

Package ‘VarCon’

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Type Package

Title VarCon: an R package for retrieving neighboring nucleotides of an SNV

Version 1.18.0

Description VarCon is an R package which converts the positional information from the annotation of a single nucleotide variation (SNV) (either referring to the coding sequence or the reference genomic sequence). It retrieves the genomic reference sequence around the position of the single nucleotide variation. To assess, whether the SNV could potentially influence binding of splicing regulatory proteins VarCon calculates the HEXplorer score as an estimation. Besides, VarCon additionally reports splice site strengths of splice sites within the retrieved genomic sequence and any changes due to the SNV.

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Encoding UTF-8

LazyData true

VignetteBuilder knitr

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Depends Biostrings, BSgenome, GenomicRanges, R (>= 4.1)

Suggests testthat, knitr, rmarkdown

biocViews FunctionalGenomics, AlternativeSplicing

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calculateHZEIperNT	<i>Generates table with HZEI scores per nucleotide of a sequence.</i>
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Description

This function generates a table with HZEI scores per index nucleotide.

Usage

```
calculateHZEIperNT(seq)
```

Arguments

seq	Nucleotide sequence longer than 11nt and only containing bases "A", "G", "C", "T".
-----	--

Value

Dataframe with HZEI value per index position.

Examples

```
calculateHZEIperNT("TTCCAAACGAACTTTGTAGGGA")
```

 calculateMaxEntScanScore

Calculate MaxEntScan score of a splice site sequence

Description

This function calculates the MaxEntScan score of either splice donor or acceptor sequences.

Usage

```
calculateMaxEntScanScore(seqVector, ssType)
```

Arguments

seqVector	Character vector of nucleotide sequence of a splice site sequences. SA sequences should be 23nt long (20 intronic, 3 exonic) and splice donor sequences should be 9nt long (3 exonic, 6 intronic) only contain bases "A", "G", "C", "T".
ssType	Numeric indicator, if the entered sequence is a splice donor (5) or acceptor (3)

Value

Character vector of the MaxEntScan scores generated from the entered seqVector.

Examples

```
calculateMaxEntScanScore("TTCCAAACGAACTTTTGTAGGGA", 3)
calculateMaxEntScanScore("GAGGTAAGT", 5)
```

 gene2transcript

Small data frame specifying a transcript to certain genes for synonymous use.

Description

Small data frame specifying a transcript to certain genes for synonymous use.

Usage

```
gene2transcript
```

Format

data frame

gene_name HGNC gene name

gene_ID Ensembl gene ID

transcript_ID Ensembl transcript ID

Examples

```
gene2transcript
```

generateHEXplorerPlot	<i>Generates plot with HZEI values and splice site strengths from a list holding information about an SNV.</i>
-----------------------	--

Description

This function generates a plot depicting the HZEI score changes and changes in the HBS or Max-EntScan score, from a sequence variation.

Usage

```
generateHEXplorerPlot(variationInfoList, ntWindow)
```

Arguments

variationInfoList	Output from the getSeqInfoFromVariation function.
ntWindow	Numeric value defining the sequence surrounding of interest.

Value

Plot stating the HZEI values per nt and splice site strength with and without the SNV.

Examples

```
#Defining exemplary input data
transcriptTable <- transCoord      # Using pseudo transcript table
transcriptID <- "pseudo_ENST00000650636"    # Using pseudo transcript
variation <- "c.412C>G/p.(T89M)"
ntWindow <- 20
gene2transcript <- data.frame(gene_name = "Example_gene",
gene_ID = "pseudo_ENSG00000147099", transcript_ID = "pseudo_ENST00000650636")

results <- getSeqInfoFromVariation(referenceDnaStringSet, transcriptID, variation, ntWindow=ntWindow, transcr

generateHEXplorerPlot(results)
```

getMaxEntInfo	<i>Generates table with MaxEntScan scores per potential SA position.</i>
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Description

This function generates a table with MaxEntScan scores per potential SA position.

Usage

```
getMaxEntInfo(seq)
```

Arguments

seq Nucleotide sequence longer than 22nt and only containing bases "A", "G", "C", "T".

Value

Dataframe of potential acceptor index positions and corresponding MaxEntScan scores.

Examples

```
getMaxEntInfo("TTCCAAACGAACCTTTGTAGGGA")
```

```
getSeqInfoFromVariation
```

Collects information about genomic context of sequence variants.

