# Package 'hdxmsqc'

December 2, 2025

```
mass spectrometry experiments
Version 1.6.0
Description The hdxmsqc package enables us to analyse and visualise the quality
      of HDX-MS experiments. Either as a final quality check before downstream
      analysis and publication or as part of a interative procedure to determine
      the quality of the data. The package builds on the QFeatures and Spectra
     packages to integrate with other mass-spectrometry data.
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Type Package

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# Description

A small HDX-MS dataset for BRD4 in apo state and in complex with IBET151

# Author(s)

My Name <ocorook@gmail.com>

BRD4df\_full

This is data to be included in my package

# Description

A complete HDX-MS dataset for BRD4 in apo state and in complex with IBET151

# Author(s)

My Name <ococrook@gmail.com>

chargeCorrelationHdx 3

 ${\it charge Correlation Hdx} \qquad {\it Charge states should have correlated incorperation but they need not} \\ be {\it exactly the same}$ 

#### **Description**

Charge states should have correlated incorperation but they need not be exactly the same

#### Usage

```
chargeCorrelationHdx(object, experiment = NULL, timepoints = NULL)
```

#### **Arguments**

object An object of class QFeatures

experiment A character vector indicating the experimental conditions timepoints A numeric vector indicating the experimental timepoints

#### Author(s)

Oliver Crook

#### **Examples**

```
data("BRD4df_full")
BRD4df_filtered <- isMissingAtRandom(object = BRD4df_full)
BRD4df_full_imputed <- impute(BRD4df_filtered, method = "zero", i = 1)
experiment <- c("wt", "iBET")
timepoints <- rep(c(0, 15, 60, 600, 3600, 14000), each = 3)
monoStat <- chargeCorrelationHdx(object = BRD4df_full_imputed,
experiment = experiment,
timepoints = timepoints)</pre>
```

compatibleUptake

Check whether deuterium uptakes are compatible with difference overlapping sequences.

## **Description**

Check whether deuterium uptakes are compatible with difference overlapping sequences.

#### Usage

```
compatibleUptake(object, overlap = 5, experiment = NULL, timepoints = NULL)
```

#### **Arguments**

object	An object of class QFeatures

overlap How much overlap is required to check consistentcy. Default is sequences within

5 residues

experiment A character vector indicating the experimental conditions timepoints A numeric vector indicating the experimental timepoints

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#### Author(s)

Oliver Crook

# **Examples**

```
data("BRD4df")
result <- compatibleUptake(BRD4df, experiment = 1, timepoints = 1)</pre>
```

computeMassError

Empirical versus theoretical mass errors

# Description

Empirical versus theoretical mass errors

# Usage

```
computeMassError(object, eCentroid = "Exp.Cent", tCentroid = "Theor.Cent")
```

# Arguments

object An object of class QFeatures

eCentroid character string indicating column identifier for experimental centroid

tCentroid character string indicating column identifier for theoretical centroid

## Value

The error difference between the empirical and theoretical centroid

#### Author(s)

Oliver Crook

```
data("BRD4df")
result <- computeMassError(BRD4df, "Exp.Cent", "Theor.Cent")
head(result)</pre>
```

computeMonotoneStats 5

computeMonotoneStats Monotonicity based outlier detection.

## **Description**

Monotonicity based outlier detection.

## Usage

```
computeMonotoneStats(object, experiment = NULL, timepoints = NULL)
```

# Arguments

object An object of class QFeatures

experiment A character vector indicating the experimental conditions timepoints A numeric vector indicating the experimental timepoints

## Author(s)

Oliver Crook

# **Examples**

```
data("BRD4df")
result <- computeMonotoneStats(BRD4df, experiment = 1, timepoint = 1)</pre>
```

exchangeableAmides

Compute exchangeable amides.

# Description

Computes the number of exchangeable amides based on the sequnece

## Usage

