

```
perform_lm_anova(
   object,
   formula_char,
   all_features = FALSE,
   lm_args = NULL,
   anova_args = NULL,
   assay.type = NULL
)
```

#### **Arguments**

object a SummarizedExperiment object

formula\_char character, the formula to be used in the linear model (see Details)

all\_features should all features be included in FDR correction?

lm\_args list of arguments to lm, list names should be parameter names

anova\_args list of arguments to anova, list names should be parameter names

character, assay to be used in case of multiple assays

#### **Details**

The linear model is fit on combined\_data(object). Thus, column names in pheno data can be specified. To make the formulas flexible, the word "Feature" must be used to signal the role of the features in the formula. "Feature" will be replaced by the actual Feature IDs during model fitting, see the example.

## Value

A data frame with one row per feature, with all the relevant statistics of the linear model as columns.

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#### See Also

1m

#### **Examples**

```
data(toy_notame_set, package = "notame")
# A simple example without QC samples
# Features predicted by Group and Time
lm_anova_results <- perform_lm_anova(notame::drop_qcs(toy_notame_set),
    formula_char = "Feature ~ Group + Time")</pre>
```

perform\_logistic

Logistic regression

## **Description**

Fits a logistic regression model separately for each feature. Returns all relevant statistics.

## Usage

```
perform_logistic(
  object,
  formula_char,
  all_features = FALSE,
  assay.type = NULL,
   ...
)
```

## **Arguments**

```
object a SummarizedExperiment object

formula_char character, the formula to be used in the linear model (see Details)

all_features should all features be included in FDR correction?

assay.type character, assay to be used in case of multiple assays

... additional parameters passed to glm
```

## **Details**

The logistic regression model is fit on combined\_data(object). Thus, column names in pheno data can be specified. To make the formulas flexible, the word "Feature" must be used to signal the role of the features in the formula. "Feature" will be replaced by the actual Feature IDs during model fitting, see the example.

## Value

A data frame with one row per feature, with all the relevant statistics of the linear model as columns.

#### See Also

glm

#### **Examples**

```
data(toy_notame_set, package = "notame")
# A simple example without QC samples
# Time predicted by features
logistic_results <- perform_logistic(notame::drop_qcs(toy_notame_set),
    formula_char = "Time ~ Feature + Group"
)</pre>
```

perform\_non\_parametric

Pairwise and paired non-parametric tests

#### **Description**

Performs pairwise and paired non-parametric tests. The default is Mann- Whitney U test, use is\_paired for Wilcoxon signed rank tests.

## Usage

```
perform_non_parametric(
  object,
  formula_char,
  is_paired = FALSE,
  id = NULL,
  all_features = FALSE,
  assay.type = NULL,
  ...
)
```

#### **Arguments**

```
object a SummarizedExperiment object

formula_char character, the formula to be used in the tests

is_paired logical, use paired test

id character, name of the subject identification column for paired version

all_features should all features be included in FDR correction?

assay.type character, assay to be used in case of multiple assays

other parameters passed to test wilcox.test
```

## Details

P-values of each comparison are corrected separately from each other. The model is fit on combined\_data(object). Thus, column names in pheno data can be specified. To make the formulas flexible, the word "Feature" must be used to signal the role of the features in the formula. "Feature" will be replaced by the actual features during model fitting. For example, if testing for equality of means in study groups, use "Feature ~ Group".

#### Value

A data frame with the results.

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#### See Also

```
wilcox.test
```

#### **Examples**

```
data(toy_notame_set, package = "notame")
# Including QCs as a study group for example for pairwise tests
mann_whitney_results <- perform_non_parametric(toy_notame_set,
    formula_char = "Feature ~ Group")
# Using paired mode (pairs with QC are skipped as there are no common IDs in
# 'toy_notame_set')
wilcoxon_signed_results <- perform_non_parametric(toy_notame_set,
    formula_char = "Feature ~ Time",
    is_paired = TRUE,
    id = "Subject_ID")
# Only two groups
mw_results <-perform_non_parametric(notame::drop_qcs(toy_notame_set),
    formula_char = "Feature ~ Group")</pre>
```

perform\_oneway\_anova

Welch's ANOVA and classic ANOVA

#### **Description**

Performs ANOVA with Welch's correction as default, to deal with heterogeneity of variances. Can also perform classic ANOVA with assumption of equal variances. Uses base R function oneway.test.

