Package 'stepNorm'

December 5, 2025

Version 1.83.0

Date 2008-10-08

Title Stepwise normalization functions for cDNA microarrays

Author Yuanyuan Xiao <yxiao@itsa.ucsf.edu>, Yee Hwa (Jean) Yang <jean@biostat.ucsf.edu>

Depends R (>= 1.8.0), marray, methods

Imports marray, MASS, methods, stats

Maintainer Yuanyuan Xiao <yxiao@itsa.ucsf.edu>

Description Stepwise normalization functions for cDNA microarray data.

License LGPL

URL http://www.biostat.ucsf.edu/jean/

biocViews Microarray, TwoChannel, Preprocessing

git_url https://git.bioconductor.org/packages/stepNorm

git_branch devel

git_last_commit efe8204

git_last_commit_date 2025-10-29

Repository Bioconductor 3.23

Date/Publication 2025-12-04

Contents

calcAIC 2
calcBIC 3
fit2DWithin 5
fitWithin 7
maCompPlate2 9
makeStepList 11
marrayFit-class 13
seqWithinNorm 14

stepNorm-internal 17
stepWithinNorm 17
withinNorm 20

Index **23**

`calcAIC` *Extract AIC from a Fitted Model*

Description

Computes the Akaike Information Criterion for a fitted parametric model.

Usage

```
calcAIC(fit, subset=TRUE, scale = 0, enp, loss.fun = square)
```

Arguments

`fit`	fitted model; see details below
`scale`	optional numeric specifying the scale parameter of the model; see `scale` in `step`.
`subset`	A "logical" or "numeric" vector indicating the subset of points used to compute the fitted model.
`enp`	equivalent number of parameters in the fitted model. If missing, the `enp` component from fit will be used.
`loss.fun`	the loss function used to calculate deviance; default uses the squared deviations from the fitted values; one could also use, for example, absolute deviations (`abs`).

Details

The argument `fit` can be an object of class `marrayFit`, in which case the `residuals` component from the `marrayFit` object will be extracted to calculate the deviance; the user can also pass in a numeric vector, in which case it will be interpreted as the residuals and the user needs to specify the argument `enp`.

The criterion used is

$$AIC = -2 * logL + k * enp,$$

where L is the likelihood and `enp` the equivalent number of parameters of `fit`. For linear models (as in marrayFit), $-2logL$ is computed from the deviance.

`k = 2` corresponds to the traditional AIC and is the penalty for the number of parameters.

Value

A numeric vector of length 4, giving

`Dev`	the deviance of the `fit`.
`enp`	the equivalent number of parameters of the `fit`.
`penalty`	the penalty for number of parameters.
`Criterion`	the Akaike Information Criterion for `fit`.

Author(s)

Yuanyuan Xiao, <yxiao@itsa.ucsf.edu>,
Jean Yee Hwa Yang, <jean@biostat.ucsf.edu>

See Also

AIC, deviance, calcBIC.

Examples

```
## load in swirl data
data(swirl)

## fit a model
fit <- fitWithin(fun="medfit")
## res is an object of class marrayFit
res <- fit(swirl[,1])

## calculate AIC
calcAIC(res)
## or could pass in the residual vector, but then argument "enp" needs to be specified
calcAIC(res$residual, enp=1)
```

---

calcBIC *Extract BIC from a Fitted Model*

Description

Computes the Bayesian Information Criterion for a fitted parametric model.

Usage

```
calcBIC(fit, subset=TRUE, scale = 0, enp, loss.fun = square)
```

Arguments

fit	fitted model; see details below
subset	A "logical" or "numeric" vector indicating the subset of points used to compute the fitted model.
scale	optional numeric specifying the scale parameter of the model; see scale in step.
enp	equivalent number of parameters in the fitted model. If missing, the enp component from fit will be used.
loss.fun	the loss function used to calculate deviance; the default uses the squared deviation from the fitted values; one could also use absolute deviations (abs).

Details

The argument fit can be an object of class marrayFit, in which case the residuals component from the marrayFit object will be extracted to calculate the deviance; the user can also pass in a numeric vector, in which case it will be interpreted as the residuals and the user needs to specify the argument enp.

The criterion used is

$$BIC = -2 * logL + k * enp,$$

where L is the likelihood and enp the equivalent number of parameters of fit. For linear models (as in marrayFit), $-2logL$ is computed from the deviance.

k = log(n) corresponds to the BIC and is the penalty for the number of parameters.

Value

A numeric vector of length 4, giving

Dev	the deviance of the fit.
enp	the equivalent number of parameters of the fit.
penalty	the penalty for number of parameters.
Criterion	the Akaike Information Criterion for fit.

