

Package ‘ggbio’

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Version 1.58.0

Title Visualization tools for genomic data

Description The ggbio package extends and specializes the grammar of graphics for biological data. The graphics are designed to answer common scientific questions, in particular those often asked of high throughput genomics data. All core Bioconductor data structures are supported, where appropriate. The package supports detailed views of particular genomic regions, as well as genome-wide overviews. Supported overviews include ideograms and grand linear views. High-level plots include sequence fragment length, edge-linked interval to data view, mismatch pileup, and several splicing summaries.

Depends methods, BiocGenerics, ggplot2 (>= 1.0.0)

Imports grid, grDevices, graphics, stats, utils, gridExtra, scales, reshape2, gtable, Hmisc, biovizBase (>= 1.29.2), Biobase, S4Vectors (>= 0.13.13), IRanges (>= 2.11.16), Seqinfo, GenomeInfoDb (>= 1.45.5), GenomicRanges (>= 1.61.1), SummarizedExperiment (>= 1.39.1), Biostrings (>= 2.77.2), Rsamtools (>= 2.25.1), GenomicAlignments (>= 1.45.1), BSgenome (>= 1.77.1), VariantAnnotation (>= 1.55.1), rtracklayer (>= 1.69.1), GenomicFeatures (>= 1.61.4), OrganismDbi, ensemblDb (>= 2.33.1), AnnotationDbi, AnnotationFilter, rlang

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URL <https://lawremi.github.io/ggbio/>

BugReports <https://github.com/lawremi/ggbio/issues>

License Artistic-2.0

LazyLoad Yes

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 layout_karyogram-method.R layout_linear-method.R
 stat_aggregate-method.R stat_coverage-method.R
 stat_identity-method.R stat_mismatch-method.R
 stat_stepping-method.R stat_gene-method.R stat_table-method.R
 stat_bin-method.R stat_slice-method.R stat_reduce-method.R
 coord_genome-method.R autoplot-method.R hack.R
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arrangeGrobByParsingLegend

Arrange grobs by parse their legend.

Description

Arrange grobs and parse their legend, then put it together on the right.

Usage

```
arrangeGrobByParsingLegend(..., nrow = NULL, ncol = NULL,  
  widths = c(4, 1), legend.idx = NULL)
```

Arguments

- ... ggplot graphics.
- nrow number of row for layout.
- ncol number of columns for layout
- widths width ratio for plot group and legend group.
- legend.idx legend index you want to keep.

Value

a

Author(s)

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Examples

```
library(ggplot2)
p1 <- qplot(x = mpg, y= cyl, data = mtcars, color = carb)
p2 <- qplot(x = mpg, y= cyl, data = mtcars, color = wt)
p3 <- qplot(x = mpg, y= cyl, data = mtcars, color = qsec)
p4 <- qplot(x = mpg, y= cyl, data = mtcars, color = gear)
arrangeGrobByParsingLegend(p1, p2, p3, p4)
arrangeGrobByParsingLegend(p1, p2, p3, p4, ncol = 1)
arrangeGrobByParsingLegend(p1, p2, p3, p4, legend.idx = 2)
```

autoplot

Generic autoplot function

Description

autoplot is a generic function to visualize various data object, it tries to give better default graphics and customized choices for each data type, quick and convenient to explore your genomic data compare to low level ggplot method, it is much simpler and easy to produce fairly complicate graphics, though you may lose some flexibility for each layer.

Usage

```
## S4 method for signature 'GRanges'
autoplot(object, ..., chr, xlab, ylab, main, truncate.gaps = FALSE,
          truncate.fun = NULL, ratio = 0.0025, space.skip = 0.1,
          legend = TRUE, geom = NULL, stat = NULL,
          chr.weight = NULL,
          coord = c("default", "genome", "truncate_gaps"),
          layout = c("linear", "karyogram", "circle"))

## S4 method for signature 'GRangesList'
autoplot(object, ..., xlab, ylab, main, indName = "grl_name",
          geom = NULL, stat = NULL, coverage.col = "gray50",
          coverage.fill = coverage.col, group.selfish = FALSE)

## S4 method for signature 'IRanges'
autoplot(object, ..., xlab, ylab, main)

## S4 method for signature 'Seqinfo'
autoplot(object, ideogram = FALSE, ... )

## S4 method for signature 'GAlignments'
autoplot(object, ..., xlab, ylab, main, which,
          geom = NULL, stat = NULL)

## S4 method for signature 'BamFile'
autoplot(object, ..., which, xlab, ylab, main,
          bsgenome, geom = "line", stat = "coverage", method = c("raw",
          "estimate"), coord = c("linear", "genome"),
```

```

        resize.extra = 10, space.skip = 0.1, show.coverage =
        TRUE)

## S4 method for signature 'character'
autoplot(object, ..., xlab, ylab, main, which)

## S4 method for signature 'TxDbOREnsDb'
autoplot(object, which, ..., xlab, ylab, main, truncate.gaps =
        FALSE, truncate.fun = NULL, ratio = 0.0025,
        mode = c("full", "reduce"), geom =
        c("alignment"), stat = c("identity", "reduce"),
        names.expr = "tx_name", label = TRUE)

## S4 method for signature 'BSgenome'
autoplot(object, which, ...,
        xlab, ylab, main, geom = NULL)

## S4 method for signature 'Rle'
autoplot(object, ..., xlab, ylab, main, binwidth, nbin = 30,
        geom = NULL, stat = c("bin", "identity", "slice"),
        type = c("viewSums", "viewMins", "viewMaxs", "viewMeans"))

## S4 method for signature 'RleList'
autoplot(object, ..., xlab, ylab, main, nbin = 30, binwidth,
        facetByRow = TRUE, stat = c("bin", "identity", "slice"),
        geom = NULL, type = c("viewSums", "viewMins", "viewMaxs", "viewMeans"))

## S4 method for signature 'matrix'
autoplot(object, ..., xlab, ylab, main,
        geom = c("tile", "raster"), axis.text.angle = NULL,
        hjust = 0.5, na.value = NULL,
        rownames.label = TRUE, colnames.label = TRUE,
        axis.text.x = TRUE, axis.text.y = TRUE)

## S4 method for signature 'ExpressionSet'
autoplot(object, ..., type = c("heatmap", "none",
        "scatterplot.matrix", "pcp", "MA", "boxplot",
        "mean-sd"), test.method =
        "t", rotate = FALSE, pheno.plot = FALSE, main_to_pheno
        = 4.5, padding = 0.2)

## S4 method for signature 'RangedSummarizedExperiment'
autoplot(object, ..., type = c("heatmap", "link", "pcp", "boxplot", "scatterplot.matrix"), pheno.pl
        main_to_pheno = 4.5, padding = 0.2, assay.id = 1)

## S4 method for signature 'VCF'

```

```

autoplot(object, ...,
          xlab, ylab, main,
          assay.id,
          type = c("default", "geno", "info", "fixed"),
          full.string = FALSE,
          ref.show = TRUE,
          genome.axis = TRUE,
          transpose = TRUE)

## S4 method for signature 'OrganismDb'
autoplot(object, which, ...,
          xlab, ylab, main,
          truncate.gaps = FALSE,
          truncate.fun = NULL,
          ratio = 0.0025,
          geom = c("alignment"),
          stat = c("identity", "reduce"),
          columns = c("TXNAME", "SYMBOL", "TXID", "GENEID"),
          names.expr = "SYMBOL",
          label = TRUE,
          label.color = "gray40")

## S4 method for signature 'VRanges'
autoplot(object, ..., which = NULL,
          arrow = TRUE, indel.col = "gray30",
          geom = NULL,
          xlab, ylab, main)

## S4 method for signature 'TabixFile'
autoplot(object, which, ...)

```

Arguments

<code>object</code>	object to be plot.
<code>columns</code>	columns passed to method works for TxDb, EnsDb and OrganismDb.
<code>label.color</code>	when label turned on for gene model, this parameter controls label color.
<code>arrow</code>	arrow passed to <code>genome_alignment</code> function to control intron arrow attributes.
<code>indel.col</code>	indel colors.
<code>ideogram</code>	Whether to call <code>plotIdeogram</code> or not, default is FALSE, if TRUE, <code>layout_karyogram</code> will be called.
<code>transpose</code>	logical value, default TRUE, always make features from VCF as x, so we can use it to map to genomic position.
<code>axis.text.angle</code>	axis text angle.
<code>axis.text.x</code>	logical value indicates whether to show x axis and labels or not.
<code>axis.text.y</code>	logical value indicates whether to show y axis and labels or not.
<code>hjust</code>	horizontal just for axis text.
<code>rownames.label</code>	logical value indicates whether to show rownames of matrix as y label or not.
<code>colnames.label</code>	logical value indicates whether to show colnames of matrix as y label or not.

na.value	color for NA value.
rotate	
pheno.plot	show pheno plot or not.
main_to_pheno	main matrix plot width to pheno plot width ratio.
padding	padding between plots.
assay.id	index for assay you are going to use.
geom	Geom to use (Single character for now). Please see section Geometry for details.
truncate.gaps	logical value indicate to truncate gaps or not.
truncate.fun	shrinkage function. Please see shrinkagefun in package biovizBase.
ratio	used in maxGap.
mode	Display mode for genomic features.
space.skip	space ratio between chromosome spaces in coordate genome.
coord	Coodinate system.
chr.weight	numeric vectors which sum to <1, the names of vectors has to be matched with seqnames in seqinfo, and you can only specify part of the seqnames, other lengths of chromosomes will be assined proportionally to their seqlengths, for example, you could specify chr1 to be 0.5, so the chr1 will take half of the space and other chromosomes squeezed to take left of the space.
legend	A logical value indicates whether to show legend or not. Default is TRUE
which	A GRanges object to subset the result, usually passed to the ScanBamParam function. For autoplot, EnsDb, which can in addition also be an object extending AnnotationFilter , an AnnotationFilterList combining such objects or a formula representing a filter expression. See examples below or documentation of AnnotationFilter for more details.
show.coverage	A logical value indicates whether to show coverage or not. This is used for geom "mismatch.summary".
resize.extra	A numeric value used to add buffer to intervals to compute stepping levels on.
bsgenome	A BSgenome object. Only need for geom "mismatch.summary".
xlab	x label.
ylab	y label.
label	logic value, default TRUE. To show label by the side of features.
facetByRow	A logical value, default is TRUE ,facet RleList by row. If FALSE, facet by column.
type	For Rle/RleList, "raw" plot everything, so be careful, that would be pretty slow if you have too much data. For "viewMins", "viewMaxs", "viewMeans", "viewSums", require extra arguments to slice the object. so users need to at least provide lower, more details and control please refer the the manual of slice function in IRanges. For "viewMins", "viewMaxs", we use viewWhichMin and viewWhichMax to get x scale, for "viewMeans", "viewSums", we use window midpoint as x. For ExpresionSet, plotting types.
layout	Layout including linear, circular and karyogram. for GenomicRangesList, it only supports circular layout.
method	method used for parsing coverage from bam files. 'estimate' use fast esitimated method and 'raw' use relatively slow parsing method.

<code>test.method</code>	test method
<code>...</code>	Extra parameters. Usually are those parameters used in autoplot to control aesthetics or geometries.
<code>main</code>	title.
<code>stat</code>	statistical transformation.
<code>indName</code>	When coerce <code>GRangesList</code> to <code>GRanges</code> , names created for each group.
<code>coverage.col</code>	coverage stroke color.
<code>coverage.fill</code>	coverage fill color.
<code>group.selfish</code>	Passed to <code>addStepping</code> , control whether to show each group as unique level or not. If set to <code>FALSE</code> , if two groups are not overlapped with each other, they will probably be layout in the same level to save space.
<code>names.expr</code>	names expression used for creating labels. For <code>EnsDb</code> objects either <code>"tx_id"</code> , <code>"gene_name"</code> or <code>"gene_id"</code> .
<code>binwidth</code>	width of the bins.
<code>nbin</code>	number of bins.
<code>genome.axis</code>	logical value, if <code>TRUE</code> , whenever possible, try to parse genomic position for each column(e.g. <code>RangedSummarizedExperiment</code>), show column as exactly the genomic position instead of showing them side by side and indexed from 1.
<code>full.string</code>	logical value. If <code>TRUE</code> , show full string of indels in plot for VCF.
<code>ref.show</code>	logical value. If <code>TRUE</code> , show REF in VCF at bottom track.
<code>chr</code>	characters indicates the seqnames to be subseted.

Value

A `ggplot` object, so you can use common features from `ggplot2` package to manipulate the plot.

Introduction

`autoplot` is redefined as generic `s4` method inside this package, user could use `autoplot` in the way they are familiar with, and we are also setting limitation inside this package, like

- scales X scales is always genomic coordinates in most cases, x could be specified as start/end/midpoint when it's special geoms for interval data like point/line
- colors Try to use default color scheme defined in `biovizBase` package as possible as it can

Geometry

We have developed new geom for different objects, some of them may require extra parameters you need to provide. Some of the geom are more like `geom + stat` in `ggplot2` package. e.g. `"coverage.line"` and `"coverage.polygon"`. We simply combine them together, but in the future, we plan to make it more general.

This package is designed for only biological data, especially genomic data if users want to explore the data in a more flexible way, you could simply coerce the `GRanges` to a `data.frame`, then just use formal `autoplot` function in `ggplot2`, or `autoplot` generic for `data.frame`.

Some objects share the same geom so we introduce all the geom together in this section

Showing all the intervals as stepped rectangle, colored by strand automatically.

For `TxDb` or `EnsDb` objects, showing full model.

- segment** Showing all the intervals as stepped segments, colored by strand automatically.
 For object BSgenome, show nucleotides as colored segment.
 For Rle/RleList, show histogram-like segments.
- line** Showing interval as line, the interval data could also be just single position when start = end, x is one of start/end/midpoint, y value is unquoted name in elementMetadata column names. y value is required.
- point** Showing interval as point, the interval data could also be just single position when start = end, x is one of start/end/midpoint, y value is unquoted name in elementMetadata column names. y value is required.
 For object BSgenome, show nucleotides as colored point.
- coverage.line** Coverage showing as lines for interval data.
- coverage.polygon** Coverage showing as polygon for interval data.
- splice** Splicing summary. The size and width of the line and rectangle should represent the counts in each model. Need to provide model.
- single** For TxDb or EnsDb objects, showing single(reduced) model only.
- tx** For TxDb or EnsDb objects, showing transcripts isoforms.
- mismatch.summary** Showing color coded mismatched stacked bar to indicate the proportion of mismatching at each position, the reference is set to gray.
- text** For object BSgenome, show nucleotides as colored text.
- rectangle** For object BSgenome, show nucleotides as colored rectangle.

Faceting

Faceting in ggbio package is a little different from ggplot2 in several ways

- The faceted column could only be seqnames or regions on the genome. So we limited the formula passing to facet argument, e.g something ~ seqnames, is accepted formula, you can change "something" to variable name in the elementMetadata. But you can not change the second part.
- Sometime, we need to view different regions, so we also have a facet_gr argument which accept a GRanges. If this is provided, it will override the default seqnames and use provided region to facet the graphics, this might be useful for different gene centric views.

Author(s)

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Examples

```
set.seed(1)
N <- 1000
library(GenomicRanges)
gr <- GRanges(seqnames =
  sample(c("chr1", "chr2", "chr3"),
    size = N, replace = TRUE),
  IRanges(
    start = sample(1:300, size = N, replace = TRUE),
    width = sample(70:75, size = N, replace = TRUE)),
  strand = sample(c("+", "-", "*"), size = N,
    replace = TRUE),
  value = rnorm(N, 10, 3), score = rnorm(N, 100, 30),
```

```

sample = sample(c("Normal", "Tumor"),
  size = N, replace = TRUE),
pair = sample(letters, size = N,
  replace = TRUE))

idx <- sample(1:length(gr), size = 50)

#####
### code chunk number 3: default
#####
autoplot(gr[idx])

#####
### code chunk number 4: bar-default-pre
#####
set.seed(123)
gr.b <- GRanges(seqnames = "chr1", IRanges(start = seq(1, 100, by = 10),
  width = sample(4:9, size = 10, replace = TRUE)),
  score = rnorm(10, 10, 3), value = runif(10, 1, 100))
gr.b2 <- GRanges(seqnames = "chr2", IRanges(start = seq(1, 100, by = 10),
  width = sample(4:9, size = 10, replace = TRUE)),
  score = rnorm(10, 10, 3), value = runif(10, 1, 100))
gr.b <- c(gr.b, gr.b2)
head(gr.b)

#####
### code chunk number 5: bar-default
#####
p1 <- autoplot(gr.b, geom = "bar")
## use value to fill the bar
p2 <- autoplot(gr.b, geom = "bar", aes(fill = value))
tracks(default = p1, fill = p2)

#####
### code chunk number 6: autoplot.Rnw:236-237
#####
autoplot(gr[idx], geom = "arch", aes(color = value), facets = sample ~ seqnames)

#####
### code chunk number 7: gr-group
#####
gra <- GRanges("chr1", IRanges(c(1,7,20), end = c(4,9,30)), group = c("a", "a", "b"))
## if you desn't specify group, then group based on stepping levels, and gaps are computed without
## considering extra group method
p1 <- autoplot(gra, aes(fill = group), geom = "alignment")
## when use group method, gaps only computed for grouped intervals.
## default is group.selfish = TRUE, each group keep one row.
## in this way, group labels could be shown as y axis.
p2 <- autoplot(gra, aes(fill = group, group = group), geom = "alignment")
## group.selfish = FALSE, save space
p3 <- autoplot(gra, aes(fill = group, group = group), geom = "alignment", group.selfish = FALSE)
tracks('non-group' = p1, 'group.selfish = TRUE' = p2, 'group.selfish = FALSE' = p3)

