

# Package ‘Site2Target’

November 14, 2025

**Type** Package

**Title** An R package to associate peaks and target genes

**Version** 1.2.0

**Description** Statistics implemented for both peak-wise and gene-wise associations. In peak-wise associations, the p-value of the target genes of a given set of peaks are calculated. Negative binomial or Poisson distributions can be used for modeling the unweighted peaks targets and log-normal can be used to model the weighted peaks. In gene-wise associations a table consisting of a set of genes, mapped to specific peaks, is generated using the given rules.

**BugReports** <https://github.com/fls-bioinformatics-core/Site2Target/issues>

**Depends** R (>= 4.4)

**License** GPL-2

**Encoding** UTF-8

**LazyData** false

**Imports** S4Vectors, stats, utils, BiocGenerics, GenomeInfoDb, MASS,  
IRanges, GenomicRanges

**biocViews** Annotation, ChIPSeq, Software, Epigenetics, GeneExpression,  
GeneTarget

**RoxygenNote** 7.3.2

**Suggests** BiocStyle, knitr, rmarkdown, testthat (>= 3.0.0)

**VignetteBuilder** knitr

**Config/testthat/edition** 3

**git\_url** <https://git.bioconductor.org/packages/Site2Target>

**git\_branch** RELEASE\_3\_22

**git\_last\_commit** 902e93a

**git\_last\_commit\_date** 2025-10-29

**Repository** Bioconductor 3.22

**Date/Publication** 2025-11-13

**Author** Peyman Zarrineh [cre, aut] (ORCID:  
<<https://orcid.org/0000-0003-4820-4101>>)

**Maintainer** Peyman Zarrineh <peyman.zarrineh@manchester.ac.uk>

## Contents

addColumn2geneWiseAssociation . . . . .	2
addRelation2geneWiseAssociation . . . . .	4
data . . . . .	5
extendSitesInGivenRegions . . . . .	7
genewiseAssociation . . . . .	7
getCenterOfPeaks . . . . .	9
getNameFromCoordinates . . . . .	9
getTargetGenesNumber . . . . .	10
getTargetGenesPvals . . . . .	11
getTargetGenesPvalsWithDNAInteractions . . . . .	12
getTargetGenesPvalsWithIntensities . . . . .	13
getTargetGenesPvalsWithIntensitiesAndDNAInteractions . . . . .	14
granges2String . . . . .	15
removeReserveCharacter . . . . .	16
site2GeneDistance . . . . .	16
Site2Target . . . . .	17
string2Granges . . . . .	18
Table2Granges . . . . .	18
<b>Index</b>	<b>20</b>

---

addColumn2geneWiseAssociation

*Add column to gene-wise association*

---

### Description

Add a column of values based on the type either genes or peaks.

The Input is either coordinates or names of genes or peaks plus a column of relevant values. This function add these values as a column to gene or peak table as well as the interaction table.

### Usage

```
addColumn2geneWiseAssociation(
  type = "",
  name = NULL,
  coordinates = NULL,
  columnName = NA,
  column,
  inFile = "geneWiseAssociation",
  outFile = "geneWiseAssociation"
)
```

**Arguments**

type	type of columns to be added. Either "gene" or "peak"
name	Names of genes or peaks
coordinates	Coordinates of genes or peaks in granges format
columnName	Column name that should be added to the tables
column	Column values that should be added to the tables
inFile	The name of the input folder (default "genewiseAssociation")
outFile	The name of the output folder (default "genewiseAssociation")

**Value**

No value returns just column would be added to the tables

**See Also**

[genewiseAssociation](#)

**Examples**

```
geneFile=system.file("extdata", "gene_expression.tsv", package="Site2Target")
geneCoords <- Table2Granges(geneFile)
geneTable <- read.table(geneFile, header=TRUE)

geneDEIndices <- which((abs(geneTable$logFC)>1)==TRUE)
indicesLen <- length(geneDEIndices)
if(indicesLen >0)
{
  geneTable <- geneTable[geneDEIndices,]
  geneCoords <- geneCoords[geneDEIndices]
}
geneDENames <- geneTable$name
geneDElogFC <- geneTable$logFC
geneCoordsDE <- geneCoords

tfFile =system.file("extdata", "MEIS_binding.tsv", package="Site2Target")
TFCoords <- Table2Granges(tfFile)
tfTable <- read.table(tfFile, header=TRUE)
tfIntensities <- tfTable$intensities

stats <-
genewiseAssociation(associationBy="distance",
  geneCoordinates=geneCoordsDE,
  geneNames=geneDENames,
  peakCoordinates=TFCoords,
  distance=50000,
  outFile="Gene_TF_50K")

stats

# add expression log fold changes to the table
addColumn2geneWiseAssociation(type="gene", name=geneDENames,
  columnName="Expr_logFC", column=geneDElogFC, inFile="Gene_TF_50K",
  outFile="Gene_TF_50K")

# add peak intensities to the table
```

```
addColumn2geneWiseAssociation(type="peak", coordinates=TFCoords,
    columnName="Binding_Intensities", column=tfIntensities,
    inFile="Gene_TF_50K", outFile="Gene_TF_50K")
```