Author(s)

Yuanyuan Xiao, <yxiao@itsa.ucsf.edu>,
Jean Yee Hwa Yang, <jean@biostat.ucsf.edu>

See Also

AIC, deviance, calcAIC.

Examples

```
## load in swirl data
data(swirl)

## fit a model
fit <- fitWithin(fun="medfit")
## res is an object of class marrayFit
res <- fit(swirl[,1])

## calculate BIC
calcBIC(res)
## or could pass in the residual vector, but then argument "enp" needs to be specified
calcBIC(res$residual, enp=1)
```

`fit2DWithin` *Bivariate location normalization function for cDNA microarray data*

Description

This function performs 2D location normalization on cDNA micoroarray. It operates on class `marrayRaw` or class `marrayNorm`. It allows the user to choose from a set of four basic normalization procedures.

Usage

```
fit2DWithin(x1.fun = "maSpotRow", x2.fun = "maSpotCol", y.fun = "maM",
subset=TRUE, fun = aov2Dfit, ...)
```

Arguments

`x1.fun`	Name of accessor method for spot row coordinates, usually `maSpotRow`.
`x2.fun`	Name of accessor method for spot column coordinates, usually `maSpotCol`.
`y.fun`	Name of accessor method for spot statistics, usually the log-ratio `maM`.
`subset`	A "logical" or "numeric" vector indicating the subset of points used to compute the normalization values.
`fun`	Character string specifying the normalization procedures: **rlm2Dfit** for robust linear regression using the `rlm` function **loess2Dfit** for robust local regression using the `loess` function **aov2Dfit** for linear regression using the `lm` function **spatialMedfit** for spatial median normalization
`...`	Misc arguments for `fun`

Details

The spot statistic named in `y` is regressed on spot row and column coordinates, using the function specified by the argument `fun`. Typically, `rlm2Dfit` and `loess2Dfit`, which treat row and column coordinates as numeric vectors, require a lot fewer parameters than `aov2Dfit` which specifies these two variables as categorical. `spatialMedfit` could yet fit the most complicated model, depending on size of the smoothing window specified; details see Wison et al (2003).

Value

The function `fit2DWithin` returns a function (F) with bindings for `x1.fun`, `x2.fun`, `y.fun`, `subset` and `fun`. When the function F is evaluated with an object of class `marrayNorm` or `marrayRaw`, it carries out normalization and returns an object of class `marrayFit` that contains the normalization information as a list with the following components:

`varfun`	: A character vector of names of predictor variables.
`x`	: A numeric matrix of predictor variables.
`y`	: A numeric matrix of responses.
`residuals`	: A numeric matrix of normalized values (typically log ratios (M)).
`fitted`	: A numeric matrix of the fitted values.
`enp`	: The equivalent number of parameters; see `loess`.
`df.residual`	: The residual degrees of freedom.
`fun`	: A character string indicating the name of the function used for normalization.

Note that the `residuals` component stores the normalized ratios.

Author(s)

Yuanyuan Xiao, <yxiao@itsa.ucsf.edu>,
Jean Yee Hwa Yang, <jean@biostat.ucsf.edu>

References

Y. H. Yang, S. Dudoit, P. Luu, and T. P. Speed (2001). Normalization for cDNA microarray data. In M. L. Bittner, Y. Chen, A. N. Dorsel, and E. R. Dougherty (eds), *Microarrays: Optical Technologies and Informatics*, Vol. 4266 of *Proceedings of SPIE*.

D. L. Wilson, M. J. Buckley, C. A. Helliwell and I. W. Wilson (2003). New normalization methods for cDNA microarray data. *Bioinformatics*, Vol. 19, pp. 1325-1332.

See Also

`fitWithin`

Examples

```
## use the swirl data as example
data(swirl)

## 2D rlm normalization
rlm2D <- fit2DWithin(fun="rlm2Dfit")
swirl1.rlm <- rlm2D(swirl[,1])
norm.M <- swirl1.rlm$residuals ## matrix of normalized ratios

## 2D loess normalization, default span=0.2
loess2D <- fit2DWithin(fun="loess2Dfit")
swirl1.loess <- loess2D(swirl[,1])
## 2D loess normalization, span=0.4
## Not run:
loess2D.1 <- fit2DWithin(fun="loess2Dfit", span=0.4)
swirl1.loess.1 <- loess2D.1(swirl[,1])
## End(Not run)

## 2D aov normalization
aov2D <- fit2DWithin(fun="aov2Dfit")
swirl1.aov <- aov2D(swirl[,1])

## 2D spatial median normalization, default window width=3
spatialMed2D <- fit2DWithin(fun="spatialMedfit")
swirl1.spatialMed <- spatialMed2D(swirl[,1])
## 2D loess normalization, window width=9
## Not run:
spatialMed2D.1 <- fit2DWithin(fun="spatialMedfit", width=9)
swirl1.spatialMed.1 <- spatialMed2D.1(swirl[,1])
## End(Not run)
```

`fitWithin` *Simple location normalization function for cDNA microarray data*

Description

This function performs location normalization on cDNA micoroarray. It operates on class `marrayRaw` or class `marrayNorm`. It allows the user to choose from a set of three basic normalization procedures.