```

```
#####
### code chunk number 8: gr-facet-strand
#####
autoplot(gr, stat = "coverage", geom = "area",
         facets = strand ~ seqnames, aes(fill = strand))

#####
### code chunk number 9: gr-autoplot-circle
#####
autoplot(gr[idx], layout = 'circle')

#####
### code chunk number 10: gr-circle
#####
seqlengths(gr) <- c(400, 500, 700)
values(gr)$to.gr <- gr[sample(1:length(gr), size = length(gr))]
idx <- sample(1:length(gr), size = 50)
gr <- gr[idx]
ggplot() + layout_circle(gr, geom = "ideo", fill = "gray70", radius = 7, trackWidth = 3) +
  layout_circle(gr, geom = "bar", radius = 10, trackWidth = 4,
               aes(fill = score, y = score)) +
  layout_circle(gr, geom = "point", color = "red", radius = 14,
               trackWidth = 3, grid = TRUE, aes(y = score)) +
  layout_circle(gr, geom = "link", linked.to = "to.gr", radius = 6, trackWidth = 1)

#####
### code chunk number 11: seqinfo-src
#####
data(hg19Ideogram, package = "biovizBase")
sq <- seqinfo(hg19Ideogram)
sq

#####
### code chunk number 12: seqinfo
#####
autoplot(sq[paste0("chr", c(1:22, "X"))])

#####
### code chunk number 13: ir-load
#####
set.seed(1)
N <- 100
ir <- IRanges(start = sample(1:300, size = N, replace = TRUE),
              width = sample(70:75, size = N, replace = TRUE))
## add meta data
df <- DataFrame(value = rnorm(N, 10, 3), score = rnorm(N, 100, 30),
                sample = sample(c("Normal", "Tumor"),
                               size = N, replace = TRUE),
                pair = sample(letters, size = N,
                              replace = TRUE))
```

```
values(ir) <- df
ir
```

```
#####
### code chunk number 14: ir-exp
#####
p1 <- autoplot(ir)
p2 <- autoplot(ir, aes(fill = pair)) + theme(legend.position = "none")
p3 <- autoplot(ir, stat = "coverage", geom = "line", facets = sample ~. )
p4 <- autoplot(ir, stat = "reduce")
tracks(p1, p2, p3, p4)
```

```
#####
### code chunk number 15: grl-simul
#####
set.seed(1)
N <- 100
## =====
## simulated GRanges
## =====
gr <- GRanges(seqnames =
  sample(c("chr1", "chr2", "chr3"),
    size = N, replace = TRUE),
  IRanges(
    start = sample(1:300, size = N, replace = TRUE),
    width = sample(30:40, size = N, replace = TRUE),
    strand = sample(c("+", "-", "*"), size = N,
      replace = TRUE),
    value = rnorm(N, 10, 3), score = rnorm(N, 100, 30),
    sample = sample(c("Normal", "Tumor"),
      size = N, replace = TRUE),
    pair = sample(letters, size = N,
      replace = TRUE))

grl <- split(gr, values(gr)$pair)
```

```
#####
### code chunk number 16: grl-exp
#####
## default gap.geom is 'chevron'
p1 <- autoplot(grl, group.selfish = TRUE)
p2 <- autoplot(grl, group.selfish = TRUE, main.geom = "arrowrect", gap.geom = "segment")
tracks(p1, p2)
```

```
#####
### code chunk number 17: grl-name
#####
autoplot(grl, aes(fill = ..grl_name..))
## equal to
## autoplot(grl, aes(fill = grl_name))
```

```
#####
### code chunk number 18: rle-simul
#####
library(IRanges)
set.seed(1)
lambda <- c(rep(0.001, 4500), seq(0.001, 10, length = 500),
            seq(10, 0.001, length = 500))

## @knitr create
xVector <- rpois(1e4, lambda)
xRle <- Rle(xVector)
xRle

#####
### code chunk number 19: rle-bin
#####
p1 <- autoplot(xRle)
p2 <- autoplot(xRle, nbin = 80)
p3 <- autoplot(xRle, geom = "heatmap", nbin = 200)
tracks('nbin = 30' = p1, "nbin = 80" = p2, "nbin = 200(heatmap)" = p3)

#####
### code chunk number 20: rle-id
#####
p1 <- autoplot(xRle, stat = "identity")
p2 <- autoplot(xRle, stat = "identity", geom = "point", color = "red")
tracks('line' = p1, "point" = p2)

#####
### code chunk number 21: rle-slice
#####
p1 <- autoplot(xRle, type = "viewMaxs", stat = "slice", lower = 5)
p2 <- autoplot(xRle, type = "viewMaxs", stat = "slice", lower = 5, geom = "heatmap")
tracks('bar' = p1, "heatmap" = p2)

#####
### code chunk number 22: rlel-simul
#####
xRleList <- RleList(xRle, 2L * xRle)
xRleList

#####
### code chunk number 23: rlel-bin
#####
p1 <- autoplot(xRleList)
p2 <- autoplot(xRleList, nbin = 80)
p3 <- autoplot(xRleList, geom = "heatmap", nbin = 200)
tracks('nbin = 30' = p1, "nbin = 80" = p2, "nbin = 200(heatmap)" = p3)

#####
### code chunk number 24: rlel-id
```

```
#####
p1 <- autoplot(xRleList, stat = "identity")
p2 <- autoplot(xRleList, stat = "identity", geom = "point", color = "red")
tracks('line' = p1, "point" = p2)

#####
### code chunk number 25: rlel-slice
#####
p1 <- autoplot(xRleList, type = "viewMaxs", stat = "slice", lower = 5)
p2 <- autoplot(xRleList, type = "viewMaxs", stat = "slice", lower = 5, geom = "heatmap")
tracks('bar' = p1, "heatmap" = p2)

#####
### code chunk number 26: txdb
#####
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
data(genesymbol, package = "biovizBase")
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene

#####
### code chunk number 27: txdb-visual
#####
p1 <- autoplot(txdb, which = genesymbol["ALDOA"], names.expr = "tx_name:::gene_id")
p2 <- autoplot(txdb, which = genesymbol["ALDOA"], stat = "reduce", color = "brown",
               fill = "brown")
tracks(full = p1, reduce = p2, heights = c(5, 1)) + ylab("")

#####
### EnsDb
#####
## Fetching gene models from an EnsDb object.
library(EnsDb.Hsapiens.v75)
ensdb <- EnsDb.Hsapiens.v75
## We use a GenenameFilter to specifically retrieve all transcripts for that gene.
p1 <- autoplot(ensdb, which = GenenameFilter("ALDOA"), names.expr = "gene_name")
## Instead of providing the GenenameFilter, we can also use filter expressions
p2 <- autoplot(ensdb, which = ~ genename == "ALDOA", stat = "reduce",
               color = "brown", fill = "brown")
tracks(full = p1, reduce = p2, heights = c(5, 1)) + ylab("")

## Alternatively, we can specify a GRangesFilter and display all genes
## that are (partially) overlapping with that genomic region:
gr <- GRanges(seqnames=16, IRanges(30768000, 30770000), strand="+")
autoplot(ensdb, GRangesFilter(gr, "any"), names.expr="gene_name")
## Just submitting the GRanges object also works.
autoplot(ensdb, gr, names.expr="gene_name")

## Or genes encoded on both strands.
gr <- GRanges(seqnames = 16, IRanges(30768000, 30770000), strand = "*")
autoplot(ensdb, GRangesFilter(gr), names.expr="gene_name")

## Also, we can specify directly the gene ids and plot all transcripts of these
## genes (not only those overlapping with the region)
```

```

autoplot(ensdb, GeneIdFilter(c("ENSG00000196118", "ENSG00000156873")))

#####
### code chunk number 28: ga-load
#####
library(GenomicAlignments)
data("genesymbol", package = "biovizBase")
bamfile <- system.file("extdata", "SRR027894subRBM17.bam",
                        package="biovizBase")
library(GenomeInfoDb) # for keepStandardChromosomes()
which <- keepStandardChromosomes(genesymbol["RBM17"])
## need to set use.names = TRUE
ga <- readGAlignments(bamfile,
                      param = ScanBamParam(which = which),
                      use.names = TRUE)

#####
### code chunk number 29: ga-exp
#####
p1 <- autoplot(ga)
p2 <- autoplot(ga, geom = "rect")
p3 <- autoplot(ga, geom = "line", stat = "coverage")
tracks(default = p1, rect = p2, coverage = p3)

#####
### code chunk number 30: bf-load (eval = FALSE)
#####
## library(Rsamtools)
## bamfile <- "../wgEncodeCaltechRnaSeqK562R1x75dAlignsRep1V2.bam"
## bf <- BamFile(bamfile)

#####
### code chunk number 31: bf-est-cov (eval = FALSE)
#####
## autoplot(bamfile)
## autoplot(bamfile, which = c("chr1", "chr2"))
## autoplot(bf)
## autoplot(bf, which = c("chr1", "chr2"))
##
## data(genesymbol, package = "biovizBase")
## autoplot(bamfile, method = "raw", which = genesymbol["ALDOA"])
##
## library(BSgenome.Hsapiens.UCSC.hg19)
## autoplot(bf, stat = "mismatch", which = genesymbol["ALDOA"], bsgenome = Hsapiens)

#####
### code chunk number 32: char-bam (eval = FALSE)
#####
## bamfile <- "../wgEncodeCaltechRnaSeqK562R1x75dAlignsRep1V2.bam"
## autoplot(bamfile)

#####

```

```

### code chunk number 33: char-gr
#####
library(rtracklayer)
test_path <- system.file("tests", package = "rtracklayer")
test_bed <- file.path(test_path, "test.bed")
autoplot(test_bed, aes(fill = name))

#####
### matrix
#####
volcano <- volcano[20:70, 20:60] - 150
autoplot(volcano)
autoplot(volcano, xlab = "xlab", main = "main", ylab = "ylab")
## special scale theme for 0-centered values
autoplot(volcano, geom = "raster")+scale_fill_fold_change()

## when a matrix has colnames and rownames label them by default
colnames(volcano) <- sort(sample(1:300, size = ncol(volcano), replace = FALSE))
autoplot(volcano)+scale_fill_fold_change()

rownames(volcano) <- letters[sample(1:24, size = nrow(volcano), replace = TRUE)]
autoplot(volcano)

## even with row/col names, you could also disable it and just use numeric index
autoplot(volcano, colnames.label = FALSE)
autoplot(volcano, rownames.label = FALSE, colnames.label = FALSE)

## don't want the axis has label??
autoplot(volcano, axis.text.x = FALSE)
autoplot(volcano, axis.text.y = FALSE)

# or totally remove axis
colnames(volcano) <- lapply(letters[sample(1:24, size = ncol(volcano),
replace = TRUE)],
function(x){
  paste(rep(x, 7), collapse = "")
})

## Oops, overlapped
autoplot(volcano)
## tweak with it.
autoplot(volcano, axis.text.angle = -45, hjust = 0)

## when character is the value
x <- sample(c(letters[1:3], NA), size = 100, replace = TRUE)
mx <- matrix(x, nrow = 5)
autoplot(mx)
## tile gives you a white margin
rownames(mx) <- LETTERS[1:5]
autoplot(mx, color = "white")
colnames(mx) <- LETTERS[1:20]
autoplot(mx, color = "white")
autoplot(mx, color = "white", size = 2)
## weird in aes(), though works
## default tile is flexible

```



```

autoplot(mx, aes(width = 0.6, height = 0.6))
autoplot(mx, aes(width = 0.6, height = 0.6), na.value = "white")
autoplot(mx, aes(width = 0.6, height = 0.6)) + theme_clear()

#####
### Views
#####
lambda <- c(rep(0.001, 4500), seq(0.001, 10, length = 500),
            seq(10, 0.001, length = 500))
xVector <- dnorm(1:5e3, mean = 1e3, sd = 200)
xRle <- Rle(xVector)
v1 <- Views(xRle, start = sample(.4e3:.6e3, size = 50, replace = FALSE), width = 1000)
autoplot(v1)
names(v1) <- letters[sample(1:24, size = length(v1), replace = TRUE)]
autoplot(v1)
autoplot(v1, geom = "tile", aes(width = 0.5, height = 0.5))
autoplot(v1, geom = "line")
autoplot(v1, geom = "line", aes(color = row)) + theme(legend.position = "none")
autoplot(v1, geom = "line", facets = NULL)
autoplot(v1, geom = "line", facets = NULL, alpha = 0.1)

#####
### ExpressionSet
#####
library(Biobase)
data(sample.ExpressionSet)
sample.ExpressionSet
set.seed(1)
## select 50 features
idx <- sample(seq_len(dim(sample.ExpressionSet)[1]), size = 50)
eset <- sample.ExpressionSet[idx,]
eset
autoplot(as.matrix(pData(eset)))

## default heatmap
p1 <- autoplot(eset)
p2 <- p1 + scale_fill_fold_change()
p2
autoplot(eset)
autoplot(eset, geom = "tile", color = "white", size = 2)
autoplot(eset, geom = "tile", aes(width = 0.6, height = 0.6))

autoplot(eset, pheno.plot = TRUE)
idx <- order(pData(eset)[,1])
eset2 <- eset[,idx]
autoplot(eset2, pheno.plot = TRUE)

## parallel coordainte plot
autoplot(eset, type = "pcp")

## boxplot
autoplot(eset, type = "boxplot")

## scatterplot.matrix
## slow, be carefull

```

```

##autoplot(eset[, 1:7], type = "scatterplot.matrix")

## mean-sd
autoplot(eset, type = "mean-sd")

#####
### RangedSummarizedExperiment
#####
library(SummarizedExperiment)
nrows <- 200; ncols <- 6
counts <- matrix(runif(nrows * ncols, 1, 1e4), nrows)
counts2 <- matrix(runif(nrows * ncols, 1, 1e4), nrows)
rowRanges <- GRanges(rep(c("chr1", "chr2"), c(50, 150)),
                      IRanges(floor(runif(200, 1e5, 1e6)), width=100),
                      strand=sample(c("+", "-"), 200, TRUE))
colData <- DataFrame(Treatment=rep(c("ChIP", "Input"), 3),
                    row.names=LETTERS[1:6])
sset <- SummarizedExperiment(assays=SimpleList(counts=counts,
                                              counts2 = counts2),
                           rowRanges=rowRanges, colData=colData)
autoplot(sset) + scale_fill_fold_change()
autoplot(sset, pheno.plot = TRUE)

#####
### pcp
#####
autoplot(sset, type = "pcp")

#####
### boxplot
#####
autoplot(sset, type = "boxplot")

#####
### scatterplot matrix
#####
##autoplot(sset, type = "scatterplot.matrix")

#####
### vcf
#####

## Not run:
library(VariantAnnotation)
vcffile <- system.file("extdata", "chr22.vcf.gz", package="VariantAnnotation")
vcf <- readVcf(vcffile, "hg19")
## default use type 'geno'
## default use genome position
autoplot(vcf)
## or disable it

```

```

autoplot(vcf, genome.axis = FALSE)
## not transpose
autoplot(vcf, genome.axis = FALSE, transpose = FALSE, rownames.label = FALSE)
autoplot(vcf)
## use
autoplot(vcf, assay.id = "DS")
## equivalent to
autoplot(vcf, assay.id = 2)
## doesn't work when assay.id cannot find
autoplot(vcf, assay.id = "NO")
## use AF or first
autoplot(vcf, type = "info")
## geom bar
autoplot(vcf, type = "info", aes(y = THETA))
autoplot(vcf, type = "info", aes(y = THETA, fill = VT, color = VT))
autoplot(vcf, type = "fixed")
autoplot(vcf, type = "fixed", size = 10) + xlim(c(50310860, 50310890)) + ylim(0.75, 1.25)

p1 <- autoplot(vcf, type = "fixed") + xlim(50310860, 50310890)
p2 <- autoplot(vcf, type = "fixed", full.string = TRUE) + xlim(50310860, 50310890)
tracks("full.string = FALSE" = p1, "full.string = TRUE" = p2)+
  scale_y_continuous(breaks = NULL, limits = c(0, 3))
p3 <- autoplot(vcf, type = "fixed", ref.show = FALSE) + xlim(50310860, 50310890) +
  scale_y_continuous(breaks = NULL, limits = c(0, 2))
p3

## End(Not run)

#####
### code chunk number 56: bs-v
#####
library(BSgenome.Hsapiens.UCSC.hg19)
data(genesymbol, package = "biovizBase")
p1 <- autoplot(Hsapiens, which = resize(genesymbol["ALDOA"], width = 50))
p2 <- autoplot(Hsapiens, which = resize(genesymbol["ALDOA"], width = 50), geom = "rect")
tracks(text = p1, rect = p2)

#####
### code chunk number 57: sessionInfo
#####
sessionInfo()

```

Description

Show interval data as alignment.