```
exchangeableAmides(sequence)
```

# Arguments

sequence

The sequence of the peptide

#### Value

Returns a numeric indicating the number of exchangeable amides

```
exchangeableAmides(sequence = "HDAEHAHEAPRKL")
```

6 fourierIsotope

fourierIsotope

fourier transform approach to computing isotopic distribution

# Description

fourier transform approach to computing isotopic distribution

# Usage

```
fourierIsotope(
  elements,
  incorp = 0,
  num_exch_sites = 0,
  charge = 1,
  isotopes = NULL
)
```

# Arguments

elements A list of elements

incorp The deuterium incoperation

 $num\_exch\_sites$  The number of exchangable amides. Default is 0.

charge The charge state of the peptide

isotopes The number of isotopes to compute. The default is NULL, in whiich a default

heuristic is used to make a good guess that covers the expected peaks.

## Value

A list of mass and intensity value corresponding to the isotope distribution

## Author(s)

Oliver Crook

```
fourierIsotope(c(C = 0, H = 2, N = 0, O = 1, S = 0, P = 0))
```

generateSpectra 7

generateSpectra

generate Spectra using a fourier transform

## **Description**

generate Spectra using a fourier transform

## Usage

```
generateSpectra(
   sequences,
   incorps,
   charges,
   customs = list(code = NULL, elements = NULL)
)
```

#### **Arguments**

sequences A vector of peptide sequences
incorps A vector of deuterium incoperation
charges A vector of charge states of the peptide
customs Custom elements supplied as a list

#### Value

A Spectra object corresponding to the isotope distributions

#### Author(s)

Oliver Crook

# **Examples**

```
generateSpectra(sequence = "HDAEHAHEAPRKL", incorps = c(0.5), charges = 2)
```

hdxmsqc

A package to perfrom quality control for mass-spectrometry based hydrogen deuterium exchange experiment.

## **Description**

'hdxmsqc' provides the functionality to assess the quality and perform quality control of HDX-MS experiments. Raw and processed data can be visualized and analyzed to identify potential issues with the data. The package is designed to work with data from any HDX-MS platform. Typically, users will have exported results from either HDExaminer or DynamX software. There is not need to filter the data in either of those software systems.

#### Author(s)

Oliver Crook

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imTimeOutlier

Ion Mobility time based outlier analysis

#### **Description**

Ion Mobility time based outlier analysis

# Usage

```
imTimeOutlier(
  object,
  rightIMS = "rightIMS",
  leftIMS = "leftIMS",
  searchIMS = "Search.IMS"
)
```

## **Arguments**

object An object of class QFeatures

rightIMS A string indicating the right boundary of the ion mobility separation time. De-

faults is "rightIMS".

leftIMS A string indicating the left boundary of the ion mobility separation time. Default

is "leftIMS".

searchIMS A string indicating the actual ion mobility search time. The default is "Search.IMS"

# Author(s)

Oliver Crook

# **Examples**

```
data("BRD4df_full")
BRD4df_filtered <- isMissingAtRandom(object = BRD4df_full)
BRD4df_full_imputed <- impute(BRD4df_filtered, method = "zero", i = 1)
imTimeOutlier(object = BRD4df_full_imputed)</pre>
```

intensityOutliers

Intensity based deviations

## Description

Intensity based deviations

## Usage

```
intensityOutliers(object, fcolIntensity = "Max.Inty")
```

isMissingAtRandom 9

## **Arguments**

object An object of class QFeatures

fcolIntensity character to intensity intensity columns. Default is "Max.Inty" and uses regular

expressions to find relevant columns

# Value

The Cook's distance to characterise outleirs

#### Author(s)

Oliver Crook

## **Examples**

```
data("BRD4df_full")
intensityOutliers(BRD4df_full)
```

isMissingAtRandom

Missing at random versus missing not at random

## **Description**

Missing at random versus missing not at random

# Usage