---

```
addRelation2geneWiseAssociation
```

*Add a relation column to gene-peak interaction table*

---

### **Description**

Get coordinates of interactions (ex. HiC interactions) and a column of interaction values (ex. HiC intensities ) and add them as a column to gene-peak interaction table.

### **Usage**

```
addRelation2geneWiseAssociation(
    strand1 = NULL,
    strand2 = NULL,
    columnName,
    column,
    inFile = "geneWiseAssociation",
    outFile = "geneWiseAssociation"
)
```

### **Arguments**

strand1	granges of DNA strand1 linked to DNA strand2
strand2	granges of DNA strand2 linked to DNA strand1
columnName	Column name that should be added to the interaction table
column	Column values that should be added to the interaction table
inFile	The name of the input folder (default "genewiseAssociation")
outFile	The name of the output folder (default "genewiseAssociation")

### **Value**

No value would be returned just a column be added to link table

### **See Also**

[genewiseAssociation](#)

## Examples

```

geneFile=system.file("extdata", "gene_expression.tsv", package="Site2Target")
geneCoords <- Table2Granges(geneFile)
geneTable <- read.table(geneFile, header=TRUE)

geneDEIndices <- which((abs(geneTable$logFC)>1)==TRUE)
indicesLen <- length(geneDEIndices)
if(indicesLen >0)
{
  geneTable <- geneTable[geneDEIndices,]
  geneCoords <- geneCoords[geneDEIndices]
}
geneDENames <- geneTable$name
geneDElogFC <- geneTable$logFC
geneCoordsDE <- geneCoords

tfFile =system.file("extdata", "MEIS_binding.tsv", package="Site2Target")
TFCoords <- Table2Granges(tfFile)
tfTable <- read.table(tfFile, header=TRUE)

stats <-
genewiseAssociation(associationBy="distance",
                    geneCoordinates=geneCoordsDE,
                    geneNames=geneDENames,
                    peakCoordinates=TFCoords,
                    distance=50000,
                    outFile="Gene_TF_50K")

stats

HiCFile =system.file("extdata", "HiC_intensities.tsv", package="Site2Target")
HiCstr1 <- Table2Granges(HiCFile, chrColName="Strand1_chr",
                        startColName="Strand1_start", endColName="Strand1_end")
HiCstr2 <- Table2Granges(HiCFile, chrColName="Strand2_chr",
                        startColName="Strand2_start", endColName="Strand2_end")
HiCTable <- read.table(HiCFile, header=TRUE)
HiCintensities <- HiCTable$intensities

addRelation2geneWiseAssociation(strand1=HiCstr1, strand2=HiCstr2,
                                columnName="HiC_Intensities", column=HiCintensities,
                                inFile="Gene_TF_50K", outFile="Gene_TF_50K")

```

---

data

*MEIS cardiomyocytes datasets used in the package*

---

## Description

Human cardiomyocytes datasets are reduced in size by only using chr21. log fold changes of Gene expression WT vs MEIS KO from RNA-seq experiments, and binding sites of MEIS derived from a ChIP-seq experiment are the main experimental datasets representing relevant gene and peak information. HiC interactions and topologically associating domains (TADs) are derived from a HiC experiments are auxiliary datasets related to DNA-DNA interactions.

**Format**

Gene expression WT vs MEIS KO in chr21. MEIS binding sites in chr21. TADs, and HiC interactions in chr21.