Usage

```
fitWithin(x.fun = "maA", y.fun = "maM", z.fun = TRUE, subset=TRUE, fun = "medfit", ...)
```

Arguments

`x.fun`	Name of accessor method for spot intensity, usually `maA`.
`y.fun`	Name of accessor method for spot statistics, usually the log-ratio `maM`.

z.fun	Name of accessor method for spot statistic used to stratify the data, usually a layout parameter, e.g. maPrintTip or maCompPlate. If z is not a character, e.g. NULL, the data are not stratified.
subset	A "logical" or "numeric" vector indicating the subset of points used to compute the normalization values.
fun	Character string specifying the normalization procedure: **medfit** for global median location normalization **rlmfit** for global intensity or A-dependent location normalization using the rlm function **loessfit** for global intensity or A-dependent location normalization using the loess function
...	Miscs arguments to be passed in fun

Details

Normalization is typically performed on the expression ratios of cDNA microarray data, using the function specified by argument fun. Currently, this function is to be chosen from: medfit (median), rlmfit (rlm) and loessfit(loess). When z.fun is provided as a character string, for example, maPrintTip, the normalization procedure is operated within each print-tip of the slide.

Value

The function fitWithin returns a function(F) with bindings for x.fun, y.fun, z.fun, subset and fun. When the function F is evaluated with an object of class marrayNorm or marrayRaw, it carries out normalization and returns an object of class marrayFit that contains the normalization information as a list with the following list components:

varfun	: A character vector of names of predictor variables.
x	: A numeric matrix of predictor variables.
y	: A numeric matrix of repsonses.
residuals	: A numeric matrix of normalized values (typically log ratios (M)).
fitted	: A numeric matrix of the fitted values.
enp	: The equivalent number of parameters; see loess.
df.residual	: The residual degrees of freedom.
fun	: A character string indicating the name of the function used for normalization.

Note that the residuals component stores the normalized ratios.

Author(s)

Yuanyuan Xiao, <yxiao@itsa.ucsf.edu>,
Jean Yee Hwa Yang, <jean@biostat.ucsf.edu>

References

Y. H. Yang, S. Dudoit, P. Luu, and T. P. Speed (2001). Normalization for cDNA microarray data. In M. L. Bittner, Y. Chen, A. N. Dorsel, and E. R. Dougherty (eds), *Microarrays: Optical Technologies and Informatics*, Vol. 4266 of *Proceedings of SPIE*.

See Also

fit2DWithin

Examples

```
## using the swirl data as example
data(swirl)

## median normalization
med <- fitWithin(fun="medfit")
swirl1.med <- med(swirl[,1])
norm.M <- swirl1.med$residuals ## matrix of normalized ratios

## rlm normalization
rlmF <- fitWithin(fun="rlmfit")
swirl1.rlm <- rlmF(swirl[,1])

## loess normalization, default span=0.4
loessF <- fitWithin(fun="loessfit")
swirl1.loess <- loessF(swirl[,1])
## loess normalization, span=0.2
loessF.1 <- fitWithin(fun="loessfit", span=0.2)
swirl1.loess.1 <- loessF.1(swirl[,1])


## within-printtip loess normalization
loessP <- fitWithin(z.fun="maPrintTip", fun="loessfit")
swirl1.loessP <- loessP(swirl[,1])
```

---

maCompPlate2 *Generate plate IDs*

Description

This function is a modification of the maCompPlate function in the marray library. It generates plate IDs from the dimensions of the grid and spot matrices. Unlike the maCompPlate function, the number of spots is not necessarily a multiple of the number of wells on a plate, therefore this function allows empty spots on the slide.

Usage

```
maCompPlate2(no.plates = NULL, n = 384)
```

Arguments

no.plates	object of class "numeric", number of plates used specified by the user. If a number is not specified, then it is assumed that there are no empty spots on the slide.
n	object of class "numeric", number of wells in each plate, usually 384 or 96.

Details

This function can be used to handle three cases: 1) the number of spots is a multiple of the number of wells on a plate (usually 96 or 384); 2) the number of spots is not a multiple of the number of wells on a plate, and several of spots on the slide are therefore left empty. In this case, the user needs to specify the number of total plates used; plate IDs of empty spots will be NAs; 3)the number of spots is not a multiple of the number of wells on a plate, but all spots on the slide are spotted, therefore there is one plate not fully used. In this case, the user does not need to specify the number of total plates (as this will not be an integer), the function assumes no empty spots on the slide automatically. See Examples below.