```
isMissingAtRandom(object, threshold = NULL, filter = TRUE)
```

# **Arguments**

object An object of class QFeatures

threshold A threshold indicated how many missing values indicate whether missingness is

not at random. Default is NULL, which means leads to a threshold which is half

the number of columns.

filter A logial indicating whether to filter out data that is deemed missing not at ran-

dom

data("BRD4df\_full")

isMissingAtRandom(BRD4df\_full)

#### Value

Adds a missing not at random indicator column

## Author(s)

Oliver Crook

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```
isotopicDistributionHDXfourier
```

fourier transform approach to computing isotopic distribution

# **Description**

fourier transform approach to computing isotopic distribution

# Usage

```
isotopicDistributionHDXfourier(
  sequence,
  incorp = 0,
  charge = 1,
  custom = list(code = NULL, elements = NULL)
)
```

# Arguments

sequence A peptide

incorp The deuterium incoperation charge The charge state of the peptide

custom amino acids can be provided here provide a list of the elements.

#### Value

A list of mass and intensity value corresponding to the isotope distribution

#### Author(s)

Oliver Crook

## **Examples**

```
isotopicDistributionHDXfourier(sequence = "HDAEHAHEAPRKL")
```

plotImTimeOutlier

Ion Mobility time based outlier analysis

## **Description**

Ion Mobility time based outlier analysis

## Usage

```
plotImTimeOutlier(
  object,
  rightIMS = "rightIMS",
  leftIMS = "leftIMS",
  searchIMS = "Search.IMS"
)
```

plotIntensityOutliers 11

## **Arguments**

object An object of class QFeatures

rightIMS A string indicating the right boundary of the ion mobility separation time. De-

faults is "rightIMS".

leftIMS A string indicating the left boundary of the ion mobility separation time. Default

is "leftIMS".

searchIMS A string indicating the actual ion mobility search time. The default is "Search.IMS"

#### Author(s)

Oliver Crook

#### **Examples**

```
library(RColorBrewer)
data("BRD4df_full")
BRD4df_filtered <- isMissingAtRandom(object = BRD4df_full)
BRD4df_full_imputed <- impute(BRD4df_filtered, method = "zero", i = 1)
plotImTimeOutlier(object = BRD4df_full_imputed)</pre>
```

plotIntensityOutliers Intensity based deviation plot

## **Description**

Intensity based deviation plot

#### Usage

```
plotIntensityOutliers(object, fcolIntensity = "Max.Inty")
```

## **Arguments**

object An object of class QFeatures

 $fcolIntensity \quad character\ to\ intensity\ intensity\ columns.\ Default\ is\ "Max.Inty"\ and\ uses\ regular$ 

expressions to find relevant columns

#### Value

A ggplot2 object showing intensity based outliers

# Author(s)

Oliver Crook

```
data("BRD4df_full")
library(RColorBrewer)
plotIntensityOutliers(BRD4df_full)
```

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plotMassError

Mass error plot

## **Description**

Mass error plot

## Usage

```
plotMassError(object, eCentroid = "Exp.Cent", tCentroid = "Theor.Cent")
```

## **Arguments**

object An object of class QFeatures

eCentroid character string indicating column identifier for experimental centroid tCentroid character string indicating column identifier for theoretical centroid

#### Value

a ggplot2 object which can be used to visualise the

#### Author(s)

Oliver Crook

## **Examples**

```
library(RColorBrewer)
data("BRD4df")
result <- plotMassError(BRD4df, "Exp.Cent", "Theor.Cent")</pre>
```

plotMissing

missing value plot

## **Description**

```
missing value plot
```

## Usage

```
plotMissing(object, ...)
```

# Arguments

object An object of class QFeatures
... Additional arguemnts to pheatmap

## Value

a pheatmap showing missing values

plotMonotoneStat 13

#### Author(s)

Oliver Crook

## **Examples**

```
data("BRD4df_full")
library(pheatmap)
library(RColorBrewer)
plotMissing(BRD4df_full)
```

plotMonotoneStat

Monotonicity based outlier detection, plot.