Usage

```
## S4 method for signature 'GRanges'
```

```

geom_alignment(data, ..., xlab, ylab, main, facets = NULL, stat =
  c("stepping", "identity"), range.geom = c("rect",
  "arrowrect"), gap.geom = c("chevron", "arrow",
  "segment"), rect.height = NULL, group.selfish = TRUE,
  label = TRUE)

## S4 method for signature 'TxDbOREnsDb'
geom_alignment(data, ..., which, columns = c("tx_id", "tx_name",
  "gene_id"), names.expr = "tx_name", facets = NULL,
  truncate.gaps = FALSE, truncate.fun = NULL, ratio =
  0.0025)

## S4 method for signature 'GRangesList'
geom_alignment(data, ..., which = NULL,
  cds.rect.h = 0.25,
  exon.rect.h = cds.rect.h,
  utr.rect.h = cds.rect.h/2,
  xlab, ylab, main,
  facets = NULL, geom = "alignment",
  stat = c("identity", "reduce"),
  range.geom = "rect",
  gap.geom = "arrow",
  utr.geom = "rect",
  names.expr = NULL,
  label = TRUE,
  label.color = "gray40",
  label.size = 3,
  arrow.rate = 0.015,
  length = unit(0.1, "cm"))

## S4 method for signature 'OrganismDb'
geom_alignment(data, ..., which,
  columns = c("TXNAME", "SYMBOL", "TXID", "GENEID"),
  names.expr = "SYMBOL",
  facets = NULL,
  truncate.gaps = FALSE,
  truncate.fun = NULL, ratio = 0.0025
)

```

Arguments

<code>data</code>	A GRanges, data.frame, TxDb or EnsDb object.
<code>...</code>	Extra parameters such as <code>aes()</code> passed.
<code>which</code>	GRanges object to subset the TxDb or EnsDb object. For EnsDb: can also be a single object extending AnnotationFilter , an AnnotationFilterList combining such objects or a filter expression in form of a formula.
<code>cds.rect.h</code>	cds heights.
<code>exon.rect.h</code>	exon heights.
<code>utr.rect.h</code>	utr heights.
<code>label.color</code>	label color.
<code>label.size</code>	label size.

arrow.rate	arrow rate.
length	arrow length.
columns	columns to get from object.
xlab	Label for x
ylab	Label for y
main	Title for plot.
facets	Faceting formula to use.
stat	For GRanges : Character vector specifying statistics to use. "stepping" with randomly assigned stepping levels as y variable. "identity" allow users to specify y value in aes. For TxDb : default "identity" give full gene model and "reduce" for reduced model.
gap.geom	Geom for 'gap' computed from the data you passed based on the group information.
rect.height	Half height of the arrow body.
group.selfish	Passed to addStepping, control whether to show each group as unique level or not. If set to FALSE, if two groups are not overlapped with each other, they will probably be layout in the same level to save space.
truncate.gaps	logical value indicate to truncate gaps or not.
truncate.fun	shrinkage function. Please see shrinkagefun in package biovizBase.
ratio	used in maxGap.
geom	geometric object. only support "gene" now.
range.geom	geom for main intervals or exons.
utr.geom	geom for utr region.
names.expr	expression for showing y label.
label	logical value. Whether to label the intervals with names specified by argument names.expr.

Value

A 'Layer'.

Author(s)

Tengfei Yin

Examples

```
set.seed(1)
N <- 100
require(GenomicRanges)
## =====
## simulated GRanges
## =====
gr <- GRanges(seqnames =
  sample(c("chr1", "chr2", "chr3"),
    size = N, replace = TRUE),
  IRanges(
```

```

        start = sample(1:300, size = N, replace = TRUE),
        width = sample(70:75, size = N, replace = TRUE)),
strand = sample(c("+", "-", "*"), size = N,
  replace = TRUE),
value = rnorm(N, 10, 3), score = rnorm(N, 100, 30),
sample = sample(c("Normal", "Tumor"),
  size = N, replace = TRUE),
pair = sample(letters, size = N,
  replace = TRUE))

## =====
## default
## =====
ggplot(gr) + geom_alignment()
## or
ggplot() + geom_alignment(gr)

## =====
## facetting and aesthetics
## =====
ggplot(gr) + geom_alignment(facets = sample ~ seqnames, mapping = aes(color = strand, fill = strand))

## =====
## stat:stepping
## =====
ggplot(gr) + geom_alignment(stat = "stepping", mapping = aes(group = pair))

## =====
## group.selfish controls when
## =====
ggplot(gr) + geom_alignment(stat = "stepping", mapping = aes(group = pair), group.selfish = FALSE)

## =====
## main/gap geom
## =====
ggplot(gr) + geom_alignment(range.geom = "arrowrect", gap.geom = "chevron")

## =====
## For TxDb
## =====
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
data(genesymbol, package = "biovizBase")
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
## made a track comparing full/reduce stat.
ggbio() + geom_alignment(data = txdb, which = genesymbol["RBM17"])
p1 <- ggplot(txdb) + geom_alignment(which = genesymbol["RBM17"])
p1
p2 <- ggplot(txdb) + geom_alignment(which = genesymbol["RBM17"], stat = "reduce")
tracks(full = p1, reduce = p2, heights = c(3, 1))
tracks(full = p1, reduce = p2, heights = c(3, 1)) + theme_tracks_sunset()
tracks(full = p1, reduce = p2, heights = c(3, 1)) +
  theme_tracks_sunset(axis.line.color = NA)
## change y labels
ggplot(txdb) + geom_alignment(which = genesymbol["RBM17"], names.expr = "tx_id:::gene_id")

```

geom_arch

*Arch geoms for GRanges object***Description**

Show interval data as arches.

Usage

```
## S4 method for signature 'data.frame'
geom_arch(data, ..., n = 25, max.height = 10)

## S4 method for signature 'GRanges'
geom_arch(data, ..., xlab, ylab, main, facets = NULL,
          rect.height = 0, n = 25, max.height = 10)
```

Arguments

data	A GRanges or data.frame object.
...	Extra parameters passed to autoplot function, aes mapping support height, x, xend. • xstart of the arches • xendend of the arches • heightheight of arches
xlab	Label for x
ylab	Label for y
main	Title for plot.
n	Integer values at which interpolation takes place to create 'n' equally spaced points spanning the interval [<code>'min(x)'</code> , <code>'max(x)'</code>].
facets	Faceting formula to use.
rect.height	When data is GRanges, this padding the arches from original y value to allow users putting arches 'around' the interval rectangles.
max.height	Max height of all arches.

Details

To draw a interval data as arches, we need to provide a special geom for this purpose. Arches is popular in gene viewer or genomoe browser, when they try to show isoforms or gene model. `geom_arch`, just like any other `geom_*` function in `ggplot2`, you can pass `aes()` to it to map variable to height of arches.

Value

A 'Layer'.

Author(s)

Tengfei Yin

Examples

```

set.seed(1)
N <- 100
library(GenomicRanges)

## =====
##  simulated GRanges
## =====
gr <- GRanges(seqnames =
  sample(c("chr1", "chr2", "chr3"),
    size = N, replace = TRUE),
  IRanges(
    start = sample(1:300, size = N, replace = TRUE),
    width = sample(70:75, size = N, replace = TRUE)),
  strand = sample(c("+", "-", "*"), size = N,
    replace = TRUE),
  value = rnorm(N, 10, 3), score = rnorm(N, 100, 30),
  sample = sample(c("Normal", "Tumor"),
    size = N, replace = TRUE),
  pair = sample(letters, size = N,
    replace = TRUE))

## =====
##  default
## =====
ggplot(gr) + geom_arch()
# or
ggplot() + geom_arch(gr)

## =====
##  facetting and aesthetics
## =====
ggplot(gr) + geom_arch(mapping = aes(color = value, height = value, size = value),
  alpha = 0.2, facets = sample ~ seqnames)

```

geom_arrow

Arrow geoms for GRanges object

Description

Show interval data as arrows.

Usage

```

## S4 method for signature 'GRanges'
geom_arrow(data, ..., xlab, ylab, main,
  angle = 30, length = unit(0.12, "cm"), type = "open",
  stat = c("stepping", "identity"), facets = NULL,
  arrow.rate = 0.03, group.selfish = TRUE)

```


Arguments

<code>data</code>	A GRanges object.
<code>...</code>	Extra parameters such as <code>aes()</code> passed.
<code>xlab</code>	Label for x
<code>ylab</code>	Label for y
<code>main</code>	Title for plot.
<code>angle</code>	The angle of the arrow head in degrees (smaller numbers produce narrower, pointier arrows). Essentially describes the width of the arrow head.
<code>length</code>	A unit specifying the length of the arrow head (from tip to base).
<code>type</code>	One of "open" or "closed" indicating whether the arrow head should be a closed triangle.
<code>stat</code>	Character vector specifying statistics to use. "stepping" with randomly assigned stepping levels as y variable. "identity" allow users to specify y value in aes.
<code>facets</code>	Faceting formula to use.
<code>arrow.rate</code>	Arrow density of the arrow body.
<code>group.selfish</code>	Passed to <code>addStepping</code> , control whether to show each group as unique level or not. If set to FALSE, if two groups are not overlapped with each other, they will probably be layout in the same level to save space.

Value

A 'Layer'.

Author(s)

Tengfei Yin

Examples

```

set.seed(1)
N <- 100
require(GenomicRanges)
## =====
## simulated GRanges
## =====
gr <- GRanges(seqnames =
  sample(c("chr1", "chr2", "chr3"),
    size = N, replace = TRUE),
  IRanges(
    start = sample(1:300, size = N, replace = TRUE),
    width = sample(70:75, size = N, replace = TRUE)),
  strand = sample(c("+", "-", "*"), size = N,
    replace = TRUE),
  value = rnorm(N, 10, 3), score = rnorm(N, 100, 30),
  sample = sample(c("Normal", "Tumor"),
    size = N, replace = TRUE),
  pair = sample(letters, size = N,
    replace = TRUE))

## =====

```

```
## default
## =====
ggplot(gr) + geom_arrow()
# or
ggplot() + geom_arrow(gr)

## =====
## facetting and aesthetics
## =====
ggplot(gr) + geom_arrow(facets = sample ~ seqnames, mapping = aes(color = strand, fill = strand))

## =====
## stat:identity
## =====
ggplot(gr) + geom_arrow(stat = "identity", mapping = aes(y = value))

## =====
## stat:stepping
## =====
ggplot(gr) + geom_arrow(stat = "stepping", mapping = aes(y = value, group = pair))

## =====
## group.selfish
## =====
ggplot(gr) + geom_arrow(stat = "stepping", mapping = aes(y = value, group = pair), group.selfish = FALSE)

## =====
## other options to control arrow angle, density, ...
## =====
library(grid)
ggplot(gr) + geom_arrow(stat = "stepping", mapping = aes(y = value, group = pair),
  arrow.rate = 0.01, length = unit(0.3, "cm"), angle = 45,
  group.selfish = FALSE)
```

geom_arrowrect

Arrowrect geoms for GRanges object

Description

Show interval data as rectangle with a arrow head.

Usage

```
## S4 method for signature 'GRanges'
geom_arrowrect(data, ..., xlab, ylab, main,
  facets = NULL, stat = c("stepping", "identity"),
  rect.height = NULL, arrow.head = 0.06,
  arrow.head.rate = arrow.head, arrow.head.fix = NULL,
  group.selfish = TRUE)
```

Arguments

data	A GRanges object.
...	Extra parameters such as aes() passed.
xlab	Label for x
ylab	Label for y
main	Title for plot.
facets	Faceting formula to use.
stat	Character vector specifying statistics to use. "stepping" with randomly assigned stepping levels as y variable. "identity" allow users to specify y value in aes.
rect.height	Half height of the arrow body.
arrow.head	Arrow head to body ratio.
arrow.head.rate	Arrow head to body ratio. same with arrow.head.
arrow.head.fix	fixed length of arrow head.
group.selfish	Passed to addStepping, control whether to show each group as unique level or not. If set to FALSE, if two groups are not overlapped with each other, they will probably be layout in the same level to save space.

Value

A 'Layer'.

Author(s)

Tengfei Yin

Examples

```
set.seed(1)
N <- 100
require(GenomicRanges)
## =====
## simulated GRanges
## =====
gr <- GRanges(seqnames =
  sample(c("chr1", "chr2", "chr3"),
    size = N, replace = TRUE),
  IRanges(
    start = sample(1:300, size = N, replace = TRUE),
    width = sample(70:75, size = N, replace = TRUE)),
  strand = sample(c("+", "-", "*"), size = N,
    replace = TRUE),
  value = rnorm(N, 10, 3), score = rnorm(N, 100, 30),
  sample = sample(c("Normal", "Tumor"),
    size = N, replace = TRUE),
  pair = sample(letters, size = N,
    replace = TRUE))

## =====
## default
```

```

## =====
ggplot(gr) + geom_arrowrect()
## or
ggplot() + geom_arrowrect(gr)

## =====
## facetting and aesthetics
## =====
ggplot(gr) + geom_arrowrect(facets = sample ~ seqnames, mapping = aes(color = strand, fill = strand))

## =====
## stat:identity
## =====
ggplot(gr) + geom_arrowrect(stat = "identity", mapping = aes(y = value))

## =====
## stat:stepping
## =====
ggplot(gr) + geom_arrowrect(stat = "stepping", mapping = aes(y = value, group = pair))

## =====
## group.selfish controls when
## =====
ggplot(gr) + geom_arrowrect(gr, stat = "stepping", mapping = aes(y = value, group = pair), group.selfish = FALSE)

```

geom_bar

Segment geoms for GRanges object

Description

Show interval data as vertical bar, width equals to interval width and use 'score' or specified 'y' as y scale.

Usage

```

## S4 method for signature 'ANY'
geom_bar(data, ...)
## S4 method for signature 'GRanges'
geom_bar(data,..., xlab, ylab, main)

```

Arguments

data	Typically a GRanges or data.frame object.
...	Extra parameters such as aes() or color, size passed.
xlab	Label for x
ylab	Label for y
main	Title for plot.

Details

Useful for showing bed like files, when imported as GRanges, have a extra 'score' column, use it as default y, you could also specify y by using aes(y =).

Value

A 'Layer'.

Examples

```
## load
library(GenomicRanges)

## simul
set.seed(123)
gr.b <- GRanges(seqnames = "chr1", IRanges(start = seq(1, 100, by = 10),
      width = sample(4:9, size = 10, replace = TRUE)),
      score = rnorm(10, 10, 3), value = runif(10, 1, 100))
gr.b2 <- GRanges(seqnames = "chr2", IRanges(start = seq(1, 100, by = 10),
      width = sample(4:9, size = 10, replace = TRUE)),
      score = rnorm(10, 10, 3), value = runif(10, 1, 100))
gr.b <- c(gr.b, gr.b2)
## default use score as y

## bar
ggplot(gr.b) + geom_bar(mapping = aes(fill = value))
## or
ggplot() + geom_bar(gr.b, mapping = aes(fill = value))
ggplot(gr.b) + geom_bar(mapping = aes(y = value))
## equal to
autoplot(gr.b, geom = "bar")
```

geom_chevron

Chevron geoms for GRanges object

Description

Break normal intervals stroed in GRanges object and show them as chevron, useful for showing model or splice summary.

Usage

```
## S4 method for signature 'GRanges'
geom_chevron(data, ..., xlab, ylab, main,
  offset = 0.1,
  facets = NULL,
  stat = c("stepping", "identity"),
  chevron.height.rescale = c(0.1, 0.8),
  group.selfish = TRUE)
```

Arguments

<code>data</code>	A GRanges object.
<code>...</code>	Extra parameters passed to autoplot function.
<code>xlab</code>	Label for x
<code>ylab</code>	Label for y
<code>main</code>	Title for plot.
<code>offset</code>	A numeric value or characters. If it's numeric value, indicate how much you want the chevron to wiggle, usually the rectangle for drawing GRanges is of height unit 1, so it's better between -0.5 and 0.5 to make it nice looking. Unless you specify offset as one of those columns, this will use height of the chevron to indicate the columns. Of course you could use size of the chevron to indicate the column variable easily, please see the examples.
<code>facets</code>	faceting formula to use.
<code>stat</code>	character vector specifying statistics to use. "stepping" with randomly assigned stepping levels as y variable. "identity" allow users to specify y value in aes.
<code>chevron.height.rescale</code>	A numeric vector of length 2. When the offset parameters is a character which is one of the data columns, this parameter rescale the offset.
<code>group.selfish</code>	Passed to addStepping, control whether to show each group as unique level or not. If set to FALSE, if two groups are not overlapped with each other, they will probably be layout in the same level to save space.

Details

To draw a normal GRanges as Chevron, we need to provide a special geom for this purpose. Chevron is popular in gene viewer or genome browser, when they try to show isoforms or gene model. `geom_chevron`, just like any other `geom_*` function in `ggplot2`, you can pass `aes()` to it to use height of chevron or width of chevron to show statistics summary.

Value

A 'Layer'.

Author(s)

Tengfei Yin

Examples

```
set.seed(1)
N <- 100
require(GenomicRanges)

## =====
##  simulated GRanges
##  =====
gr <- GRanges(seqnames =
  sample(c("chr1", "chr2", "chr3"),
    size = N, replace = TRUE),
  IRanges(
    start = sample(1:300, size = N, replace = TRUE),
```

```

        width = sample(70:75, size = N, replace = TRUE)),
        strand = sample(c("+", "-", "*"), size = N,
            replace = TRUE),
        value = rnorm(N, 10, 3), score = rnorm(N, 100, 30),
        sample = sample(c("Normal", "Tumor"),
            size = N, replace = TRUE),
        pair = sample(letters, size = N,
            replace = TRUE))

## =====
## default
##
## =====
ggplot(gr) + geom_chevron()
## or
ggplot() + geom_chevron(gr)

## =====
## facetting and aesthetics
## =====
ggplot(gr) + geom_chevron(facets = sample ~ seqnames, mapping = aes(color = strand))

## =====
## stat:identity
## =====
ggplot(gr) + geom_chevron(stat = "identity", mapping = aes(y = value))

## =====
## stat:stepping
## =====
ggplot(gr) + geom_chevron(stat = "stepping", mapping = aes(group = pair))

## =====
## group.selfish controls when
## =====
ggplot(gr) + geom_chevron(stat = "stepping", mapping = aes(group = pair), group.selfish = FALSE,
    xlab = "xlab", ylab = "ylab", main = "main")

p <- qplot(x = mpg, y = cyl, data = mtcars)

## =====
## offset
## =====
gr2 <- GRanges("chr1", IRanges(c(1, 10, 20), width = 5))
gr2.p <- gaps(gr2)
## resize to connect them
gr2.p <- resize(gr2.p, fix = "center", width = width(gr2.p)+2)

ggplot(gr2) + geom_rect() + geom_chevron(gr2.p)

```

```
## notice the rectangle height is 0.8
## offset = 0 just like a line
ggplot(gr2) + geom_rect() + geom_chevron(gr2.p, offset = 0)

## equal height
ggplot(gr2) + geom_rect() + geom_chevron(gr2.p, offset = 0.4)

## =====
## chevron.height
## =====
values(gr2.p)$score <- c(100, 200)
ggplot(gr2) + geom_rect() + geom_chevron(gr2.p, offset = "score")
## chevron.height
ggplot(gr2) + geom_rect() + geom_chevron(gr2.p, offset = "score",
                                          chevron.height.rescale = c(0.4, 10))
```

geom_rect

Rect geoms for GRanges object

Description

Show interval data as rectangle.