Description

This function collects information about genomic context of sequence variants.

Usage

```
getSeqInfoFromVariation(referenceDnaStringSet, transcriptID,  
variation, ntWindow=20, transcriptTable, gene2transcript=gene2transcript)
```

Arguments

referenceDnaStringSet DNAStringset from the reference genome fasta file.

transcriptID Ensembl ID of the transcript of interest.

variation A sequence variation either referring to coding sequence or the genomic sequence (c.12A>T, or g.182284A>T).

ntWindow Numeric value defining the sequence surrounding of interest.

transcriptTable Table of transcripts and their exon coordinates and CDS coordinates.

gene2transcript Gene to transcript conversion table with the gene name in the first column and the gene ID in the second and the transcript ID in the third column.

Value

List of informations about the entered variation.

Examples

```
#Defining exemplary input data
transcriptTable <- transCoord
transcriptID <- "pseudo_ENST00000650636"
variation <- "c.412C>G/p.(T89M)"
gene2transcript <- data.frame(gene_name = "Example_gene",
gene_ID = "pseudo_ENSG00000147099", transcriptID = "pseudo_ENST00000650636")

results <- getSeqInfoFromVariation(referenceDnaStringSet, transcriptID,
variation, ntWindow=20, transcriptTable, gene2transcript=gene2transcript)

#Using a predefined gene to transcript conversion
transcriptID <- "Example_gene"
results <- getSeqInfoFromVariation(referenceDnaStringSet, transcriptID,
variation, ntWindow=20, transcriptTable, gene2transcript=gene2transcript)
```

hbg	<i>Donor sequences and their HBS</i>
-----	--------------------------------------

Description

Donor sequences and their HBS

Usage

hbg

Format

- A data frame with columns:
- seq** 11nt long donor sequence
- hbs** HBS of the donor sequence

Examples

hbg

hex	<i>Hexamers and Z scores</i>
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Description

Hexamers and Z scores

Usage

hex

Format

A data frame with columns:

seq Sequence of the hexamer.

value ZEI-score of the hexamer from HEXplorer.

first First codon within the hexamer.

second Second codon within the hexamer.

first_AA First encoded amino acid within the hexamer (three letter code).

second_AA Second encoded amino acid within the hexamer (three letter code).

AA Both encoded amino acid within the hexamer

Examples

hex

prepareReferenceFasta *Imports Fasta file from filepath.*

Description

This function imports Fasta file of the reference genome into R environment as DNASTringset.

Usage

```
prepareReferenceFasta(filepath)
```

Arguments

filepath R conform filepath to the fasta file of the reference genome to use.

Value

Creates new DNASTringSet from the object stated by the entered filepath.

Examples

```
## Loading exemplary DNASTringSet
filepath <- system.file("extdata", "fastaEx.fa", package="Biostrings")
referenceDnaStringSet <- prepareReferenceFasta(filepath)
```

referenceDnaStringSet	<i>Small DNStringset as exemplary reference genome sequence</i>
-----------------------	---

Description

Small DNStringset as exemplary reference genome sequence

Usage

```
referenceDnaStringSet
```

Format

```
DNStringset
```

width Length of feature sequence

seq Sequence of the feature

names Name of the feature

Examples

```
referenceDnaStringSet
```

startVarConApp	<i>Start GUI of VarCon.</i>
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Description

Start graphical user interface for the VarCon application.

Usage

```
startVarConApp()
```

Value

Shiny app

Examples

```
## Not run:
startVarConApp()

## End(Not run)
```

transCoord*Small table as exemplary transcript table with exon coordinates*

Description

Small table as exemplary transcript table with exon coordinates.

Usage

transCoord

Format

data frame

Gene.stable.ID Ensembl gene ID

Transcript.stable.ID Ensembl Transcript ID

Strand Strand of the feature

Exon.region.start..bp. Smalles coordinate of the exon end coordinates of a specific exon

Exon.region.end..bp. Largest coordinate of the exon end coordinates of a specific exon

cDNA.coding.start Start of the coding sequence

cDNA.coding.end End of the coding sequence

CDS.start Covered coding nucleotides start

CDS.end Covered coding nucleotides end

Exon.rank.in.transcript Rank of the exon within the respective transcript

Exon.stable.ID Ensembl exon ID

Chromosome.scaffold.name Name of the chromosome

Examples

transCoord

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