**gene\_expression.tsv** Gene expression

**MEIS\_binding.tsv** MEIS binding sites

**TADs.tsv** TADs

**HiC\_intensities.tsv** HiC interactions

**Value**

Just description of data

**Examples**

```
## Gene expression table

# Read gene coordinates
geneFile=system.file("extdata", "gene_expression.tsv",
                     package="Site2Target")
geneCoords <- Table2Granges(geneFile)

# Read gene table
geneTable <- read.table(geneFile, header=TRUE)

## TF binding table

# Read peak coordinates
tfFile =system.file("extdata", "MEIS_binding.tsv",
                   package="Site2Target")
TFCoords <- Table2Granges(tfFile)

# Read MEIS binding intensities
tfTable <- read.table(tfFile, header=TRUE)

## DNA-DNA interactions

# Read TAD regions
TADsFile =system.file("extdata", "TADs.tsv",
                     package="Site2Target")
TADs <- Table2Granges(TADsFile)

# Read HiC interactions
HiCFile =system.file("extdata", "HiC_intensities.tsv",
                    package="Site2Target")
HiCstr1 <- Table2Granges(HiCFile, chrColName="Strand1_chr",
                       startColName="Strand1_start", endColName="Strand1_end")
HiCstr2 <- Table2Granges(HiCFile, chrColName="Strand2_chr",
                       startColName="Strand2_start", endColName="Strand2_end")

HiCTable <- read.table(HiCFile, header=TRUE)
```

---

 extendSitesInGivenRegions

*Extend sites given regions boundaries*


---

### Description

Get sites and given regions (ex. TADs or loops) coordinates.

It extends sites in a give region using a distance function

### Usage

```
extendSitesInGivenRegions(givenRegions, sites, distance = 1e+05)
```

### Arguments

givenRegions granges coordinates of given regions (ex. TAD or loops)

sites granges coordinates of sites

distance the maximum distance to associate sites to regions

### Value

A granges of the extended sites in given regions

### Examples

```
tffile =system.file("extdata", "MEIS_binding.tsv", package="Site2Target")
TFCoords <- Table2Granges(tffile)
```

```
TADsFile =system.file("extdata", "TADs.tsv",package="Site2Target")
TADs <- Table2Granges(TADsFile)
```

```
extendSitesInGivenRegions(TADs, TFCoords)
```

---

 genewiseAssociation *Generate genewise association between genes and peaks*


---

### Description

Get genomic coordinates of a set of genes and a set of peaks associate them by a fixed distance (default 50K nt). It also associate genes and peaks for provided DNA-DNA interaction from a dataset like HiC. This function can also associate genes and user provided regions (ex. TADs, subTADs, etc). It generates three tables: Gene table, peak table, and Gene-Peak association table.

**Usage**

```
genewiseAssociation(
  associationBy = "distance",
  geneCoordinates = NULL,
  geneNames = NULL,
  peakCoordinates = NULL,
  peakNames = NULL,
  distance = 50000,
  givenRegions = NULL,
  strand1 = NULL,
  strand2 = NULL,
  outFile = "genewiseAssociation"
)
```

**Arguments**

associationBy	Can be "distance", "regions", or "DNAinteractions"
geneCoordinates	Gene coordinates in granges format
geneNames	Gene names can be provided by the user
peakCoordinates	Peak coordinates in granges format
peakNames	Peak names can be provided by the user
distance	The maximum distance to associate peaks to genes. default 50K
givenRegions	granges coordinates of given regions (ex. TAD or loops)
strand1	granges of DNA strand1 linked to DNA strand2
strand2	granges of DNA strand2 linked to DNA strand1
outFile	The name of the output folder (default "genewiseAssociation")

**Value**

A vector of portions of linked genes and linked peaks

**Examples**

```
geneFile=system.file("extdata", "gene_expression.tsv", package="Site2Target")
geneCoords <- Table2Granges(geneFile)
geneTable <- read.table(geneFile, header=TRUE)

geneDEIndices <- which((abs(geneTable$logFC)>1)==TRUE)
indicesLen <- length(geneDEIndices)
if(indicesLen >0)
{
  geneTable <- geneTable[geneDEIndices,]
  geneCoords <- geneCoords[geneDEIndices]
}
geneDENames <- geneTable$name
geneDElogFC <- geneTable$logFC
geneCoordsDE <- geneCoords

tfFile =system.file("extdata", "MEIS_binding.tsv", package="Site2Target")
```

```
TFCoords <- Table2Granges(tfFile)
tfTable <- read.table(tfFile, header=TRUE)

stats <-
  genewiseAssociation(associationBy="distance",
                      geneCoordinates=geneCoordsDE,
                      geneNames=geneDENames,
                      peakCoordinates=TFCoords,
                      distance=50000,
                      outFile="Gene_TF_50K")

stats
```

---

getCenterOfPeaks      *Return center of the given granges files*

---

### Description

Get a granges and find the center of it

### Usage

```
getCenterOfPeaks(gr)
```

### Arguments

gr                    granges coordinate

### Value

granges format of the center

### Examples

```
tfFile =system.file("extdata", "MEIS_binding.tsv", package="Site2Target")
TFCoords <- Table2Granges(tfFile)
TFCoordsCenters <- getCenterOfPeaks(TFCoords)
TFCoordsCenters
```

---

getNameFromCoordinates

*Get names of genes or peaks related to a query coordinates*

---

### Description

Get names and coordinates of genes or peaks. It also get the coordinates of query regions and returns the related genes or peak names.