## Usage

```
perform_oneway_anova(
  object,
  formula_char,
  all_features = FALSE,
  assay.type = NULL,
   ...
)
```

#### **Arguments**

```
object a SummarizedExperiment object
formula_char character, the formula to be used in the linear model (see Details).
all_features should all features be included in FDR correction?
assay.type character, assay to be used in case of multiple assays
other parameters to oneway.test
```

#### **Details**

The model is fit on combined\_data(object). Thus, column names in pheno data can be specified. To make the formulas flexible, the word "Feature" must be used to signal the role of the features in the formula. "Feature" will be replaced by the actual Feature IDs during model fitting. For example, if testing for equality of means in study groups, use "Feature ~ Group".

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#### Value

A data frame with the results.

#### See Also

```
oneway.test
```

## **Examples**

```
data(toy_notame_set, package = "notame")
perform_oneway_anova(toy_notame_set, formula_char = "Feature ~ Group")
```

perform\_permanova

**PERMANOVA** 

## **Description**

Performs permutational multivariate analysis of variance. Uses package called PERMANOVA.

## Usage

```
perform_permanova(
  object,
  group,
  all_features = FALSE,
  transform = "Standardize columns",
  coef = "Pythagorean",
  assay.type = NULL,
  ...
)
```

## **Arguments**

```
object a SummarizedExperiment object
group character, name of the column to compare
all_features should all features be included?
transform Transformation to use in IniTransform. By default uses "Standardize columns".
coef Coefficient to calculate continuous distances in IniTransform. By default uses Pythagorean distances.
assay.type character, assay to be used in case of multiple assays
... other parameters to PERMANOVA
```

## Value

A PERMANOVA object.

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## **Examples**

```
data(toy_notame_set, package = "notame")
permanova_res <- perform_permanova(
  notame::drop_qcs(toy_notame_set),
  group = "Group")</pre>
```

perform\_t\_test

Pairwise and paired t-tests

## **Description**

Performs pairwise and paired t-tests. The R default is Welch's t-test (unequal variances), use var.equal = TRUE for Student's t-test. Use  $is\_paired$  for paired t-tests.

## Usage

```
perform_t_test(
  object,
  formula_char,
  is_paired = FALSE,
  id = NULL,
  all_features = FALSE,
  assay.type = NULL,
  ...
)
```

## **Arguments**

```
object a SummarizedExperiment object

formula_char character, the formula to be used in the linear model (see Details)

is_paired logical, use paired t-test

id character, name of the subject identification column for paired version

all_features should all features be included in FDR correction?

assay.type character, assay to be used in case of multiple assays

other parameters passed to t.test
```

#### **Details**

P-values of each comparison are corrected separately from each other.

## Value

A data frame with the results.

#### See Also

```
t.test
```

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#### **Examples**

```
data(toy_notame_set, package = "notame")
# Including QCs as a study group for example
t_test_results <- perform_t_test(toy_notame_set,
    formula_char = "Feature ~ Group")
# Using paired mode (pairs with QC are skipped as there are no common IDs in
# 'toy_notame_set')
t_test_results <- perform_t_test(toy_notame_set,
    formula_char = "Feature ~ Time", is_paired = TRUE, id = "Subject_ID")
# Only two groups
t_test_results <- perform_t_test(notame::drop_qcs(toy_notame_set),
    formula_char = "Feature ~ Group")</pre>
```

pls

PLS

## Description

Simple wrappers for fitting a PLS model using mixOmics package. The result can then be passed to many of the mixOmics functions for prediction, performance evaluation etc.