Usage

```
## S4 method for signature 'ANY'
geom_rect(data, ...)
## S4 method for signature 'GRanges'
geom_rect(data,..., xlab, ylab, main,
          facets = NULL, stat = c("stepping", "identity"),
          rect.height = NULL,
          group.selfish = TRUE)
```

Arguments

data	Typically a GRanges or data.frame object. When it's data.frame, it's simply calling ggplot2::geom_rect.
...	Extra parameters such as aes() or color, size passed.
xlab	Label for x
ylab	Label for y
main	Title for plot.
facets	Faceting formula to use.
stat	Character vector specifying statistics to use. "stepping" with randomly assigned stepping levels as y variable. "identity" allow users to specify y value in aes.
rect.height	Half height of the arrow body.
group.selfish	Passed to addStepping, control whether to show each group as unique level or not. If set to FALSE, if two groups are not overlapped with each other, they will probably be layout in the same level to save space.

Value

A 'Layer'.

Author(s)

Tengfei Yin

Examples

```
set.seed(1)
N <- 100
require(GenomicRanges)

## =====
##  simulated GRanges
## =====
gr <- GRanges(seqnames =
  sample(c("chr1", "chr2", "chr3"),
    size = N, replace = TRUE),
  IRanges(
    start = sample(1:300, size = N, replace = TRUE),
    width = sample(70:75, size = N, replace = TRUE),
    strand = sample(c("+", "-", "*"), size = N,
      replace = TRUE),
    value = rnorm(N, 10, 3), score = rnorm(N, 100, 30),
    sample = sample(c("Normal", "Tumor"),
      size = N, replace = TRUE),
    pair = sample(letters, size = N,
      replace = TRUE))

## =====
##  data.frame call ggplot2::geom_rect
## =====
ggplot() + geom_rect(data = mtcars, mapping = aes(xmin = mpg, ymin = wt, xmax = mpg + 10, ymax = wt + 0.2,
  fill = cyl))

## =====
##  default
## =====
ggplot(gr) + geom_rect()
# or
ggplot() + geom_rect(gr)

## =====
##  facetting and aesthetics
## =====
ggplot(gr) + geom_rect(facets = sample ~ seqnames, mapping = aes(color = strand, fill = strand))

## =====
##  stat:identity
```

```
## =====
ggplot(gr) + geom_rect(stat = "identity", mapping = aes(y = value))

## =====
## stat:stepping
## =====
ggplot(gr) + geom_rect(stat = "stepping", mapping = aes(y = value, group = pair))

## =====
## group.selfish controls when
## =====
ggplot(gr) + geom_rect(stat = "stepping", mapping = aes(y = value, group = pair), group.selfish = FALSE)
```

geom_segment

Segment geoms for GRanges object

Description

Show interval data as segments.

Usage

```
## S4 method for signature 'ANY'
geom_segment(data, ...)
## S4 method for signature 'GRanges'
geom_segment(data,..., xlab, ylab, main,
             facets = NULL, stat = c("stepping", "identity"),
             group.selfish = TRUE)
```

Arguments

data	A GRanges or data.frame object.
...	Extra parameters such as aes() or color, size passed.
xlab	Label for x
ylab	Label for y
main	Title for plot.
facets	Faceting formula to use.
stat	Character vector specifying statistics to use. "stepping" with randomly assigned stepping levels as y variable. "identity" allow users to specify y value in aes.
group.selfish	Passed to addStepping, control whether to show each group as unique level or not. If set to FALSE, if two groups are not overlapped with each other, they will probably be layout in the same level to save space.

Value

A 'Layer'.

Author(s)

Tengfei Yin

Examples

```

set.seed(1)
N <- 100
require(GenomicRanges)

## =====
##  simulated GRanges
## =====
gr <- GRanges(seqnames =
  sample(c("chr1", "chr2", "chr3"),
    size = N, replace = TRUE),
  IRanges(
    start = sample(1:300, size = N, replace = TRUE),
    width = sample(70:75, size = N, replace = TRUE),
    strand = sample(c("+", "-", "*"), size = N,
      replace = TRUE),
    value = rnorm(N, 10, 3), score = rnorm(N, 100, 30),
    sample = sample(c("Normal", "Tumor"),
      size = N, replace = TRUE),
    pair = sample(letters, size = N,
      replace = TRUE))

## =====
##  data.frame call ggplot2::geom_segment
## =====
ggplot() + geom_segment(data = mtcars, mapping = aes(x = mpg, y = wt, xend = mpg + 10, yend = wt + 0.2,
  color = cyl))

## =====
##  default
##
##  =====
ggplot(gr) + geom_segment()
## or
ggplot() + geom_segment(gr)

## =====
##  facetting and aesthetics
## =====
ggplot(gr) + geom_segment(facets = sample ~ seqnames, mapping = aes(color = strand))

## =====
##  stat:identity
## =====
ggplot(gr) + geom_segment(stat = "identity", mapping = aes(y = value))

```

```
## =====
## stat:stepping
## =====
ggplot(gr) + geom_segment(stat = "stepping", mapping = aes(y = value, group = pair))

## =====
## group.selfish controls when
## =====
ggplot(gr) + geom_segment(stat = "stepping", mapping = aes(y = value, group = pair), group.selfish = FALSE)
```

GGbio

*class ggbio***Description**

a sub class of ggplot and gg class defined in ggplot2 package, used for ggbio specific methods.

Usage

```
GGbio(ggplot = NULL, data = NULL, fetchable = FALSE, blank =
      FALSE, ...)
```

Arguments

ggplot	a ggplot or gg object.
data	raw data
fetchable	logical value, default FALSE, is there any fetch method available.
blank	logical value, default FALSE, is this plot a blank plot.
...	More properties passed to class like Cache.

Details

This class is defined to facilitate the ggbio-specific visualization method, especially when using [ggplot](#) to construct ggbio supported object, that will return a ggbio class. And internals tricks will help a lazy evaluation for following + method.

Value

a ggbio object.

Author(s)

Tengfei Yin

See Also

[ggplot](#)

Examples

```
p1 <- qplot()
g1 <- ggbio(p1)
class(g1)
```

ggplotggplot methods

Description

These methods extend ggplot to support several types of Bioconductor objects, as well as some base types like matrix. They return a ggbio object, which stores the original data object. Please check the corresponding method for [mold](#) to see how an object is coerced into a data.frame.

Usage

```
## S3 method for class 'Vector'
ggplot(data, mapping = aes(), ...,
        environment = parent.frame())
## S3 method for class 'Seqinfo'
ggplot(data, mapping = aes(), ...,
        environment = parent.frame())
## S3 method for class 'ExpressionSet'
ggplot(data, mapping = aes(), ...,
        environment = parent.frame())
## S3 method for class 'RsamtoolsFile'
ggplot(data, mapping = aes(), ...,
        environment = parent.frame())
## S3 method for class 'TxDbOREnsDb'
ggplot(data, mapping = aes(), ...,
        environment = parent.frame())
## S3 method for class 'BSgenome'
ggplot(data, mapping = aes(), ...,
        environment = parent.frame())
## S3 method for class 'matrix'
ggplot(data, mapping = aes(), ...,
        environment = parent.frame())
## S3 method for class 'character'
ggplot(data, mapping = aes(), ...,
        environment = parent.frame())
## S3 method for class 'SummarizedExperiment'
ggplot(data, mapping = aes(),
        assay.id = 1L, ..., environment = parent.frame())
## S3 method for class 'GAlignments'
ggplot(data, mapping = aes(), ...,
        environment = parent.frame())
## S3 method for class 'VCF'
ggplot(data, mapping = aes(), ...,
        environment = parent.frame())
```

Arguments

data	original data object.
mapping	the aesthetic mapping.
...	other arguments passed to specific methods.

environment	fall-back environment for evaluation of aesthetic symbols
assay.id	index of assay you are using when multiple assays exist.

Details

The biggest difference for objects returned by `ggplot` in `ggbio` from `ggplot2`, is we always keep the original data copy, this is useful because in `ggbio`, our starting point is not always `data.frame`, many special statistical transformation is computed upon original data objects instead of coerced `data.frame`. This is a hack to follow `ggplot2`'s API while allow our own defined components to trace back to original data copy and do the transformation. For objects supported by `mold` we transform them to `data.frame` stored along the original data set, for objects which not supported by `mold` method, we only store the original copy for `ggbio` specific graphics.

`ggplot()` is typically used to construct a plot incrementally, using the `+` operator to add layers to the existing `ggplot` object. This is advantageous in that the code is explicit about which layers are added and the order in which they are added. For complex graphics with multiple layers, initialization with `ggplot` is recommended. You can always call `qplot` in package `ggplot2` or `autoplot` in `ggbio` for convenient usage.

There are three common ways to invoke `ggplot`:

- `ggplot(df, aes(x, y, <other aesthetics>))`
- `ggplot(df)`
- `ggplot()`

The first method is recommended if all layers use the same data and the same set of aesthetics, although this method can also be used to add a layer using data from another data frame. The second method specifies the default data frame to use for the plot, but no aesthetics are defined up front. This is useful when one data frame is used predominantly as layers are added, but the aesthetics may vary from one layer to another. The third method initializes a skeleton `ggplot` object which is fleshed out as layers are added. This method is useful when multiple data frames are used to produce different layers, as is often the case in complex graphics.

The examples below illustrate how these methods of invoking `ggplot` can be used in constructing a graphic.

Value

a return `ggbio` object, which is a subclass of `ggplot` defined in `ggplot2` package, but that's more, a `'data'` list entry is stored with the returned object.

Author(s)

Tengfei Yin

See Also

[mold](#), [ggbio](#)

Examples

```
set.seed(1)
N <- 100
library(GenomicRanges)
## GRanges
gr <- GRanges(seqnames =
```

```

sample(c("chr1", "chr2", "chr3"),
       size = N, replace = TRUE),
IRanges(
  start = sample(1:300, size = N, replace = TRUE),
  width = sample(70:75, size = N, replace = TRUE)),
strand = sample(c("+", "-", "*"), size = N,
  replace = TRUE),
value = rnorm(N, 10, 3), score = rnorm(N, 100, 30),
sample = sample(c("Normal", "Tumor"),
  size = N, replace = TRUE),
pair = sample(letters, size = N,
  replace = TRUE))

## automatically facetting and assign y
## this must mean geom_rect support GRanges object
ggplot(gr) + geom_rect()
ggplot(gr) + geom_alignment()
ggplot() + geom_alignment(gr)

## use pure ggplot2's geom_rect, no auto facet
ggplot(gr) + ggplot2::geom_rect(aes(xmin = start, ymin = score,
                                   xmax = end, ymax = score + 1))

## GRangesList
grl <- split(gr, values(gr)$pair)
ggplot(grl) + geom_alignment()
ggplot(grl) + geom_rect()
ggplot(grl) + ggplot2::geom_rect(aes(xmin = start, ymin = score,
                                   xmax = end, ymax = score + 1))

## IRanges
ir <- ranges(gr)
ggplot(ir) + geom_rect()
ggplot(ir) + layout_circle(geom = "rect")

## Seqinfo
seqlengths(gr) <- c(400, 500, 420)
ggplot(seqinfo(gr)) + geom_point(aes(x = midpoint, y = seqlengths))

## matrix
mx <- matrix(1:12, nrow = 3)
ggplot(mx, aes(x = x, y = y)) + geom_raster(aes(fill = value))
## row is the factor
ggplot(mx, aes(x = x, y = row)) + geom_raster(aes(fill = value))
colnames(mx)
colnames(mx) <- letters[1:ncol(mx)]
mx
## has extra 'colnames'
ggplot(mx, aes(x = x, y = row)) + geom_raster(aes(fill = colnames))
rownames(mx)
rownames(mx) <- LETTERS[1:nrow(mx)]
ggplot(mx, aes(x = x, y = row)) + geom_raster(aes(fill = rownames))
## please check autoplot, matrix for more control

## Views

```

```

## ExpressionSet
library(Biobase)
data(sample.ExpressionSet)
sample.ExpressionSet
set.seed(1)
## select 50 features
idx <- sample(seq_len(dim(sample.ExpressionSet)[1]), size = 50)
eset <- sample.ExpressionSet[idx,]

ggplot(eset) + geom_tile(aes(x = x, y = y, fill = value))
## please check autoplot,matrix method which gives you more control
ggplot(eset) + geom_tile(aes(x = x, y = y, fill = sex))
ggplot(eset) + geom_tile(aes(x = x, y = y, fill = type))

## Rle
library(IRanges)
lambda <- c(rep(0.001, 4500), seq(0.001, 10, length = 500),
            seq(10, 0.001, length = 500))
xVector <- rpois(1e4, lambda)
xRle <- Rle(xVector)
ggplot(xRle) + geom_tile(aes(x = x, y = y, fill = value))

## RleList
xRleList <- RleList(xRle, 2L * xRle)
xRleList
ggplot(xRleList) + geom_tile(aes(x = x, y = y, fill = value)) +
  facet_grid(group~.)
names(xRleList) <- c("a", "b")
ggplot(xRleList) + geom_tile(aes(x = x, y = y, fill = value)) +
  facet_grid(group~.)

## RangedSummarizedExperiment
library(SummarizedExperiment)
nrows <- 200; ncols <- 6
counts <- matrix(runif(nrows * ncols, 1, 1e4), nrows)
counts2 <- matrix(runif(nrows * ncols, 1, 1e4), nrows)
rowRanges <- GRanges(rep(c("chr1", "chr2"), c(50, 150)),
                     IRanges(floor(runif(200, 1e5, 1e6)), width=100),
                     strand=sample(c("+", "-"), 200, TRUE))
colData <- DataFrame(Treatment=rep(c("ChIP", "Input"), 3),
                    row.names=LETTERS[1:6])
sset <- SummarizedExperiment(assays=SimpleList(counts=counts,
                                                counts2 = counts2),
                           rowRanges=rowRanges, colData=colData)
ggplot(sset) + geom_raster(aes(x = x, y = y , fill = value))

```

ggsave

Save a ggplot object or tracks with sensible defaults

Description

ggsave is a convenient function for saving a plot. It defaults to saving the last plot that you displayed, and for a default size uses the size of the current graphics device. It also guesses the type of graphics

device from the extension. This means the only argument you need to supply is the filename.

Usage

```
ggsave(filename, plot = last_plot(),
        device = default_device(filename), path = NULL,
        scale = 1, width = par("din")[1],
        height = par("din")[2], units = c("in", "cm", "mm"),
        dpi = 300, limitsize = TRUE, ...)
```

Arguments

filename	file name/filename of plot
plot	plot to save, defaults to last plot displayed
device	device to use, automatically extract from file name extension
path	path to save plot to (if you just want to set path and not filename)
scale	scaling factor
width	width (defaults to the width of current plotting window)
height	height (defaults to the height of current plotting window)
units	units for width and height when either one is explicitly specified (in, cm, or mm)
dpi	dpi to use for raster graphics
limitsize	when TRUE (the default), ggsave will not save images larger than 50x50 inches, to prevent the common error of specifying dimensions in pixels.
...	other arguments passed to graphics device

Details

ggsave currently recognises the extensions eps/ps, tex (pictex), pdf, jpeg, tiff, png, bmp, svg and wmf (windows only).

Grob-class	<i>Grob getter</i>
------------	--------------------

Description

'Grob' class is a container for 'grob' based object defined with grid system. Generic function Grob gets grob object supported by grid system, and make an instance of subclass of class 'Grob'.

'GrobList' is a container of list of 'Grob' object.

Usage

```
## S4 method for signature 'gg'
Grob(x)
## S4 method for signature 'gtable'
Grob(x)
## S4 method for signature 'trellis'
Grob(x)
## S4 method for signature 'lattice'
Grob(x)
## S4 method for signature 'GGbio'
Grob(x)
```

Arguments

x object of class: gg, gtable, trellis, lattice, GGbio.

Value

A Grob object.

Author(s)

Tengfei Yin

Ideogram	<i>Plot single chromosome with cytobands</i>
----------	--

Description

Plot single chromosome with cytobands.

Usage

```
plotIdeogram(obj, subchr = NULL, zoom.region = NULL, which = NULL, xlab, ylab, main, xlabel =
  FALSE, color = "red", fill = "red", alpha = 0.7,
  zoom.offset = 0.2, size = 1,
  cytobands = TRUE, aspect.ratio = 1/20, genome)
```

constructor

```
Ideogram(obj, subchr = NULL, which = NULL, xlabel = FALSE,
  cytobands = TRUE, color = "red", fill = "red", alpha =
  0.7, zoom.region = NULL, zoom.offset = 0.2, size = 1,
  aspect.ratio = 1/20, ..., genome)
```

Arguments

obj	A GenomicRanges object, which include extra information about cytobands, check biovizBase::isIdeogram.
subchr	A single character of chromosome names to show.
which	GRanges object to subset and highlight the ideogram.
zoom.region	A numeric vector of length 2 indicating zoomed region.
xlab	Label for x
ylab	Label for y
main	Title for plot.
xlabel	A logical value. Show the x label or not.
color	color for highlight region.
fill	fill color for highlight region.
alpha	alpha for highlight regio.
zoom.offset	zoomed highlights region offset around chromosome plotting region.
size	size for zoomed region rectangle boundary.

cytobands	If FALSE, plot just blank chromosome without cytobands. default is TRUE. es
aspect.ratio	aspect ratio for the chromosome ideogram plot, default is NULL.
genome	genome character passed to getIdeogram
...	passed to ggbio constructor.