**Usage**

```
getNameFromCoordinates(names, coordinates, queryCoordinates)
```

**Arguments**

```
names          Names of genes or peaks
coordinates    Coordinates of genes or peaks in granges format
queryCoordinates
               Coordinates of the query regions in granges format
```

**Value**

Names of genes or peaks in queried regions

---

```
getTargetGenesNumber  generate number of sites per gene given distances
```

---

**Description**

Get genes and sites coordinates, and associate them by given distance.

**Usage**

```
getTargetGenesNumber(geneCoordinates = NA, sites = NA, distance = 50000)
```

**Arguments**

```
geneCoordinates    granges coordinates of genes
sites              granges coordinates of sites
distance           the maximum distance to associate sites to genes. default 50K
```

**Value**

A vector sites number matched to each gene

**Examples**

```
geneFile=system.file("extdata", "gene_expression.tsv", package="Site2Target")
geneCoords <- Table2Granges(geneFile)

tfFile =system.file("extdata", "MEIS_binding.tsv", package="Site2Target")
TFCoords <- Table2Granges(tfFile)

targetNum <- getTargetGenesNumber( geneCoords, TFCoords)
```

---

getTargetGenesPvals     *Fit Negative binomial distribution to target genes*

---

### Description

Get genes and sites coordinates, and associate them by given distance or given regions (ex. TADs or loops). It tests the distribution of sites around genes either by poisson or negative binomial test.

### Usage

```
getTargetGenesPvals(  
  associationBy = "distance",  
  dist = "negative binomial",  
  geneCoordinates = NA,  
  sites = NA,  
  distance = 50000,  
  givenRegions = NA  
)
```

### Arguments

associationBy	either "distance" or "regions"
dist	either "negative binomial" or "poisson"
geneCoordinates	granges coordinates of genes
sites	granges coordinates of sites
distance	the maximum distance to associate sites to genes. default 50K
givenRegions	user provided granges regions like TADs or loops

### Value

A vector of pvalue distribution for target genes

### Examples

```
geneFile=system.file("extdata", "gene_expression.tsv", package="Site2Target")  
geneCoords <- Table2Granges(geneFile)  
  
tfFile =system.file("extdata", "MEIS_binding.tsv", package="Site2Target")  
TFCoords <- Table2Granges(tfFile)  
  
pvals <- getTargetGenesPvals( geneCoordinates=geneCoords, sites=TFCoords)
```

---

```
getTargetGenesPvalsWithDNAInteractions
```

*Fit Negative binomial distribution to target genes*

---

### Description

Get genes and sites coordinates, and associate them by given distance and user provided DNA interaction (ex. HiC). It tests the distribution of sites around genes either by poisson or negative binomial test.

### Usage

```
getTargetGenesPvalsWithDNAInteractions(
  dist = "negative binomial",
  geneCoordinates = NA,
  sites = NA,
  strand1 = NA,
  strand2 = NA,
  distance = 50000
)
```

### Arguments

dist	either "negative binomial" or "poisson"
geneCoordinates	granges coordinates of genes
sites	granges coordinates of sites
strand1	granges of DNA strand1 linked to DNA strand2
strand2	granges of DNA strand2 linked to DNA strand1
distance	the maximum distance to associate sites to genes. default 50K

### Value

A vector of pvalue distribution for target genes

### Examples

```
geneFile=system.file("extdata", "gene_expression.tsv", package="Site2Target")
geneCoords <- Table2Granges(geneFile)

tfFile =system.file("extdata", "MEIS_binding.tsv", package="Site2Target")
TFCoords <- Table2Granges(tfFile)

HiCFile =system.file("extdata", "HiC_intensities.tsv", package="Site2Target")
HiCstr1 <- Table2Granges(HiCFile, chrColName="Strand1_chr",
                        startColName="Strand1_start", endColName="Strand1_end")
HiCstr2 <- Table2Granges(HiCFile, chrColName="Strand2_chr",
                        startColName="Strand2_start", endColName="Strand2_end")
```

```
pvals <- getTargetGenesPvalsWithDNAInteractions(  
  geneCoordinates=geneCoords, sites=TFCoords, strand1=HiCstr1,  
  strand2=HiCstr2)
```

---

getTargetGenesPvalsWithIntensities

*Fit log-normal distribution to target genes*

---

### Description

Get genes and sites coordinates, and associate them by given distance or given regions (ex. TADs or loops). It tests the distribution of log-intensities of sites around genes by log-normal test. This function consider both binding sites and intensities.