## Usage

```
mixomics_pls(
  object,
  у,
  ncomp,
  all_features = FALSE,
  covariates = NULL,
  assay.type = NULL,
)
mixomics_pls_optimize(
  object,
  у,
  ncomp,
  plot_perf = FALSE,
  folds = 5,
  nrepeat = 50,
  all_features = FALSE,
  covariates = NULL,
  assay.type = NULL,
)
mixomics_spls_optimize(
  object,
  у,
  ncomp,
```

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```
plot_perf = FALSE,
  n_features = c(seq_len(10), seq(20, 300, 10)),
  folds = 5,
    nrepeat = 50,
    all_features = FALSE,
    covariates = NULL,
    assay.type = NULL,
    ...
)
```

## **Arguments**

a SummarizedExperiment object
character vector, column names of the grouping variable to predict
number of X components
logical, should all features be included in the model? if FALSE, flagged features are left out
character, column names of pheno datato use as covariates in the model, in addition to molecular features
character, assay to be used in case of multiple assays
any parameters passed to pls or spls
plot performance of models in cross-validation
the number of folds to use in k-fold cross validation
the number of times to repeat the cross validation. Lower this for faster testing.
the number of features to try for each component

## Details

- mixomics\_pls A simple PLS model with set number of components and all features
- mixomics\_pls\_optimize Test different numbers of components
- mixomics\_spls\_optimize sPLS model: Test different numbers of components and features

#### Value

An object of class "mixo\_pls" or "mixo\_spls". For the optimized and sparse models, a list with object of class "mixo\_plsda" and a performance plot.

#### See Also

```
pls, perf, spls, tune.spls
```

```
data(toy_notame_set, package = "notame")
pls_res <- mixomics_pls(toy_notame_set, y = "Injection_order", ncomp = 3)
# Cross-validation repeated only 5 times for quick run time
pls_opt <- mixomics_pls_optimize(toy_notame_set,
    y = "Injection_order", ncomp = 3, nrepeat = 5)
spls_opt <- mixomics_spls_optimize(toy_notame_set,
    y = "Injection_order", ncomp = 3,
    n_features = c(1:10, 12, 15, 20), nrepeat = 5</pre>
```

 $pls_da$ 

```
# Plot score plot of any final model
mixOmics::plotIndiv(pls_res,
   comp = seq_len(2), group = toy_notame_set$Group,
   ind.names = FALSE, title = "PLS scores plot", legend = TRUE)
# Proportion of variance explained
pls_res$prop_expl_var$X[seq_len(2)] |> round(digits = 3) * 100
```

pls\_da

PLS-DA

## **Description**

A simple wrapper for fitting a PLS-DA model using mixOmics package. The object can then be passed to many of the mixOmics functions for prediction, performance evaluation etc.

- mixomics\_plsda A simple PLS-DA model with set number of components and all features
- mixomics\_plsda\_optimize Test different numbers of components, choose the one with minimal balanced error rate
- mixomics\_splsda\_optimize Test different numbers of components and features, choose the one with minimal balanced error rate

## Usage

```
mixomics_plsda(
  object,
  у,
  ncomp,
  all_features = FALSE,
  covariates = NULL,
  assay.type = NULL,
)
mixomics_plsda_optimize(
  object,
  у,
  ncomp,
  plot_perf = FALSE,
  folds = 5,
  nrepeat = 50,
  all_features = FALSE,
  covariates = NULL,
  assay.type = NULL,
mixomics_splsda_optimize(
  object,
```

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```
y,
ncomp,
dist,
plot_perf = FALSE,
n_features = c(seq_len(10), seq(20, 300, 10)),
folds = 5,
nrepeat = 50,
all_features = FALSE,
covariates = NULL,
assay.type = NULL,
...
)
```

## **Arguments**

object	a SummarizedExperiment object
У	character, column name of the grouping variable to predict
ncomp	the number of X components
all_features	logical, should all features be included in the model? if FALSE, flagged features are left out
covariates	character, column names of pheno data to use as covariates in the model, in addition to molecular features
assay.type	character, assay to be used in case of multiple assays
	any parameters passed to plsda
plot_perf	plot performance of models in cross-validation
folds	the number of folds to use in k-fold cross validation
nrepeat	the number of times to repeat the cross validation. Lower this for faster testing.
dist	the distance metric to use, one of "max.dist", "mahalanobis.dist", "centroids.dist". use mixomics_plsda_optimize to find the best distance metric
n_features	the number of features to try for each component

## Value

An object of class "mixo\_plsda" or for the optimized and sparse models, a list with object of class "mixo\_plsda" and a performance plot.