Details

User could provide the whole ideogram and use subchr to point to particular chromosome.

Value

A ggplot object.

Author(s)

Tengfei Yin

Examples

```
## Not run:
library(biovizBase)
p.ideo <- Ideogram(genome = "hg19")
p.ideo
library(GenomicRanges)
p.ideo + xlim(GRanges("chr2", IRanges(1e8, 1e8+10000)))
Ideogram(genome = "hg19", xlabel = TRUE)

## End(Not run)
```

layout_circle	<i>Create a circle layout</i>
---------------	-------------------------------

Description

Create a circle layout.

Usage

```
## S4 method for signature 'GRanges'
layout_circle(data, ..., geom = c("point", "line", "link", "ribbon",
  "rect", "bar", "segment", "hist", "scale", "heatmap", "ideogram",
  "text"), linked.to, radius = 10, trackWidth = 5,
  space.skip = 0.015, direction = c("clockwise",
  "anticlockwise"), link.fun = function(x, y, n = 30)
  bezier(x, y, evaluation = n), rect.inter.n = 60, rank,
  ylim = NULL,
  scale.n = 60, scale.unit = NULL, scale.type = c("M",
  "B", "sci"), grid.n = 5, grid.background = "gray70",
  grid.line = "white", grid = FALSE, chr.weight = NULL)

## S4 method for signature 'missing'
layout_circle(data, ...)
circle(...)
```

Arguments

<code>data</code>	A GRanges object.
<code>...</code>	Extra parameters such as aesthetics mapping in <code>aes()</code> , or <code>color</code> , <code>size</code> , etc. For <code>circle</code> function, it passed to <code>layout_circle</code> .
<code>geom</code>	The geometric object to use display the data.
<code>linked.to</code>	Character indicates column that specifying end of the linking lines, that column should be a GRanges object.
<code>radius</code>	Numeric value indicates radius. Default is 10.
<code>trackWidth</code>	Numeric value indicates the track width.
<code>space.skip</code>	Numeric value indicates the ratio of skipped region between chunks(chromosomes in GRanges) to the whole track space.
<code>direction</code>	Space layout orders.
<code>link.fun</code>	Function used for interpolate the linking lines. Default is <code>Hmisc::bezier</code> .
<code>rect.inter.n</code>	<code>n</code> passed to interpolate function in rectangle transformation(from a rectangle) to a section in circular view.
<code>rank</code>	For default equal trackWidth, use rank to specify the circle orders.
<code>ylim</code>	Numeric range to control y limits.
<code>scale.n</code>	Approximate number of ticks you want to show on the whole space. used when <code>scale.unit</code> is NULL.
<code>scale.unit</code>	Unit used for computing scale. Default is NULL,
<code>scale.type</code>	Scale type used for
<code>grid</code>	logical value indicate showing grid background for track or not.
<code>grid.n</code>	integer value indicate horizontal grid line number.
<code>grid.background</code>	grid background color.
<code>grid.line</code>	grid line color.
<code>chr.weight</code>	numeric vectors which sum to <1, the names of vectors has to be matched with <code>seqnames</code> in <code>seqinfo</code> , and you can only specify part of the <code>seqnames</code> , other lengths of chromosomes will be assigned proportionally to their <code>seqlengths</code> , for example, you could specify <code>chr1</code> to be 0.5, so the <code>chr1</code> will take half of the space and other chromosomes squeezed to take left of the space.

Value

A 'Layer'.

Author(s)

Tengfei Yin

Examples

```
N <- 100
library(GenomicRanges)
## =====
## simulated GRanges
## =====
```

```

gr <- GRanges(seqnames =
  sample(c("chr1", "chr2", "chr3"),
    size = N, replace = TRUE),
  IRanges(
    start = sample(1:300, size = N, replace = TRUE),
    width = sample(70:75, size = N, replace = TRUE)),
  strand = sample(c("+", "-", "*"), size = N,
    replace = TRUE),
  value = rnorm(N, 10, 3), score = rnorm(N, 100, 30),
  sample = sample(c("Normal", "Tumor"),
    size = N, replace = TRUE),
  pair = sample(letters, size = N,
    replace = TRUE))

seqlengths(gr) <- c(400, 500, 700)
values(gr)$to.gr <- gr[sample(1:length(gr), size = length(gr))]

## doesn't pass gr to the ggplot
ggplot() + layout_circle(gr, geom = "ideo", fill = "gray70", radius = 7, trackWidth = 3) +
  layout_circle(gr, geom = "bar", radius = 10, trackWidth = 4, aes(fill = score, y = score)) +
  layout_circle(gr, geom = "point", color = "red", radius = 14,
    trackWidth = 3, grid = TRUE, aes(y = score)) +
  layout_circle(gr, geom = "link", linked.to = "to.gr", radius = 6,
    trackWidth = 1)

## more formal API
ggplot(gr) + layout_circle(geom = "ideo", fill = "gray70", radius = 7, trackWidth = 3) +
  layout_circle(geom = "bar", radius = 10, trackWidth = 4, mapping = aes(fill = score, y = score)) +
  layout_circle(geom = "point", color = "red", radius = 14,
    trackWidth = 3, grid = TRUE, mapping = aes(y = score)) +
  layout_circle(geom = "link", linked.to = "to.gr", radius = 6, trackWidth = 1)

```

layout_karyogram	Create a karyogram layout
------------------	---------------------------

Description

Create a karyogram layout.

Usage

```

## S4 method for signature 'GRanges'
layout_karyogram(data, ..., xlab, ylab, main,
  facets = seqnames ~ ., cytobands = FALSE, geom = "rect",
  stat = NULL, ylim = NULL, rect.height = 10)

```

Arguments

data	a GRanges object, which could contain extra information about cytobands. If you want an accurate genome mapping, please provide seqlengths with this GRanges object, otherwise it will emit a warning and use data space to estimate the chromosome space which is very rough.
------	--

...	Extra parameters such as aes() or arbitrary color and size.
xlab	character vector or expression for x axis label.
ylab	character vector or expression for y axis label.
main	character vector or expression for plot title.
facets	faceting formula to use.
cytobands	logical value indicate to show the cytobands or not.
geom	The geometric object to use display the data.
stat	character vector specifying statistics to use.
ylim	limits for y axis, usually the chromosome spaces y limits are from 0 to rect.height, which 10, so if you want to stack some data on top of it, you can set limits to like c(10, 20).
rect.height	numeric value indicate half of the rectangle plotting region, used for alignment of multiple layers.

Value

A 'Layer'.

Author(s)

Tengfei Yin

Examples

```
### R code from vignette source 'karyogram.Rnw'

#####
### code chunk number 1: loading
#####
library(ggbio)
data(hg19IdeogramCyto, package = "biovizBase")
head(hg19IdeogramCyto)
## default pre-set color stored in
getOption("biovizBase")$cytobandColor

#####
### code chunk number 2: default
#####
autoplot(hg19IdeogramCyto, layout = "karyogram", cytobands = TRUE)

#####
### code chunk number 3: change-order
#####
library(GenomeInfoDb) # for keepSeqlevels()
hg19 <- keepSeqlevels(hg19IdeogramCyto, paste0("chr", c(1:22, "X", "Y")))
head(hg19)
autoplot(hg19, layout = "karyogram", cytobands = TRUE)

#####
```

```

#### code chunk number 4: cyto-normal
#####
library(GenomicRanges)
## it's a 'ideogram'
biovizBase::isIdeogram(hg19)
## set to FALSE
autoplot(hg19, layout = "karyogram", cytobands = FALSE, aes(fill = gieStain)) +
  scale_fill_giensa()

#####
#### code chunk number 5: load-RNAediting
#####
data(darned_hg19_subset500, package = "biovizBase")
dn <- darned_hg19_subset500
head(dn)
## add seqlengths
## we have seqlegnth information in another data set
data(hg19Ideogram, package = "biovizBase")
seqlengths(dn) <- seqlengths(hg19Ideogram)[names(seqlengths(dn))]
## now we have seqlengths
head(dn)
## then we change order
dn <- keepSeqlevels(dn, paste0("chr", c(1:22, "X")))
autoplot(dn, layout = "karyogram")
## this equivalent to
## autoplot(seqinfo(dn))

#####
#### code chunk number 6: load-RNAediting-color
#####
## since default is geom rectangle, even though it's looks like segment
## we still use both fill/color to map colors
autoplot(dn, layout = "karyogram", aes(color = exReg, fill = exReg))

#####
#### code chunk number 7: load-RNAediting-color-NA
#####
## since default is geom rectangle, even though it's looks like segment
## we still use both fill/color to map colors
autoplot(dn, layout = "karyogram", aes(color = exReg, fill = exReg)) +
  scale_color_discrete(na.value = "brown")

#####
#### code chunk number 8: load-RNAediting-color-fake
#####
dn2 <- dn
seqlengths(dn2) <- rep(max(seqlengths(dn2)), length(seqlengths(dn2)))
autoplot(dn2, layout = "karyogram", aes(color = exReg, fill = exReg))

#####
#### code chunk number 9: plotKaryogram (eval = FALSE)
#####

```

```

## plotKaryogram(dn)
## plotKaryogram(dn, aes(color = exReg, fill = exReg))

#####
### code chunk number 10: low-default
#####
## plot ideogram
p <- ggplot(hg19) + layout_karyogram(cytobands = TRUE)
p
## equevalant autoplot(hg19, layout = "karyogram", cytobands = TRUE)

#####
### code chunk number 11: low-default-addon
#####
p <- p + layout_karyogram(dn, geom = "rect", ylim = c(11, 21), color = "red")
## commented line below won't work
## the cytoband fill color has been used already.
## p <- p + layout_karyogram(dn, aes(fill = exReg, color = exReg), geom = "rect")
p

#####
### code chunk number 12: edit-space
#####
## plot chromosome space
p <- autoplot(seqinfo(dn))
## make sure you pass rect as geom
## otherwise you just get background
p <- p + layout_karyogram(dn, aes(fill = exReg, color = exReg), geom = "rect")
values(dn)$pvalue <- rnorm(length(dn))
p + layout_karyogram(dn, aes(x = start, y = pvalue), ylim = c(10, 30), geom = "line", color = "red")
p

#####
### code chunk number 13: sessionInfo
#####
sessionInfo()

```

Plot

Plot class

Description

genealize a graphic object to a Plot object.

Usage

```

## S4 method for signature 'gg'
Plot(x)
## S4 method for signature 'trellis'
Plot(x, mutable = FALSE)

```



```
## S4 method for signature 'GGbio'
Plot(x)
## S4 method for signature 'Ideogram'
Plot(x)
```

Arguments

x	object of gg, GGbio, trellis, Ideogram.
mutable	whether a plot response to + method or not.

Value

A Plot object.

Author(s)

Tengfei Yin

plotFragLength	<i>Plot estimated fragment length for paired-end RNA-seq data</i>
----------------	---

Description

Plot estimated fragment length for paired-end RNA-seq data against single reduced data model.

Usage

```
## S4 method for signature 'character,GRanges'
plotFragLength(data, model,
               gap.ratio = 0.0025,
               geom = c("segment", "point", "line"),
               type = c("normal", "cut"),
               heights = c(400, 100),
               annotation = TRUE)
```

Arguments

data	A character indicate the bam file.
model	A reduced model to compute estimated fragment length. please see details.
gap.ratio	When type is set to "cut", it will provide a compact view, which cut the common gaps in a certain ratio.
geom	One or all three geoms could be drawn at the same time. y value of "point" and "line" indicate the estimated fragment length. and if geom is set to "segment", the segment is from the left most position to paired right most position, should be equal to "isize".
type	"normal" return a uncut view, loose but the coordinate is true genomic coordinates. "cut" cut the view in a compact way.
heights	Numeric vector indicate the heights of tracks.
annotation	A logical value. TRUE shows model, and FALSE shows only fragment length with labels.

Details

We use a easy way to define this estimated fragment length, we collect all paired reads and model, reduce model first, then find common gaps, remove common gaps between paired-end reads, and compute the new estimated fragment length.

Value

A ggplot object when annotation = FALSE and a frame grob if annotation = TRUE

Author(s)

Tengfei Yin

Examples

```
## Not run:
data(genesymbol)
bamfile <- system.file("extdata", "SRR027894subRBM17.bam", package="biovizBase")
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
model <- exonsBy(txdb, by = "tx")
model.new <- subsetByOverlaps(model, genesymbol["RBM17"])
exons.rbm17 <- subsetByOverlaps(exons(txdb), genesymbol["RBM17"])
exons.new <- reduce(exons.rbm17)
plotFragLength(bamfile, exons.new, geom = "line")
plotFragLength(bamfile, exons.new, geom = c("point", "segment"))
plotFragLength(bamfile, exons.new, geom = c("point", "segment"), annotation = FALSE)
plotFragLength(bamfile, exons.new, geom = c("point", "segment"), type = "cut",
               gap.ratio = 0.001)

## End(Not run)
```

plotGrandLinear

Manhattan for GWAS

Description

A Manhattan plot is special scatter plot used to visualize data with a large number of data points, with a distribute of some higher-magnitude values. For example, in the GWAS(genome-wide association studies). Here we mainly focus on GWAS Manhattan plots. X-axis is genomic coordinates and Y-axis is negative logarithm of the associated P-value for each single nucleotide polymorphism. So higher the value, more stronger the association they are.

Usage

```
plotGrandLinear(obj, ..., facets, space.skip = 0.01, geom = NULL,
               cutoff = NULL, cutoff.color = "red", cutoff.size = 1,
               legend = FALSE, xlim, ylim, xlab, ylab, main,
               highlight.gr = NULL, highlight.name = NULL,
               highlight.col = "red", highlight.label = TRUE,
               highlight.label.size = 5, highlight.label.offset =
               0.05, highlight.label.col = "black", spaceline =
               FALSE)
```

Arguments

obj	GRanges object which contains extra p value, before users pass this object, they need to make sure the pvalue has been changed to $-\log_{10}(p)$.
...	extra arguments passed. such as color, size, alpha.
facets	facets formula, such as group ~ .
space.skip	numeric value for skip ratio, between chromosome spaces.default is 0.01.
geom	geometric object, default is "point".
cutoff	A numeric vector which used as cutoff for Manhattan plot.
cutoff.color	A character specifying the color used for cutoff. Default is "red".
cutoff.size	A numeric value which used as cutoff line size.
legend	A logical value indicate whether to show legend or not. Default is FALSE which disabled the legend.
xlim	limits for x scale.
ylim	limits for y scale.
xlab	Label for xscale.
ylab	Label for yscale.
main	title.
highlight.gr	a GRanges object, this wil highlight overlapped region with provided intervals.
highlight.name	if NULL, using rownames of GRanges object provided by argument highlight.gr, otherwise use character to indicate column used as labeled names.
highlight.col	highlight colors.
highlight.label	logical value, label the highlighted region of not.
highlight.label.size	highlight label size.
highlight.label.offset	highlight label offset.
highlight.label.col	highlight label color.
spaceline	show line between chromosomes.

Details

Please use seqlengths of the object and space.skip arguments to control the layout of the coordiant genome transformation.

aes(y = ...) is requiried.

aes(color =) is used to mapping to data variables, if just pass "color" without aes(), then will recycle the color to represent each chromosomes.please see the example below.

Value

Return a ggplot object.

Author(s)

Tengfei Yin

Examples

```

## load
library(ggbio)
data(hg19IdeogramCyto, package = "biovizBase")
data(hg19Ideogram, package = "biovizBase")
library(GenomicRanges)

## simul_gr
library(biovizBase)
gr <- GRanges(rep(c("chr1", "chr2"), each = 5),
               IRanges(start = rep(seq(1, 100, length = 5), times = 2),
                       width = 50))

autoplot(gr)

## coord:genome
autoplot(gr, coord = "genome")
gr.t <- transformToGenome(gr)
head(gr.t)

## is
is_coord_genome(gr.t)
metadata(gr.t)$coord

## simul_snp
chrs <- as.character(levels(seqnames(hg19IdeogramCyto)))
seqlths <- seqlengths(hg19Ideogram)[chrs]
set.seed(1)
nchr <- length(chrs)
nsnps <- 100
gr.snp <- GRanges(rep(chrs, each=nsnps),
                  IRanges(start =
                        do.call(c, lapply(chrs, function(chr){
                          N <- seqlths[chr]
                          runif(nsnps, 1, N)
                        })), width = 1),
                  SNP=sapply(1:(nchr*nsnps), function(x) paste("rs", x, sep="")),
                  pvalue = -log10(runif(nchr*nsnps)),
                  group = sample(c("Normal", "Tumor"), size = nchr*nsnps,
                                replace = TRUE)
                  )

## shorter
seqlengths(gr.snp)
nms <- seqnames(seqinfo(gr.snp))
nms.new <- gsub("chr", "", nms)
names(nms.new) <- nms
library(GenomeInfoDb) # for renameSeqlevels() and keepSeqlevels()
gr.snp <- renameSeqlevels(gr.snp, nms.new)
seqlengths(gr.snp)

## unordered
autoplot(gr.snp, coord = "genome", geom = "point", aes(y = pvalue), space.skip = 0.01)

```

```

## sort
gr.snp <- keepSeqlevels(gr.snp, c(1:22, "X", "Y"))
autoplot(gr.snp, coord = "genome", geom = "point", aes(y = pvalue), space.skip = 0.01)

## with_seq
names(seqLths) <- gsub("chr", "", names(seqLths))
seqlengths(gr.snp) <- seqLths[names(seqlengths(gr.snp))]
autoplot(gr.snp, coord = "genome", geom = "point", aes(y = pvalue), space.skip = 0.01)

## line
autoplot(gr.snp, coord = "genome", geom = "line", aes(y = pvalue, group = seqnames,
                                                    color = seqnames))

## plotGrandLinear
plotGrandLinear(gr.snp, aes(y = pvalue))

## morecolor
plotGrandLinear(gr.snp, aes(y = pvalue, color = seqnames))
plotGrandLinear(gr.snp, aes(y = pvalue), color = c("green", "deepskyblue"))
plotGrandLinear(gr.snp, aes(y = pvalue), color = c("green", "deepskyblue", "red"))
plotGrandLinear(gr.snp, aes(y = pvalue), color = "red")

## cutoff
plotGrandLinear(gr.snp, aes(y = pvalue), cutoff = 3, cutoff.color = "blue", cutoff.size = 4)

## cutoff-low
plotGrandLinear(gr.snp, aes(y = pvalue)) + geom_hline(yintercept = 3, color = "blue", size = 4)

## longer
## let's make a long name
nms <- seqnames(seqinfo(gr.snp))
nms.new <- paste("chr00000", nms, sep = "")
names(nms.new) <- nms
gr.snp <- renameSeqlevels(gr.snp, nms.new)
seqlengths(gr.snp)

## rotate
plotGrandLinear(gr.snp, aes(y = pvalue)) + theme(axis.text.x=element_text(angle=-90, hjust=0))

## sessionInfo
sessionInfo()

```

plotRangesLinkedToData

Plot Ranges Linked with Data

Description

Plot GRanges object structure and linked to a even spaced parallell coordinates plot which represting the data in elementeMetadata.