### Usage

```
getTargetGenesPvalsWithIntensities(  
  associationBy = "distance",  
  intensities,  
  geneCoordinates = NA,  
  sites = NA,  
  distance = 50000,  
  givenRegions = NA  
)
```

### Arguments

associationBy	either "distance" or "regions"
intensities	intensity values associated to sites
geneCoordinates	granges coordinates of genes
sites	granges coordinates of sites
distance	the maximum distance to associate sites to genes. default 50K
givenRegions	user provided granges regions like TADs or loops

### Value

A vector of pvalue distribution for target genes

**Examples**

```

geneFile=system.file("extdata", "gene_expression.tsv", package="Site2Target")
geneCoords <- Table2Granges(geneFile)

tfFile =system.file("extdata", "MEIS_binding.tsv", package="Site2Target")
TFCoords <- Table2Granges(tfFile)
tfTable <- read.table(tfFile, header=TRUE)
tfIntensities <- tfTable$intensities

pvals <- getTargetGenesPvalsWithIntensities(geneCoordinates=geneCoords,
                                             sites=TFCoords, intensities=tfIntensities)

```

---

```

getTargetGenesPvalsWithIntensitiesAndDNAInteractions
Fit log-normal distribution to target genes

```

---

**Description**

Get genes and sites coordinates, and associate them by given distance and user provided DNA interaction (ex. HiC). It tests the distribution of log-intensities of sites around genes by log-normal test. This function consider both binding sites and intensities.

**Usage**

```

getTargetGenesPvalsWithIntensitiesAndDNAInteractions(
  geneCoordinates,
  sites,
  intensities,
  strand1,
  strand2,
  distance = 50000
)

```

**Arguments**

geneCoordinates	granges coordinates of genes
sites	granges coordinates of sites
intensities	intensity values associated to sites
strand1	granges of DNA strand1 linked to DNA strand2
strand2	granges of DNA strand2 linked to DNA strand1
distance	the maximum distance to associate sites to genes. default 50K

**Value**

A vector of pvalue distribution for target genes

**Examples**

```
geneFile=system.file("extdata", "gene_expression.tsv", package="Site2Target")
geneCoords <- Table2Granges(geneFile)

tfFile =system.file("extdata", "MEIS_binding.tsv", package="Site2Target")
TFCoords <- Table2Granges(tfFile)
tfTable <- read.table(tfFile, header=TRUE)
tfIntensities <- tfTable$intensities

HiCFile =system.file("extdata", "HiC_intensities.tsv", package="Site2Target")
HiCstr1 <- Table2Granges(HiCFile, chrColName="Strand1_chr",
                        startColName="Strand1_start", endColName="Strand1_end")
HiCstr2 <- Table2Granges(HiCFile, chrColName="Strand2_chr",
                        startColName="Strand2_start", endColName="Strand2_end")

pvals <- getTargetGenesPvalsWithIntensitiesAndDNAInteractions(
  geneCoordinates=geneCoords, sites=TFCoords,
  intensities=tfIntensities, strand1=HiCstr1,
  strand2=HiCstr2)
```

---

`granges2String`*Convert granges to strings of coordinates*

---

**Description**

Get genomic coordinates granges and convert them to strings

**Usage**

```
granges2String(gr)
```

**Arguments**

`gr` granges coordinates

**Value**

string of coordinates

**Examples**

```
tfFile =system.file("extdata", "MEIS_binding.tsv", package="Site2Target")
TFCoords <- Table2Granges(tfFile)
strCoords <- granges2String(TFCoords)
head(strCoords)
```

removeReserveCharacter

*Remove reserved characters from a string*

---

**Description**

Remove reserved characters (such as \*, +, -, etc) from a string

**Usage**

```
removeReserveCharacter(name)
```

**Arguments**

name                    A string of characters

**Value**

A string without reserved characters

**Examples**

```
removeReserveCharacter("A&%B^f6")
```

---

site2GeneDistance

*Return the distance between paired peaks and genes*

---

**Description**

Get a granges of genes and peaks and return their distances

**Usage**

```
site2GeneDistance(geneCoordinates, peakCoordinates)
```

**Arguments**

geneCoordinates  
                          granges coordinates of genes

peakCoordinates  
                          granges coordinates of peaks

**Value**

the respective distances of paired genes and peaks

---

Site2Target

Associate peaks and target genes

---

## Description

Statistical implementation for both peak-wise and gene-wise associations. Here is an example of a peak-wise and a gene-wise association of differential genes WT vs KO of a transcription factor and binding sites of this transcription factor.