## **Description**

Monotonicity based outlier detection, plot.

# Usage

```
plotMonotoneStat(object, experiment = NULL, timepoints = NULL)
```

## **Arguments**

object An object of class QFeatures

experiment A character vector indicating the experimental conditions timepoints A numeric vector indicating the experimental timepoints

## Author(s)

Oliver Crook

```
library("RColorBrewer")
data("BRD4df_full")
experiment <- c("wt", "iBET")
timepoints <- rep(c(0, 15, 60, 600, 3600, 14000), each = 3)
monoStat <- computeMonotoneStats(object = BRD4df_full,
experiment = experiment,
timepoints = timepoints)</pre>
```

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plotrTimeOutliers

Retention time based analysis

# Description

Retention time based analysis

# Usage

```
plotrTimeOutliers(
  object,
  leftRT = "leftRT",
  rightRT = "rightRT",
  searchRT = "Search.RT"
)
```

# Arguments

object	An object of class QFeatures
leftRT	A character indicated pattern associated with left boundary of retention time search. Default is "leftRT".
rightRT	A character indicated pattern associated with right boundary of retneton time search. Default is "rightRT".
searchRT	The actual search retention time pattern. Default is "Search.RT"

## Value

a ggplot2 object showing distribution of retention time windows.

## Author(s)

Oliver Crook

```
data("BRD4df_full")
library(RColorBrewer)
plotrTimeOutliers(BRD4df_full)
```

processHDE 15

processHDE	Function to curate and HDExaminer file so that in contains all the information in a sensible format. This object can then be straightforwardly passed to a object of class QFeatures

# Description

Function to curate and HDExaminer file so that in contains all the information in a sensible format. This object can then be straightforwardly passed to a object of class QFeatures

# Usage

```
processHDE(HDExaminerFile, proteinStates = NULL)
```

# **Arguments**

```
HDExaminerFile an object of class data.frame containing an HDExaminer data proteinStates a character vector indicating the protein states
```

#### Value

A wide format data frame with HDExaminer data

#### Author(s)

Oliver Crook

# **Examples**

```
sample_data <- data.frame(read.csv(system.file("extdata", "ELN55049_AllResultsTables_Uncurated.csv", package
processHDE(sample_data)</pre>
```

qualityControl Quality Control table function. Generate a table that collates quality control metrics

# Description

Quality Control table function. Generate a table that collates quality control metrics

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#### Usage

```
qualityControl(
  object,
  massError = NULL,
  intensityOutlier = NULL,
  retentionOutlier = NULL,
  monotonicityStat = NULL,
  mobilityOutlier = NULL,
  chargeCorrelation = NULL,
  replicateCorrelation = NULL,
  replicateOutlier = NULL,
  sequenceCheck = NULL,
  spectraCheck = NULL,
  experiment = NULL,
  timepoints = NULL,
  undeuterated = FALSE
)
```

#### **Arguments**

An object of class Qfeatures, with the data used for the analysis object massError The output of the computeMassError function intensityOutlier The output of the intensityOutliers function retentionOutlier The output of the rTimeOutliers function monotonicityStat The output of the computeMonotoneStats function mobilityOutlier The output of the imTimeOutliers function chargeCorrelation The output of the chargeCorrelationsHdx function replicateCorrelation The output of the replicateCorrelation function replicateOutlier The output of the replicateOutlier function sequenceCheck The output of the compatibleUptake function spectraCheck The output of the spectraSimiarity function The experimental conditions. experiment The timepoints used in the analysis, must include repeat for replicates timepoints A logical indicating whether only the undeuterated data should be exported undeuterated

#### Value

An object of class DataFrame containing a summary of the quality control results.