Usage

```
## S4 method for signature 'RangedSummarizedExperiment'
plotRangesLinkedToData(data, ...,
  stat.y = seq_len(ncol(data)), stat.ylab = names(assays(data)[stat.assay]),
  stat.assay = 1L)

## S4 method for signature 'GenomicRanges_OR_GRangesList'
plotRangesLinkedToData(data, ...,
  stat.y = seq_len(ncol(mcols(data))),
  stat.ylab, sig, sig.col = c("black", "red"),
  stat.coord.trans = coord_trans(),
  annotation = list(), width.ratio = 0.8,
  theme.stat = theme_gray(), theme.align = theme_gray(),
  linetype = 3, heights)
```

Arguments

<code>data</code>	GRanges object with a DataFrame as elementMetadata.
<code>...</code>	Parameters passed to control lines in top plot.
<code>stat.y</code>	integer (variable position starting in DataFrame of data, start from 1) or strings (variable names) which indicate the column names.
<code>stat.ylab</code>	y label for stat track(the top track).
<code>stat.assay</code>	default 1L, element of assays.
<code>sig</code>	a character of element meta data column of logical value, indicates which row is significant. and will be shown in link lines and rectangle.
<code>sig.col</code>	colors for significant, valid when you specify "sig" argument, the first color indicates FALSE, non-significant, the second color indicates TRUE.
<code>stat.coord.trans</code>	transformation used for top plot.
<code>annotation</code>	A list of ggplot object.
<code>width.ratio</code>	Control the segment length of statistic layer.
<code>theme.stat</code>	top plot theme.
<code>theme.align</code>	alignment themes.
<code>linetype</code>	linetype
<code>heights</code>	Heights of each track.

Details

Inspired by some graphics produced in some other packages, for example in package DEXseq, the author provides graphics with gene models and linked to an even spaced statistics summary. This is useful because we always plot everything along the genomic coordinates, but genomic features like exons are not evenly distributed, so we could actually treat the statistics associated with exons like categorical data, and show them as "Paralell Coordinates Plots". This is one special layout which represent the data in a nice manner and also keep the genomic structure information. With abliity of tracks, it's possible to generate such type of a graphic along with other annotations.

The data we want is a normal GRanges object, and make sure the intervals are not overlaped with each other(currently), and you may have multiple columns which store the statistics for multiple samples, then we produce the graphic we introduced above and users could pass other annotation track in the function which will be shown below the main linked track.

The reason you need to pass annotation into the function instead of binding them by tracks later is because binding manually with annotation tracks is tricky and this function doesn't return a ggplot object.

Value

return a frame grob; side-effect (plotting) if plot=T.

Author(s)

Tengfei Yin

Examples

```
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
library(ggbio)
data(genesymbol, package = "biovizBase")
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
model <- exonsBy(txdb, by = "tx")
model17 <- subsetByOverlaps(model, genesymbol["RBM17"])
exons <- exons(txdb)
exon17 <- subsetByOverlaps(exons, genesymbol["RBM17"])
## reduce to make sure there is no overlap
## just for example
exon.new <- reduce(exon17)
## suppose
values(exon.new)$sample1 <- rnorm(length(exon.new), 10, 3)
values(exon.new)$sample2 <- rnorm(length(exon.new), 10, 10)
values(exon.new)$score <- rnorm(length(exon.new))
values(exon.new)$significant <- sample(c(TRUE,FALSE), size = length(exon.new),replace = TRUE)

plotRangesLinkedToData(exon.new, stat.y = c("sample1", "sample2"))
plotRangesLinkedToData(exon.new, stat.y = 1:2)
plotRangesLinkedToData(exon.new, stat.y = 1:2, size = 3, linetype = 4)
plotRangesLinkedToData(exon.new, stat.y = 1:2, size = 3, linetype = 4,
                        sig = "significant")
plotRangesLinkedToData(exon.new, stat.y = 1:2, size = 3, linetype = 4,
                        sig = "significant", sig.col = c("gray90","red"))
```

plotSpliceSum

Plot Splice Summary from RNA-seq data

Description

Plot splice summary by simply counting overlaped junction read in weighted way or not.

Usage

```
## For character,GRangesList
## S4 method for signature 'character,GRangesList'
plotSpliceSum(data, model, ..., weighted = TRUE)
## For character,TxDb
## S4 method for signature 'character,TxDb'
plotSpliceSum(data, model, which,
  ..., weighted = TRUE)
## For character,EnsDb
## S4 method for signature 'character,EnsDb'
plotSpliceSum(data, model, which,
  ..., weighted = TRUE)
```

Arguments

data	A character specifying the bam file path of RNA-seq data.
model	A GRangesList which representing different isoforms, a TxDb or an EnsDb object. For the latter cases, users need to pass "which" argument which, for TxDb, is a GRanges object to specify the region and for EnsDb can be a GRanges object, an object extending AnnotationFilter , an AnnotationFilterList combining such filter objects or a filter expression in form of a formula.
which	A GRanges object specifying the region you want to get model from the TxDb object. For EnsDb : can be a GRanges object, an object extending AnnotationFilter , an AnnotationFilterList combining such filter objects or a filter expression in form of a formula.
weighted	If TRUE, weighted by simply add 1/cases matched to each model and if FALSE, simply add 1 to every case.
...	Extra arguments passed to qplot function. such as, offset which control the height of chevron.

Details

Internally we use `biovizBase::spliceSummary` for simple counting, but we encourage users to use their own robust way to make slicing summary and store it as GRangesList, then plot the summary by `qplot` function.

Value

A ggplot object.

Author(s)

Tengfei Yin

See Also

[qplot](#)

Examples

```
## Not run:
bamfile <- system.file("extdata", "SRR027894subRBM17.bam", package="biovizBase")
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
data(genesymbol)
exons <- exonsBy(txdb, by = "tx")
exons.rbm17 <- subsetByOverlaps(exons, genesymbol["RBM17"])
plotSpliceSum(bamfile, exons.rbm17)
plotSpliceSum(bamfile, exons.rbm17, weighted = FALSE, offset = 0.01)
plotSpliceSum(bamfile, txdb, which = genesymbol["RBM17"])
plotSpliceSum(bamfile, txdb, which = genesymbol["RBM17"], offset = 0.01)
plotSpliceSum(bamfile, txdb, which = genesymbol["RBM17"],
              show.label = TRUE,
              label.type = "count")

## End(Not run)
```

plotStackedOverview *Plot stacked overview*

Description

Plot stacked overview for genome with or without cytobands. It's a wrapper around layout_karyogram.

Usage

```
plotStackedOverview(obj, ..., xlab, ylab, main, geom = "rect",
                    cytobands = FALSE, rescale = TRUE,
                    rescale.range = c(0, 10))
plotKaryogram(obj, ..., xlab, ylab, main, geom = "rect",
               cytobands = FALSE, rescale = TRUE,
               rescale.range = c(0, 10))
```

Arguments

obj	a GRanges object, which could contain extra information about cytobands. If it's missing, will ask user to provide species information and download proper data set from UCSC. If you want an accurate genome mapping, please provide seqlengths with this GRanges object, otherwise it will emit a warning and use data space to estimate the chromosome space which is very rough.
...	arguments passed to graphic functions to control aesthetics. For example, if you use geom "point", you need to provide "y" in aes(), and if can also pass color, fill, size etc. to control graphics.
xlab	label for x
ylab	label for y
main	title for plot.
geom	geom plotted on the stacked layout. Default is "rect", which showing interval data as rectangles. It automatically figures out boundary so you don't have to provide information in aes, users could specify other supported geom works for data.frame.

cytobands	logical value. Default is FALSE. If TRUE, plotting cytobands, this require your data have arbitrary column as name and gieStain. the easiest way is to use getIdeogram to get your data. Notice for this function, when cytobands is TRUE, it will only plot cytobands without overlaying your data. If you really need to overlay extra data on cytobands, please plus layout_karyogram for that purpose.
rescale	logical value. Default is TRUE, which rescale your data into the rescale.range, this make sure your data will not be plotted outside the stacked overview box.
rescale.range	Numeric range of length 2. Default is (0, 10), because stacked layout draws a white background as chromosome space and this space is of height 10. We hide the y-axis since we don't need it for stacked overview. Sometime users may want to leave some margin for their data, they can use this arguments to control the rescale.

Details

Stacked overview is just a arbitrary layout for karyogram layout, it use facets seqnaems ~ . as default to stack the genome. For accurate mapping, you need to provide seqlengths information in your GRanges object. Otherwise, data space will be computed for stacked overview chromosome background, this is **_NOT_** the actual chromosome space!.

Value

A ggplot object.

Author(s)

Tengfei Yin

Examples

```
## Not run:
library(biovizBase)
data(hg19IdeogramCyto, package = "biovizBase")
library(GenomicRanges)

## you can also get ideogram by biovizBase::getIdeogram

## make shorter and clean labels
old.chrs <- seqnames(seqinfo(hg19IdeogramCyto))
new.chrs <- gsub("chr", "", old.chrs)
## lst <- as.list(new.chrs)
names(new.chrs) <- old.chrs
library(GenomeInfoDb) # for renameSeqlevels() and keepSeqlevels()
new.ideo <- renameSeqlevels(hg19IdeogramCyto, new.chrs)
new.ideo <- keepSeqlevels(new.ideo, c(as.character(1:22) , "X", "Y"))
new.ideo

## sample data
data(darned_hg19_subset500, package = "biovizBase")
idx <- is.na(values(darned_hg19_subset500)$exReg)
values(darned_hg19_subset500)$exReg[idx] <- "unknown"

## you need to add seqlengths for accruate mapping
```

```

chrnames <- unique(as.character(seqnames(darned_hg19_subset500)))
data(hg19Ideogram, package = "biovizBase")
seqlengths(darned_hg19_subset500) <- seqlengths(hg19Ideogram)[sort(chrnames)]

dn <- darned_hg19_subset500
values(dn)$score <- rnorm(length(dn))

## plotStackedOverview is a simple wrapper around this functions to
  create a stacked layout
plotStackedOverview(new.ideo, cytobands = TRUE)

plotStackedOverview(dn)
plotStackedOverview(dn, aes(color = exReg, fill = exReg))
## this will did the trick for you to rescale the space
plotStackedOverview(dn, aes(x = midpoint, y = score), geom = "line")
plotStackedOverview(dn, aes(x = midpoint, y = score), geom = "line", rescale.range = c(4, 6))
## no rescale
plotStackedOverview(dn, aes(x = midpoint, y = score), geom = "line", rescale = FALSE,
                    xlab = "xlab", ylab = "ylab", main = "main") + ylab("ylab")

## no object? will ask you for species and query the data on the fly
plotStackedOverview()
plotStackedOverview(cytobands = TRUE)

## End(Not run)

```

rescale

rescale ggplot object

Description

Rescale a numeric vector or ggplot object, could be used for static zoom-in in ggbio.

Usage

```

## S4 method for signature 'numeric'
rescale(x, to = c(0, 1),
        from = range(x, na.rm = TRUE))

## S4 method for signature 'ggplot'
rescale(x, xlim, ylim, sx = 1, sy = 1)
## S4 method for signature 'gg'
rescale(x, xlim, ylim, sx = 1, sy = 1)

```

Arguments

x	A numeric object or ggplot object to be rescaled.
to	For numeric object. it's a vector of two numeric values, specifying the range to be rescale.
from	Range of x.
xlim	For ggplot object. This specify the new limits on x-scale.

ylim	For ggplot object. This specify the new limits on y-scale.
sx	Scale fold for x-scale. Default is 1, no change.
sy	Scale fold for y-scale. Default is 1, no change.

Details

When `x` is numeric value, it's just call `scales::rescale`, please refer to the manual page to check more details. If `x` is ggplot object, it first try to estimate current `x` limits and `y` limits of the ggplot object, then rescale based on those information.

Value

Return the object of the same class as `x` after rescaling.

Author(s)

Tengfei Yin

Examples

```
library(ggbio)
head(mtcars)
range(mtcars$mpg)
p <- qplot(data = mtcars, x = mpg, y = disp, geom = "point")
p.new <- rescale(p, xlim = c(20, 25))
p.new
```

scale_fill_fold_change

scale color for fold change values

Description

In biology, lots of data are scaled to value around 0, and people like to show them as blue-white-red scale color, where negative value are blue, 0 is white and positive value is red, and they are scaled for continuous variables.

Usage

```
scale_fill_fold_change()
```

Value

a list.

Author(s)

Tengfei Yin

Examples

```
p1 <- autoplot(volcano - 150)
p1
p1 + scale_fill_fold_change()
```

scale_fill_giensa	<i>scale filled color to customized giensa color.</i>
-------------------	---

Description

scale filled color to customized giensa color.

Usage

```
scale_fill_giensa(fill = getOption("biovizBase")$cytobandColor)
```

Arguments

fill	a character vector to indicate colors, and names of vector mapped to gieStain name.
------	---

Value

a list.

Author(s)

Tengfei Yin

Examples

```
getOption("biovizBase")$cytobandColor
library(biovizBase)
data(hg19IdeogramCyto)
p1 <- autoplot(hg19IdeogramCyto, layout = "karyogram", aes(fill =
gieStain))
p1
p1 + scale_fill_giensa()
```

scale_x_sequnit	<i>scale x by unit</i>
-----------------	------------------------

Description

scale x by unit 'Mb','kb', 'bp'.

Usage

```
scale_x_sequnit(unit = c("Mb", "kb", "bp"), append = NULL)
```

Arguments

unit	unit to scale x. Default is Mb.
append	default NULL. If pass a character, it disalbe unit and arbitrarily append a text behind the original x scale numbers.

Value

'position_c'

Author(s)

Tengfei Yin

Examples

```
library(ggplot2)
p <- qplot(x = seq(1, to = 10000, length.out = 40), y = rnorm(40), geom
= "point")
## default mb
p + scale_x_sequnit()
p + scale_x_sequnit("kb")
p + scale_x_sequnit("bp")
```

stat_aggregate

Generates summaries on the specified windows

Description

Generates summaries on the specified windows

Usage

```
## S4 method for signature 'GRanges'
stat_aggregate(data, ..., xlab, ylab, main, by, FUN,
               maxgap=-1L, minoverlap=0L,
               type=c("any", "start", "end", "within", "equal"),
               select=c("all", "first", "last", "arbitrary"),
               y = NULL, window = NULL, facets = NULL,
               method = c("mean", "median", "max",
                          "min", "sum", "count", "identity"),
               geom = NULL)
```

Arguments

data	A GRanges or data.frame object.
...	Arguments passed to plot function. such as aes() and color.
xlab	Label for x
ylab	Label for y
main	Title for plot.
by	An object with 'start', 'end', and 'width' methods. Passed to aggregate.
FUN	The function, found via 'match.fun', to be applied to each window of 'x'. Passed to aggregate.

maxgap, minoverlap, type	Used in the internal call to <code>findOverlaps()</code> to detect overlaps. See <code>?findOverlaps</code> in the IRanges package for a description of these arguments.
select	It passed to <code>findOverlaps</code> . When select is "all" (the default), the results are returned as a Hits object. When select is "first", "last", or "arbitrary" the results are returned as an integer vector of length query containing the first, last, or arbitrary overlapping interval in subject, with NA indicating intervals that did not overlap any intervals in subject. If select is "all", a Hits object is returned. For all other select the return value depends on the drop argument. When select != "all" && !drop, an IntegerList is returned, where each element of the result corresponds to a space in query. When select != "all" && drop, an integer vector is returned containing indices that are offset to align with the unlisted query.
y	A character indicate the variable column for which aggregation is taken on, same as <code>aes(y =)</code> .
window	Integer value indicate window size.
facets	Faceting formula to use.
method	customized method for aggregating, if FUN is not provided.
geom	The geometric object to use display the data.

Value

A 'Layer'.