## Value

Just an example

## Examples

```
geneFile=system.file("extdata", "gene_expression.tsv", package="Site2Target")
geneCoords <- Table2Granges(geneFile)
geneTable <- read.table(geneFile, header=TRUE)

tfFile =system.file("extdata", "MEIS_binding.tsv", package="Site2Target")
TFCoords <- Table2Granges(tfFile)
tfTable <- read.table(tfFile, header=TRUE)

## Peakwise association example

pvals <- getTargetGenesPvals(geneCoordinates=geneCoords, sites=TFCoords)
topTargetNum <- 5
topTargetIndex <- order(pvals)[1:topTargetNum]

# Make a data frame of peak targets pvalues and expression logFCs

dfTopTarget <-
  data.frame(name=geneTable$name[topTargetIndex],
             pvalue=pvals[topTargetIndex],
             exprLogC=geneTable$logFC[topTargetIndex]
             )
dfTopTarget

## Genewise association example
geneDEIndices <- which((abs(geneTable$logFC)>1)==TRUE)
indicesLen <- length(geneDEIndices)
if(indicesLen >0)
{
  geneTable <- geneTable[geneDEIndices,]
  geneCoords <- geneCoords[geneDEIndices]
}
geneDENames <- geneTable$name
geneDElogFC <- geneTable$logFC
geneCoordsDE <- geneCoords

stats <-
genewiseAssociation(associationBy="distance",
                    geneCoordinates=geneCoordsDE,
                    geneNames=geneDENames,
```

```

peakCoordinates=TFCoords,
distance=50000,
outFile="Gene_TF_50K")
stats

```

---

string2Granges	<i>Convert strings to granges of coordinates</i>
----------------	--------------------------------------------------

---

### Description

Get genomic coordinates as strings and convert them to granges

### Usage

```
string2Granges(strCoordinates)
```

### Arguments

strCoordinates string of coordinates

### Value

Genomic coordinates in granges format

### Examples

```
string2Granges(c("chr1:1112-1231", "ch2:3131-3221"))
```

---

Table2Granges	<i>Take Genomic Ranges from a table file</i>
---------------	----------------------------------------------

---

### Description

Read a table file and derive genomic ranges from user provided column names.

### Usage

```

Table2Granges(
  fileName,
  chrColName = "chr",
  startColName = "start",
  endColName = "end"
)

```

**Arguments**

fileName	A table delimited file
chrColName	Chromosomes column name (default: "Chr")
startColName	Start column name (default: "start")
endColName	End column name (default: "end")

**Value**

granges format of given coordinates

**Examples**

```
geneFile=system.file("extdata", "gene_expression.tsv", package="Site2Target")
grs <- Table2Granges(fileName=geneFile,
                     chrColName="chr",
                     startColName="start",
                     endColName="end")

grs
```

# Index

[addColumn2geneWiseAssociation](#), [2](#)  
[addRelation2geneWiseAssociation](#), [4](#)  
  
[data](#), [5](#)  
  
[extendSitesInGivenRegions](#), [7](#)  
  
[genewiseAssociation](#), [3](#), [4](#), [7](#)  
[getCenterOfPeaks](#), [9](#)  
[getNameFromCoordinates](#), [9](#)  
[getTargetGenesNumber](#), [10](#)  
[getTargetGenesPvals](#), [11](#)  
[getTargetGenesPvalsWithDNAInteractions](#),  
[12](#)  
[getTargetGenesPvalsWithIntensities](#), [13](#)  
[getTargetGenesPvalsWithIntensitiesAndDNAInteractions](#),  
[14](#)  
[granges2String](#), [15](#)  
  
[removeReserveCharacter](#), [16](#)  
  
[site2GeneDistance](#), [16](#)  
[Site2Target](#), [17](#)  
[string2Granges](#), [18](#)  
  
[Table2Granges](#), [18](#)