#### Author(s)

Oliver Crook

replicateCorrelation 17

replicateCorrelation Correlation based checks

#### **Description**

Correlation based checks

# Usage

```
replicateCorrelation(object, experiment, timepoints)
```

## Arguments

object An object of class QFeatures.

experiment A character vector indicating the experimental conditions timepoints A numeric vector indicating the experimental timepoints

#### Value

Returns A list of the same length as the number of experiments indicating outlier from correlation analysis. Outliers are flagged if their deuterium uptake is highly variable.

## Author(s)

Oliver Crook

## **Examples**

```
data("BRD4df_full")
experiment <- c("wt", "iBET")
timepoints <- rep(c(0, 15, 60, 600, 3600, 14000), each = 3)
monoStat <- replicateCorrelation(object = BRD4df_full,
experiment = experiment,
timepoints = timepoints)</pre>
```

replicateOutlier

Correlation based checks

## **Description**

Correlation based checks

# Usage

```
replicateOutlier(object, experiment, timepoints)
```

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## **Arguments**

object An object of class QFeatures.

experiment A character vector indicating the experimental conditions timepoints A numeric vector indicating the experimental timepoints

#### Value

Returns A list of the same length as the number of experiments indicating outlier from correlation analysis. Outliers are flagged if their deuterium uptake is highly variable.

#### Author(s)

Oliver Crook

## **Examples**

```
data("BRD4df_full")
BRD4df_filtered <- isMissingAtRandom(object = BRD4df_full)
BRD4df_full_imputed <- impute(BRD4df_filtered, method = "zero", i = 1)
experiment <- c("wt", "iBET")
timepoints <- rep(c(0, 15, 60, 600, 3600, 14000), each = 3)
monoStat <- replicateOutlier(object = BRD4df_full_imputed,
experiment = experiment,
timepoints = timepoints)</pre>
```

rTimeOutliers

Retention time based analysis

## **Description**

Retention time based analysis

#### Usage

```
rTimeOutliers(
  object,
  leftRT = "leftRT",
  rightRT = "rightRT",
  searchRT = "Search.RT"
)
```

## Arguments

object	An object of class QFeatures
leftRT	A character indicated pattern associated with left boundary of retention time search. Default is "leftRT".
rightRT	A character indicated pattern associated with right boundary of retneton time search. Default is "rightRT".
searchRT	The actual search retention time pattern. Default is "Search.RT"

spectraSimilarity 19

## Value

A list indicating the retention time based outliers.

# Author(s)

Oliver Crook

## **Examples**

```
data("BRD4df_full")
rTimeOutliers(BRD4df_full)
```

 ${\tt spectraSimilarity}$ 

Spectral checking using data from HDsite

# Description

Spectral checking using data from HDsite

# Usage

```
spectraSimilarity(
  peaks,
  object,
  experiment = NULL,
  mzCol = 14,
  startRT = "Start.RT",
  endRT = "End.RT",
  charge = "z",
  incorpD = "X.D.left",
  maxD = "maxD",
  numSpectra = NULL,
  ppm = 300,
  BPPARAM = bpparam()
)
```

# Arguments

peaks a data.frame containing data exported from hdsite	
object a data.frame obtained from HDexaminer data	
experiment A character vector indicating the experimental conditions	
mzCol The column in the peak information indicating the base mz value	
startRT The column indicatng the start of the retention time. Default is "Start.RT"	
endRT The column indicating the end of the retention time. Default is "End.RT"	
charge The column indicating the charge information. Default is "z".	
incorpD The deuterium uptake value column. Default is "X.D.left".	
maxD The maximum allowed deuterium incorporation column. Default is "maxD"	

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numSpectra The number of spectra to analyse. Default is NULL in which all Spectra are

analysed.

ppm The ppm error

BPPARAM Bioconductor parallel options.

# Value

Two list of spectra observed and matching theoretical Spectra

# Author(s)

Oliver Crook

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