## See Also

```
plsda, perf, splsda, tune.splsda
```

```
data(toy_notame_set, package = "notame")
noqc <- notame::drop_qcs(toy_notame_set)
plsda_res <- mixomics_plsda(noqc, y = "Group", ncomp = 2)
# Cross-validation repeated only 5 times for quick run time
set.seed(38)
plsda_opt <- mixomics_plsda_optimize(noqc,
    y = "Group", ncomp = 3, nrepeat = 5
)
set.seed(38)</pre>
```

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```
splsda_opt <- mixomics_splsda_optimize(noqc,
    y = "Group", dist = "max.dist", ncomp = 2,
    n_features = c(1:10, 12, 15, 20), nrepeat = 5
)
# Plot PLS-DA scores
mixOmics::plotIndiv(plsda_res,
    comp = seq_len(2), group = notame::drop_qcs(toy_notame_set)$Group,
    ind.names = FALSE, title = "PLS-DA scores plot", legend = TRUE,
    ellipse = TRUE)
# Plot prediction areas
background <- mixOmics::background.predict(plsda_res,
    comp.predicted = 2, dist = "max.dist")
mixOmics::plotIndiv(plsda_res,
    comp = seq_len(2), group = notame::drop_qcs(toy_notame_set)$Group,
    ind.names = FALSE,
    title = "prediction areas", legend = TRUE, background = background)</pre>
```

 $summarize\_results$ 

Statistics cleaning

## Description

Uses regexp to remove unnecessary columns from statistics results data frame. Can also rename columns effectively.

## Usage

```
summarize_results(
  df,
  remove = c("Intercept", "CI95", "Std_error", "t_value", "z_value", "R2"),
  rename = NULL,
  summary = TRUE,
  p_limit = 0.05,
  fdr = TRUE
)
```

## **Arguments**

df	data frame, statistics results
remove	list, should contain strings that are matching to unwanted columns
rename	named list, names should contain matches that are replaced with values
summary	logical, should summary columns be added
p_limit	numeric, limit for p-values to be counted
fdr	logical, should summary be done with fdr-fixed values

## Value

A data frame with removed and/or renamed columns.

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#### **Examples**

```
data(toy_notame_set, package = "notame")
# Simple manipulation to linear model results
lm_results <- perform_lm(notame::drop_qcs(toy_notame_set),
    formula_char = "Feature ~ Group + Time")
lm_results <- summarize_results(lm_results,
    rename = c("GroupB" = "GroupB_vs_A", "Time2" = "Time2_vs_1")
)</pre>
```

summary\_statistics

Summary statistics

## **Description**

Computes summary statistics for each feature, possibly grouped by a factor. The statistics include mean, standard deviation (sd), median, median absolute deviation (mad), minimum (min), maximum (max) as well as 25

#### Usage

```
summary_statistics(object, grouping_cols = NULL, assay.type = NULL)
```

#### **Arguments**

```
object a SummarizedExperiment object
```

grouping\_cols character vector, the columns by which grouping should be done. Use NA to

compute statistics without grouping.

assay.type character, assay to be used in case of multiple assays

## Value

A data frame with the summary statistics.

```
data(toy_notame_set, package = "notame")
# Group by "Group"
sum_stats <- summary_statistics(toy_notame_set, grouping_cols = "Group")
# Group by Group and Time
sum_stats <- summary_statistics(toy_notame_set,
    grouping_cols = c("Group", "Time"))
# No Grouping
sum_stats <- summary_statistics(toy_notame_set)</pre>
```

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