Author(s)

Tengfei Yin

Examples

```
library(GenomicRanges)
set.seed(1)
N <- 1000
## =====
## simulated GRanges
## =====
gr <- GRanges(seqnames =
  sample(c("chr1", "chr2", "chr3"),
    size = N, replace = TRUE),
  IRanges(
    start = sample(1:300, size = N, replace = TRUE),
    width = sample(70:75, size = N, replace = TRUE)),
  strand = sample(c("+", "-", "*"), size = N,
    replace = TRUE),
  value = rnorm(N, 10, 3), score = rnorm(N, 100, 30),
  sample = sample(c("Normal", "Tumor"),
    size = N, replace = TRUE),
  pair = sample(letters, size = N,
    replace = TRUE))

ggplot(gr) + stat_aggregate(mapping = aes(y = value))
```

```
## or
## ggplot(gr) + stat_aggregate(y = "value")
ggplot(gr) + stat_aggregate(mapping = aes(y = value), window = 36)
ggplot(gr) + stat_aggregate(mapping = aes(y = value), select = "first")
## Not run:
## no hits
ggplot(gr) + stat_aggregate(mapping = aes(y = value), select = "first", type = "within")

## End(Not run)
ggplot(gr) + stat_aggregate(window = 30, mapping = aes(y = value), fill = "gray40", geom = "bar")
ggplot(gr) + stat_aggregate(window = 100, fill = "gray40", mapping = aes(y = value),
                             method = "max", geom = "bar")

ggplot(gr) + stat_aggregate(mapping = aes(y = value), geom = "boxplot")
ggplot(gr) + stat_aggregate(mapping = aes(y = value), geom = "boxplot", window = 60)
## now facets need to take place inside stat_* geom_* for an accurate computation
ggplot(gr) + stat_aggregate(mapping = aes(y = value), geom = "boxplot", window = 30,
                             facets = sample ~ seqnames)

## FIXME:
## autoplot(gr, stat = "aggregate", aes(y = value), window = 36)
## autoplot(gr, stat = "aggregate", geom = "boxplot", aes(y = value), window = 36)
```

stat_bin

Binning method

Description

Binning method especially for Rle and RleList, for data.frame it's just calling ggplot2::stat_bin.

Usage

```
## S4 method for signature 'ANY'
stat_bin(data, ...)

## S4 method for signature 'Rle'
stat_bin(data, ..., binwidth, nbin = 30,
          xlab, ylab, main, geom = c("bar", "heatmap"),
          type = c("viewSums", "viewMins",
                  "viewMaxs", "viewMeans"))

## S4 method for signature 'RleList'
stat_bin(data, ..., binwidth, nbin = 30,
          xlab, ylab, main,
          indName = "sample",
          geom = c("bar", "heatmap"),
          type = c("viewSums", "viewMins",
                  "viewMaxs", "viewMeans"))
```

Arguments

data	Typically a data.frame or Rle or RleList object.
...	arguments passed to aesthetics mapping.
binwidth	width of the bins.

nbin	number of bins.
xlab	x label.
ylab	y label.
main	title.
indName	when faceted by a RleList, name used for labeling faceted factor. Default is 'sample'.
geom	geometric types.
type	statistical summary method used within bins, shown as bar height or heatmap colors.

Value

a ggplot object.

Author(s)

Tengfei Yin

Examples

```
library(IRanges)
lambda <- c(rep(0.001, 4500), seq(0.001, 10, length = 500),
            seq(10, 0.001, length = 500))
xVector <- rpois(1e4, lambda)
xRle <- Rle(xVector)
xRleList <- RleList(xRle, 2L * xRle)

ggplot() + stat_bin(xRle)
ggplot(xRle) + stat_bin()
ggplot(xRle) + stat_bin(nbin = 100)
ggplot(xRle) + stat_bin(binwidth = 200)

p1 <- ggplot(xRle) + stat_bin(type = "viewMeans")
p2 <- ggplot(xRle) + stat_bin(type = "viewSums")
## y scale are different.
tracks(viewMeans = p1, viewSums = p2)

ggplot(xRle) + stat_bin(geom = "heatmap")
ggplot(xRle) + stat_bin(nbin = 100, geom = "heatmap")
ggplot(xRle) + stat_bin(binwidth = 200, geom = "heatmap")

## for RleList
ggplot(xRleList) + stat_bin()
ggplot(xRleList) + stat_bin(nbin = 100)
ggplot(xRleList) + stat_bin(binwidth = 200)

p1 <- ggplot(xRleList) + stat_bin(type = "viewMeans")
p2 <- ggplot(xRleList) + stat_bin(type = "viewSums")
## y scale are different.
tracks(viewMeans = p1, viewSums = p2)

ggplot(xRleList) + stat_bin(geom = "heatmap")
ggplot(xRleList) + stat_bin(nbin = 100, geom = "heatmap")
ggplot(xRleList) + stat_bin(binwidth = 200, geom = "heatmap")
```

stat_coverage	<i>Calculate coverage</i>
---------------	---------------------------

Description

Calculate coverage.

Usage

```
# for GRanges
## S4 method for signature 'GRanges'
stat_coverage(data, ..., xlim, xlab, ylab, main,
              facets = NULL, geom = NULL)
# for GRangesList
## S4 method for signature 'GRangesList'
stat_coverage(data, ..., xlim, xlab, ylab, main,
              facets = NULL, geom = NULL)

# for Bamfile
## S4 method for signature 'BamFile'
stat_coverage(data, ..., maxBinSize = 2^14,
              xlim, which, xlab, ylab,
              main, facets = NULL, geom = NULL,
              method = c("estimate", "raw"),
              space.skip = 0.1, coord = c("linear", "genome"))
```

Arguments

data	A GRanges or data.frame object.
...	Extra parameters such as aes() passed to geom_rect, geom_alignment, or geom_segment.
xlim	Limits for x.
xlab	Label for x
ylab	Label for y
main	Title for plot.
facets	Faceting formula to use.
geom	The geometric object to use display the data.
maxBinSize	maxBinSize.
method	'estimate' for parsing estimated coverage(fast), 'raw' is slow and parse the accurate coverage.
which	GRanges which defines region to subset the results.
space.skip	used for coordinate genome, skip between chromosomes.
coord	coordinate system.

Value

A 'Layer'.

Author(s)

Tengfei Yin

Examples

```

library(ggbio)
## =====
##  simulated GRanges
## =====
set.seed(1)
N <- 1000
library(GenomicRanges)

gr <- GRanges(seqnames =
  sample(c("chr1", "chr2", "chr3"),
    size = N, replace = TRUE),
  IRanges(
    start = sample(1:300, size = N, replace = TRUE),
    width = sample(70:75, size = N, replace = TRUE)),
  strand = sample(c("+", "-", "*"), size = N,
    replace = TRUE),
  value = rnorm(N, 10, 3), score = rnorm(N, 100, 30),
  sample = sample(c("Normal", "Tumor"),
    size = N, replace = TRUE),
  pair = sample(letters, size = N,
    replace = TRUE))

ggplot(gr) + stat_coverage()
ggplot() + stat_coverage(gr)

ggplot(gr) + stat_coverage(geom = "point")
ggplot(gr) + stat_coverage(geom = "area")
ggplot(gr) + stat_coverage(mapping = aes(y = ..coverage..), geom = "bar")

ggplot(gr) + stat_coverage(mapping = aes(y = ..coverage..)) + geom_point()

## for bam file
## TBD

```

stat_gene

*Calculate gene structure***Description**

Calculate gene structure.

Usage

```

## S4 method for signature 'TxDb'
stat_gene(data, ...)

```

Arguments

data	A GRanges or data.frame object.
...	Extra parameters such as aes() passed to geom_alignment.

Value

A 'Layer'.

Author(s)

Tengfei Yin

See Also

[geom_alignment](#)

Examples

```
## Not run:
## loading package
## Deprecated
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
data(genesymbol, package = "biovizBase")
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene

## made a track comparing full/reduce stat.
p1 <- ggplot() + geom_alignment(txdb, which = genesymbol["RBM17"])
p1 <- ggplot() + stat_gene(txdb, which = genesymbol["RBM17"])
## or
p1 <- ggplot(txdb) + stat_gene(which = genesymbol["RBM17"])

p1 <- ggplot(txdb) + stat_gene(which = genesymbol["RBM17"])
p2 <- ggplot(txdb) + stat_gene(which = genesymbol["RBM17"], stat =
"reduce")
p2 <- ggplot(txdb) + stat_gene(which = genesymbol["RBM17"], stat = "reduce")
## ggplot(txdb) + geom_alignment(which = genesymbol["RBM17"]) + stat_reduce()
## ggplot(txdb) + geom_alignment(which = genesymbol["RBM17"])
tracks(full = p1, reduce = p2, heights = c(3, 1))

## change y labels
ggplot(txdb) + stat_gene(which = genesymbol["RBM17"], names.expr =
"tx_id::gene_id")

## End(Not run)
```

stat_identity

Transform the data to a data.frame and for multiple geoms.

Description

Transform the data to a suitable data.frame and then one could use multiple geom or even stat to re-plot the data.

Usage

```
## S4 method for signature 'ANY'
stat_identity(data, ...)

## S4 method for signature 'GRanges'
stat_identity(data, ..., geom = NULL)

## S4 method for signature 'Rle'
stat_identity(data, ..., xlab, ylab, main, geom = NULL)

## S4 method for signature 'RleList'
stat_identity(data, ..., xlab, ylab, main,
              geom = NULL, indName = "sample")
```

Arguments

<code>data</code>	Typically a <code>GRanges</code> or <code>data.frame</code> object.
<code>...</code>	Extra parameters such as <code>aes()</code> passed to <code>geom_rect</code> , <code>geom_alignment</code> , or <code>geom_segment</code> .
<code>geom</code>	The geometric object to use display the data.
<code>xlab</code>	x label.
<code>ylab</code>	y label.
<code>main</code>	title of graphic..
<code>indName</code>	sample name.

Value

A 'Layer'.

Author(s)

Tengfei Yin

Examples

```
## load
set.seed(1)
N <- 50

require(GenomicRanges)
## simul
## =====
## simulated GRanges
## =====
gr <- GRanges(seqnames =
  sample(c("chr1", "chr2", "chr3"),
    size = N, replace = TRUE),
  IRanges(
    start = sample(1:300, size = N, replace = TRUE),
    width = sample(70:75, size = N, replace = TRUE)),
  strand = sample(c("+", "-", "*"), size = N,
    replace = TRUE),
  value = rnorm(N, 10, 3), score = rnorm(N, 100, 30),
  sample = sample(c("Normal", "Tumor"),
```

```

      size = N, replace = TRUE),
    pair = sample(letters, size = N,
      replace = TRUE))

## geom_point_start
ggplot() + stat_identity(gr, mapping = aes(x = start, y = value), geom = "point")
## or more formal
ggplot(gr) + stat_identity(mapping = aes(x = start, y = value), geom = "point")

## geom_point_midpoint
ggplot(gr) + stat_identity(mapping = aes(x = midpoint, y = value), geom = "point")

## geom_rect_all
ggplot(gr) + stat_identity(mapping = aes(xmin = start, xmax = end,
                                          ymin = value - 0.5, ymax = value + 0.5),
                          geom = "rect")

## geom_rect_y
ggplot(gr) + stat_identity(mapping = aes(y = value), geom = "rect")

## geom_line
ggplot(gr) + stat_identity(mapping = aes(x = start, y = value), geom = "line")

## geom_segment
ggplot(gr) + stat_identity(mapping = aes(y = value), geom = "segment")

## Rle/RleList
library(IRanges)
lambda <- c(rep(0.001, 4500), seq(0.001, 10, length = 500),
            seq(10, 0.001, length = 500))
xVector <- rpois(1e4, lambda)
xRle <- Rle(xVector)
xRleList <- RleList(xRle, 2L * xRle)

ggplot(xRle) + stat_identity(geom = "point")
ggplot(xRleList) + stat_identity(geom = "point")

```

stat_mismatch

Calculate mismatch summary

Description

Calculate mismatch summary

Usage

```

## for GRanges
## S4 method for signature 'GRanges'
stat_mismatch(data, ..., bsgenome,
              xlab, ylab, main,
              geom = c("segment", "bar"),
              show.coverage = TRUE)

## for BamFile
## S4 method for signature 'BamFile'

```

```
stat_mismatch(data, ..., bsgenome, which,
               xlab, ylab, main,
               geom = c("segment", "bar"),
               show.coverage = TRUE)
```

Arguments

data	A GRanges or BamFile object.
...	Extra parameters such as aes() passed to geom_rect, geom_alignment, or geom_segment.
bsgenome	BSgenome object.
which	GRanges object to subset the data.
xlab	Label for x
ylab	Label for y
main	Title for plot.
geom	The geometric object to use display the data.
show.coverage	whether to show coverage as background or not.

Value

A 'Layer'.

Author(s)

Tengfei Yin

stat_reduce	<i>Reduce an object.</i>
-------------	--------------------------

Description

Reduce GRanges, IRanges or TxDb object.

Usage

```
## S4 method for signature 'GRanges'
stat_reduce(data, ...,
            xlab, ylab, main,
            drop.empty.ranges = FALSE,
            min.gapwidth = 1L,
            facets = NULL, geom = NULL)

## S4 method for signature 'IRanges'
stat_reduce(data, ...,
            xlab, ylab, main,
            drop.empty.ranges = FALSE,
            min.gapwidth = 1L,
            with.inframe.attrib=FALSE,
            facets = NULL, geom = NULL)

## S4 method for signature 'TxDbOREnsDb'
stat_reduce(data, ...)
```

Arguments

<code>data</code>	GRanges, IRanges or TxDb object.
<code>...</code>	passed to aesthetics mapping.
<code>xlab</code>	x label.
<code>ylab</code>	y label.
<code>main</code>	title.
<code>drop.empty.ranges</code>	pass to reduce function.
<code>min.gapwidth</code>	pass to reduce function.
<code>with.inframe.attrib</code>	pass to reduce function.
<code>facets</code>	pass to reduce function.
<code>geom</code>	geometric type.

Value

a ggplot object.

Author(s)

Tengfei Yin

See Also

[reduce](#).

Examples

```
set.seed(1)
N <- 1000
library(GenomicRanges)

gr <- GRanges(seqnames =
  sample(c("chr1", "chr2", "chr3"),
    size = N, replace = TRUE),
  IRanges(
    start = sample(1:300, size = N, replace = TRUE),
    width = sample(70:75, size = N, replace = TRUE)),
  strand = sample(c("+", "-", "*"), size = N,
    replace = TRUE),
  value = rnorm(N, 10, 3), score = rnorm(N, 100, 30),
  sample = sample(c("Normal", "Tumor"),
    size = N, replace = TRUE),
  pair = sample(letters, size = N,
    replace = TRUE))

ggplot(gr) + stat_reduce()
autoplot(gr, stat = "reduce")
strand(gr) <- "*"
ggplot(gr) + stat_reduce()

library(TxDb.Hsapiens.UCSC.hg19.knownGene)
```



```
data(genesymbol, package = "biovizBase")
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
## made a track comparing full/reduce stat.
ggplot(txdb) + stat_reduce(which = genesymbol["RBM17"])
```

stat_slice

Slice Rle/RleList to view them as bar or heatmap.

Description

Slice Rle/RleList to different view by set lower or other parameters, then view summary for all those viewed region.

Usage

```
## S4 method for signature 'Rle'
stat_slice(data, ...,
            xlab, ylab, main,
            na.rm = FALSE,
            geom = NULL,
            lower=-Inf, upper=Inf,
            includeLower=TRUE, includeUpper=TRUE,
            rangesOnly = FALSE,
            type = c("viewSums", "viewMins",
                    "viewMaxs", "viewMeans"))

## S4 method for signature 'RleList'
stat_slice(data, ...,
            xlab, ylab, main,
            indName = "sample",
            na.rm = FALSE,
            geom = NULL,
            lower=-Inf, upper=Inf,
            includeLower=TRUE, includeUpper=TRUE,
            rangesOnly = FALSE,
            type = c("viewSums", "viewMins",
                    "viewMaxs", "viewMeans"))
```

Arguments

data	a data.frame or Rle or RleList object.
...	arguments passed to aesthetics mapping.
xlab	x label.
ylab	y label.
main	title.
indName	when faceted by a RleList, name used for labeling faceted factor. Default is 'sample'.
geom	geometric types.

type	statistical summary method used within bins, shown as bar height or heatmap colors.
na.rm	logical value, default FALSE, passed to function like viewMaxs for statistical summary computation.
lower	passed to slice .
upper	passed to slice .
includeLower	passed to slice .
includeUpper	passed to slice .
rangesOnly	passed to slice .

Value

a ggplot object.

Author(s)

Tengfei Yin

See Also

[slice](#)

Examples

```
library(IRanges)
lambda <- c(rep(0.001, 4500), seq(0.001, 10, length = 500),
            seq(10, 0.001, length = 500))
xVector <- rpois(1e4, lambda)
xRle <- Rle(xVector)
xRleList <- RleList(xRle, 2L * xRle)

ggplot(xRle) + stat_slice(lower = 5)
ggplot(xRle) + stat_slice(lower = 5, geom = "bar")
ggplot(xRle) + stat_slice(lower = 5, geom = "heatmap")

p1 <- ggplot(xRle) + stat_slice(type = "viewMeans", lower = 5,
                              geom = "bar")
p2 <- ggplot(xRle) + stat_slice(type = "viewSums", lower = 5,
                              geom = "bar")

## y scale are different.
tracks(viewMeans = p1, viewSums = p2)

ggplot(xRleList) + stat_slice(lower = 5)
ggplot(xRleList) + stat_slice(lower = 5, geom = "bar")
ggplot(xRleList) + stat_slice(lower = 5, geom = "heatmap")

p1 <- ggplot(xRleList) + stat_slice(type = "viewMeans", lower = 5,
                              geom = "bar")
p2 <- ggplot(xRleList) + stat_slice(type = "viewSums", lower = 5,
                              geom = "bar")

## y scale are different.
tracks(viewMeans = p1, viewSums = p2)
```

stat_stepping	<i>Calculate stepping levels</i>
---------------	----------------------------------

Description

Calculate stepping levels.

Usage

```
## S4 method for signature 'GRanges'
stat_stepping(data, ..., xlab, ylab, main,
               facets = NULL,
               geom = c("rect", "alignment", "segment"))
```

Arguments

data	A GRanges or data.frame object.
...	Extra parameters such as aes() passed to geom_rect, geom_alignment, or geom_segment.
xlab	Label for x
ylab	Label for y
main	Title for plot.
facets	Faceting formula to use.
geom	The geometric object used to display the data. For 'stepping', could be one of 'rect', 'alignment', 'segment'.

Value

A 'Layer'.

Author(s)

Tengfei Yin

Examples

```
set.seed(1)
N <- 50

require(GenomicRanges)
## simul
## =====
## simulated GRanges
## =====
gr <- GRanges(seqnames =
               sample(c("chr1", "chr2", "chr3"),
                     size = N, replace = TRUE),
               IRanges(
                 start = sample(1:300, size = N, replace = TRUE),
                 width = sample(70:75, size = N, replace = TRUE)),
               strand = sample(c("+", "-", "*"), size = N,
                              replace = TRUE),
```

```

value = rnorm(N, 10, 3), score = rnorm(N, 100, 30),
sample = sample(c("Normal", "Tumor"),
  size = N, replace = TRUE),
pair = sample(letters, size = N,
  replace = TRUE))

## default
ggplot(gr) + stat_stepping()
## or
ggplot() + stat_stepping(gr)

## facet_aes
ggplot(gr) + stat_stepping(mapping = aes(color = strand, fill = strand),
  facets = sample ~ seqnames)

## geom_segment
ggplot(gr) + stat_stepping(mapping = aes(color = strand),
  geom = "segment", xlab = "Genomic coord", ylab = "y", main = "hello")

## geom_alignment
## ggplot(gr) + stat_stepping(geom = "alignment")

## geom_alignment_group
## ggplot(gr) + stat_stepping(mapping = aes(group = pair),geom = "alignment")

```

stat_table	<i>Tabulate a GRanges object</i>
------------	----------------------------------

Description

Tabulate a GRanges object

Usage

```

## S4 method for signature 'GRanges'
stat_table(data, ..., xlab, ylab, main,
  geom = NULL, stat = NULL)
## S4 method for signature 'GRangesList'
stat_table(data, ..., xlab, ylab, main,
  facets = NULL, geom = NULL)

```

Arguments

data	A GRanges or data.frame object.
...	Extra parameters such as aes() passed to geom_rect, geom_alignment, or geom_segment.
xlab	Label for x
ylab	Label for y
main	Title for plot.
facets	Faceting formula to use.
geom	The geometric object to use display the data.
stat	The geometric object to use display the data.

Value

A 'Layer'.

Author(s)

Tengfei Yin

Examples

```
## load
set.seed(1)
N <- 100
require(ggbio)
require(GenomicRanges)
## simul
## =====
## simulated GRanges
## =====
gr <- GRanges(seqnames =
  sample(c("chr1", "chr2", "chr3"),
    size = N, replace = TRUE),
  IRanges(
    start = sample(1:300, size = N, replace = TRUE),
    width = sample(70:75, size = N, replace = TRUE)),
  strand = sample(c("+", "-", "*"), size = N,
    replace = TRUE),
  value = rnorm(N, 10, 3), score = rnorm(N, 100, 30),
  sample = sample(c("Normal", "Tumor"),
    size = N, replace = TRUE),
  pair = sample(letters, size = N,
    replace = TRUE))

gr <- c(gr[seqnames(gr) == "chr1"][sample(1:10, size = 1e4, replace = TRUE)],gr)

## default
ggplot(gr) + stat_table()
ggplot(gr) + stat_table(geom = "segment", mapping = aes(y = ..score.., color = ..score..))
ggplot(gr) + stat_table(mapping = aes(color = score))
```

theme

theme in ggbio

Description

Theme defined in ggbio for plot or tracks.

Usage

```
theme_null()
theme_noexpand()
theme_alignment(ylabel = FALSE, base_size = 12, base_family = "",
  axis = TRUE, border = TRUE, grid = TRUE)
theme_pack_panels(strip.bg = FALSE, strip.text.y = TRUE)
```

```

theme_clear(grid.y = FALSE, grid.x.minor = FALSE, grid.x.major = FALSE,
            panel.background.fill = "white", panel.border.color = NA,
            axis.ticks.x = FALSE, axis.ticks.y = TRUE, grid.color = "gray95",
            axis.line.color = "gray80")
theme_tracks_sunset(bg = "#fffedb", alpha = 1, ...)
theme_genome()

```

Arguments

alpha	alpha blending from 0(transparent) to 1(solid).
axis	logical value, show axis or not.
axis.line.color	color for axis line .
axis.ticks.x	show x ticks or not.
axis.ticks.y	show y ticks or not.
base_family	family for font.
base_size	size for font.
bg	background color for tracks.
border	logical value, show border or not.
grid	logical value, show background grid or not.
grid.color	grid line color.
grid.x.major	show x major grid line or not.
grid.x.minor	show x minor grid line or not.
grid.y	show y grid or not.
panel.background.fill	panel background fill color.
panel.border.color	panel border color.
strip.bg	if strip background is removed.
strip.text.y	if strip text is removed.
ylabel	logical value. Show labels or not.
...	passed to theme_clear.

Details

Themes speciall designed for tracks, are named following naming schema `theme_tracks_*`

Value

Return a theme.

Author(s)

Tengfei Yin

Examples

```
## load
library(ggbio)
p <- qplot(data = mtcars, x = mpg, y = wt, facets = cyl ~ .)
p + theme_null()
p + theme_clear()
p + theme_pack_panels()
p + theme_alignment()
p1 <- qplot(data = mtcars, x = mpg, y = wt)
tracks(p1 = p, p2 = p1)
tracks(p1 = p, p2 = p1) + theme_tracks_sunset()
```

Tracked	<i>Tracked class</i>
---------	----------------------

Description

Create a tracked object, designed for tracks function.

Usage

```
Tracked(mutable = TRUE, fixed = FALSE, labeled = TRUE,
        hasAxis = FALSE, bgColor = "white", height = unit(1, "null"))
```

Arguments

mutable	logical value, default TRUE. To control whether a track is updatable by applying + on it.
fixed	logical value, default FALSE. To control whether the scale response to a xlim change or not.
labeled	logical value, default TRUE. To control whether to label it all not.
hasAxis	logical value, default FALSE. To control whether to show axis for that track or not.
bgColor	character to control background color of a track.
height	unit, to control track height.

Value

a Tracked object.

Author(s)

Tengfei Yin

tracks

*Tracks for genomic graphics***Description**

`tracks` is a convenient constructor for binding graphics as tracks. You don't have to worry about adjusting different graphics, `tracks` did that for you. It's NOT just limited to bind genomic tracks, you can use this function to bind any tracks with the same definition of x axis, for example, sets of time series plots you made.

Tracks view is most common way to viewing genome features and annotation data and widely used by most genome browsers. Our assumption is that, most graphics you made with `ggbio` or by yourself using `ggplot2`, are almost always sitting on the genomic coordinates or the same x axis. And to compare annotation information along with genome features, we need to align those plots on exactly the same x axis in order to form your hypothesis. This function leaves you the flexibility to construct each tracks separately with worrying your alignments later.

Usage

```
tracks(..., heights, xlim, xlab = NULL, main = NULL,
       title = NULL, theme = NULL,
       track.plot.color = NULL,
       track.bg.color = NULL,
       main.height = unit(1.5, "lines"),
       scale.height = unit(1, "lines"),
       xlab.height = unit(1.5, "lines"),
       padding = unit(-1, "lines"),
       label.bg.color = "white",
       label.bg.fill = "gray80",
       label.text.color = "black",
       label.text.cex = 1,
       label.text.angle = 90,
       label.width = unit(2.5, "lines"))
```

Arguments

<code>...</code>	plots of class <code>ggplot</code> , generated from <code>ggplot2</code> or <code>ggbio</code> .
<code>heights</code>	numeric vector of the same length of passed graphic object to indicate the ratio of each track.
<code>xlim</code>	limits on x. could be IRanges , GRanges , numeric value
<code>xlab</code>	label for x axis.
<code>main</code>	title for the tracks.
<code>title</code>	title for the tracks, alias like <code>main</code> .
<code>theme</code>	theme object used for building tracks, this will set to default, which could be reseted later.
<code>track.plot.color</code>	Vector of characters of length 1 or the same length of passed plots, background color for each track, default is white.
<code>track.bg.color</code>	background color for the whole tracks.

<code>main.height</code>	unit. Height to control the title track height.
<code>scale.height</code>	unit. Height to control the scale track height.
<code>xlab.height</code>	unit. Height to control the xlab track height.
<code>padding</code>	single numeric value or unit, if numeric value, the unit would be "lines" by default.
<code>label.bg.color</code>	track labeling background rectangle border color.
<code>label.bg.fill</code>	track labeling background fill color.
<code>label.text.color</code>	track labeling text color.
<code>label.text.cex</code>	track labeling text size.
<code>label.text.angle</code>	angle to rotate the track labels.
<code>label.width</code>	track labeling size.

Details

tracks did following modification for passed plots.

- remove x-axis, ticks, xlab and title for each track and add scales at bottom. We suppose a new xlab and title would be provided by the tracks function for the whole tracks, but we still keep individual's y axis.
- align x-scale limits to make sure every plots sitting on exactly the same x scale.
- squeezing plots together to some extent.
- labeling tracks if names are provided, please check utilities section about labeled method.
- return a track object. This would allow many features introduced in this manual.

Value

A Tracks object.

Track class

constructor tracks will return a Tracks object, which has following slots.

`grobs` a `ggplotGrobList` object contains a list of `ggplot` object, which is our passed graphics.

`backup` a backup of all the slots for holding the original tracks, so users could edit it and reset it back at any time later, and `backup` method will reset the backedup copy.

`ylim` y limits for each plot.

`labeled` vector of logical value indicates whether a track is labeled or not, for labeled attributes please check utilities section.

`mutable` vector of logical value indicates whether a track is mutable for theme editing or not, for mutable attributes please check utilities section.

`hasAxis` vector of logical value indicates whether a track has axis or not, for `hasAxis` attributes please check utilities section.

`heights`, `xlim`, `xlab`, `main`, `title`, `theme`, `fixed`, `track.plot.color`, `track.bg.color`, `main.height`, `scale`. those slots are described in arguments section for constructor.

Utilities

Please check examples for usage.

`summary(object)` summary information about tracks object.

`fixed(x)`, `fixed(x) <- value` `x` is the ggplot object, this controls if a track has a fixed x scale or not, if the fixed attributes is TRUE, then when you pass this plot to a tracks, this plot won't be re-aligned with other tracks and will keep the original x-axis, this allow you to pass some plot like ideogram. `fixed` function will return a logical value

`labeled(x)`, `labeled(x) <- value` `x` is the ggplot object, if you pass named graphics into tracks, it will create the labels on the left for you. Several ways supported to name it. You can pass a list of graphics with names. Or you can use `tracks('name1' = p1, 'name 2' = p2, ...)` with quotes for complicated words or simply `tracks(part1 = p1, part = p2, ...)`.

`mutable(x)`, `mutable(x) <- value` `x` is the ggplot object, this controls whether a plot in the tracks mutable to theme changing or not, when you use `+` method for Tracks object, add-on edit will only be applied to the the mutable plots.

`bgColor(x)`, `bgColor(x) <- value` `x` is the ggplot object, this change the background color for single plot shown in the tracks.

`xlim(x)`, `xlim(x) <- value` when `x` is the numeric value, it calls `ggplot2::coord_cartesian(xlim = ...)` method, we doesn't use `ggplot2::xlim()` for the reason it will cut data outside the range, and we believe the best behavior would be zoom-in/out like most browser. when `x` is [IRanges](#), [GRanges](#), it get the range and passed to `ggplot2::coord_cartesian` function.

when `x` is Tracks object, `xlim(x)` will return `x` limits for that tracks. `xlim(x) <- value` replace method only works for Tracks object. `value` could be numeric, [IRanges](#), [GRanges](#) object. This will change the `x` limits associated with tracks.

`+ xlim(obj):obj` is the numeric range, or [IRanges](#), [GRanges](#) object.

`+ coord_cartesian()`: please read manual in `ggplot2`, this controls both `xlim` and `ylim`, only accept numerical range.

`+` The most nice features about [Tracks](#) object is the one inherited from `ggplot2`'s components additive features, with `+` method you can use any theme object and utilities in `ggplot2` package, to add them on a [Tracks](#) object, for example, if `x` is our [Tracks](#) object, `x + theme` would apply theme to any plots in the tracks except those are immutable.

`as(x, "grob")` Coerces a Tracks object to a grob for embedding in a larger figure.

Backup and reset

`reset(obj)` `obj` is the Tracks object, this reset the tracks back to original or backuped version.

`backup(obj)` `obj` is the Tracks object, this clear previous backup and use current setting for a new backup.

Author(s)

Tengfei Yin

See Also

[align.plots](#)

Examples

```

## make a simulated time series data set
df1 <- data.frame(time = 1:100, score = sin((1:100)/20)*10)
p1 <- qplot(data = df1, x = time, y = score, geom = "line")
df2 <- data.frame(time = 30:120, score = sin((30:120)/20)*10, value = rnorm(120-30 + 1))
p2 <- ggplot(data = df2, aes(x = time, y = score)) +
  geom_line() + geom_point(size = 4, aes(color = value))
## check p2
p1
## check p2
p2

## binding
tracks(p1, p2)

## or
tk1 <- tracks(p1, p2)
tk1

## combine
c(tk1, tk1)
tk1 + tk1

cbind(tk1, tk1)
rbind(tk1, tk1) ## different with c()!
library(grid)
x <- as(tk1, "grob")
grid.draw(cbind(x, x))

## labeling: default labeling a named graphic
## simply pass a name with it
tracks(time1 = p1, time2 = p2)
## or pass a named list with it
lst <- list(time1 = p1, time2 = p2)
tracks(lst)

## more complicated case please use quotes
tracks(time1 = p1, "second time" = p2)

## set heights
tracks(time1 = p1, time2 = p2, heights = c(1, 3))

## if you want to disable label arbitrarily
## default label is always TRUE
labeled(p2)
labeled(p2) <- FALSE
## set labeled to FALSE, remove label even the plot has a name
tracks(time1 = p1, time2 = p2)
labeled(p2) <- TRUE

## fix a plot, not synchronize with other plots
p3 <- p1
## default is always FALSE
fixed(p3)
## set to TRUE

```

```

fixed(p3) <- TRUE
fixed(p3)

tracks(time1 = p1, time2 = p2, "time3(fixed)" = p3)

fixed(p3) <- FALSE
## otherwise you could run

## control axis
hasAxis(p1)
hasAxis(p1) <- TRUE
# ready for weird looking
tracks(time1 = p1, time2 = p2)
# set it back
hasAxis(p1) <- FALSE

## mutable
mutable(p1)
tracks(time1 = p1, time2 = p2) + theme_bw()
mutable(p1) <- FALSE
# mutable for "+" method
tracks(time1 = p1, time2 = p2) + theme_bw()
mutable(p1) <- TRUE

## bgColor
bgColor(p1)
tracks(time1 = p1, time2 = p2)
bgColor(p1) <- "brown"
# mutable for "+" method
tracks(time1 = p1, time2 = p2)
# set it back
bgColor(p1) <- "white"

## apply a theme to each track
tks <- tracks(time1 = p1, time2 = p2) + theme_bw()
tks
reset(tks)

## store it with tracks
tks <- tracks(time1 = p1, time2 = p2, theme = theme_bw())
tks
tks <- tks + theme_gray()
tks
## reset will be introduced later
reset(tks)

## apply a pre-defined theme for tracks!
tracks(time1 = p1, time2 = p2) + theme_tracks_sunset()
tracks(p1, p2) + theme_tracks_sunset()

## change limits
tracks(time1 = p1, time2 = p2) + xlim(c(1, 40))
tracks(time1 = p1, time2 = p2) + xlim(1, 40)

```

```

tracks(time1 = p1, time2 = p2) + coord_cartesian(xlim = c(1, 40))
# change y
tracks(time1 = p1, time2 = p2) + xlim(1, 40) + ylim(0, 10)
library(GenomicRanges)
gr <- GRanges("chr", IRanges(1, 40))
# GRanges
tracks(time1 = p1, time2 = p2) + xlim(gr)
# IRanges
tracks(time1 = p1, time2 = p2) + xlim(ranges(gr))
tks <- tracks(time1 = p1, time2 = p2)
xlim(tks)
xlim(tks) <- c(1, 35)
xlim(tks) <- gr
xlim(tks) <- ranges(gr)

## xlab, title
tracks(time1 = p1, time2 = p2, xlab = "time")
tracks(time1 = p1, time2 = p2, main = "title")
tracks(time1 = p1, time2 = p2, title = "title")
tracks(time1 = p1, time2 = p2, xlab = "time", title = "title") + theme_tracks_sunset()

## backup and restore
tks <- tracks(time1 = p1, time2 = p2)
tks
tks <- tks + xlim(1, 40)
tks
reset(tks)
tks <- tks + xlim(1, 40)
tks
tks <- backup(tks)
tks <- tks + theme_bw()
tks
reset(tks)

## padding(need to be fixed for more delicate control)
tracks(time1 = p1, time2 = p2, padding = 2)

## track color
tracks(time1 = p1, time2 = p2, track.bg.color = "yellow")
tracks(time1 = p1, time2 = p2, track.plot.color = c("yellow", "brown"))

```

zoom

Simple navigation for ggbio object.

Description

A set of simple navigation API apply to ggbio object, let you move along the genome and zoom in/out.

Usage

```
zoom(fac = 1/2)
```

```
zoom_in(fac = 1/2)
zoom_out(fac = 2)
nextView(unit = c("view", "gene", "exon", "utr"))
prevView(unit = c("view", "gene", "exon", "utr"))
```

Arguments

fac	numeric value to indicate zoom factor, multiple of current view width. If it's smaller than 1, then it's zoom-in operation; if it's bigger than 1, then it's zoom-out operation.
unit	only support 'view' unit now.

Details

zoom_in and zoom_out are just simple wrapper around zoom function.

For more convenient, gene features based jumping we will support it in the future.

Value

A special class of navigation.

Author(s)

Tengfei Yin

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