Value

The function `maCompPlate2` returns a function with bindings for `no.plates` and `n`, which when receiving a object of `marrayRaw`, `marrayNorm` or `marrayLayout` class, it returns a vector of plate IDs (`factor`).

Author(s)

Yuanyuan Xiao

See Also

`maCompPlate`, `marrayLayout`.

Examples

```
####### case 1: no empty spots on the slide, full plates used
L<-new("marrayLayout", maNgr=4, maNgc=4, maNsr=22, maNsc=24)
### "compPlate" is a function
compPlate <- maCompPlate2(n=384)
plate <- compPlate(L)
table(plate)
### can also use:
plate<-maCompPlate(L,384)
table(plate)

####### case 2: with empty spots on the slide, full plates used
L<-new("marrayLayout", maNgr=4, maNgc=4, maNsr=22, maNsc=26)
### "compPlate" is a function
compPlate <- maCompPlate2(no.plates=22,n=384)
plate <- compPlate(L)
table(plate)
### empty spots are NAs
unique(plate)

###### case 3: no empty spots on the slide, one plate not full
L<-new("marrayLayout", maNgr=4, maNgc=4, maNsr=22, maNsc=26)
### argument no.plates not specified, the function assumes no empty spots
compPlate <- maCompPlate2(n=384)
plate <- compPlate(L)
### 23 full plates (384), the 24th not full (304)
```

```
table(plate)
### no NAs, no empty spots
unique(plate)
```

---

makeStepList *Construction of a stepwise normalization list*

Description

This function provides a user friendly way to construct a list for input to the function stepWithinNorm. The list indicates intended biases for correction and models for stepwise normalization.

Usage

```
  makeStepList(A = c("median", "rlm", "loess"), PT = c("median", "rlm",
"loess"), PL = c("median", "rlm", "loess"), Spatial2D = c("rlm2D",
"loess2D", "aov2D", "spatialMedian"))
```

Arguments

A	A character string specifying the normalization models for the adjustment of intensity or A bias: **median:** global median location normalization **rlm:** global intensity or A-dependent robust linear normalization using the rlm function **loess:** global intensity or A-dependent robust nonlinear normalization using the loess function The user can specify any of these three choices and the selected model will be compared based the goodness fit and model parsimony; If the correction of the A bias is not desired, the user can set A = NULL.
PT	A character string specifying the normalization models for the adjustment of print-tip or PT bias: **median:** within-print-tip-group median normalization **rlm:** within-print-tip-group robust linear normalization using the rlm function **loess:** within-print-tip-group robust nonlinear normalization using the loess function **none:** no normalization for the PT bias If the correction of the PT bias is not desired, the user can set PT = NULL.
PL	A character string specifying the normalization models for the adjustment of well-plate or PL bias: **median:** within-well-plate median normalization **rlm:** within-well-plate robust linear normalization using the rlm function **loess:** within-well-plate robust nonlinear normalization using the loess function

none: no normalization for the PL bias

If the correction of the PL bias is not desired, the user can set `PL = NULL`.

`Spatial2D` A character string specifying the normalization models for the adjustment of spatial 2D bias:

none: no normalization for the spatial 2D bias

aov2D: spatial bivariate location normalization using ANOVA

rlm2D: spatial bivariate location normalization using the `rlm` function

loess2D: spatial bivariate location normalization using the `loess` function

spatialMedian: spatial location normalization using a spatial median approach (see Wilson et al. (2003) in reference)

If the correction of the PL bias is not desired, the user can set `Spatial2D = NULL`.

Details

This function provides a user friendly way to specify the parameter `wf.loc` for the main stepwise normalization function `stepWithinNorm`; see examples for details.

Value

An object of class "list" for input to the `stepWithinNorm` function.

Author(s)

Yuanyuan Xiao, <yxiao@itsa.ucsf.edu>,
Jean Yee Hwa Yang, <jean@biostat.ucsf.edu>

See Also

`stepWithinNorm`.

Examples

```
# Examples use swirl dataset, for description type ? swirl
data(swirl)

# To use the default parameters, which adjusts A, PT, PL and Spatial2D
# biases sequentially and compares all models available, simple type

wf.loc <- makeStepList()

# To apply loess for the A bias, and to omit the Spatial2D step
wf.loc <- makeStepList(A=("loess"), Spatial2D=NULL)

# To compare only rlm and loess in the A bias step, and other biases as default
wf.loc <- makeStepList(A=c("rlm","loess"))

# input to the stepWithinNorm function
## Not run:
```

```
step.swirl1 <- stepWithinNorm(swirl[,1],wf.loc=wf.loc)
## End(Not run)
```

`marrayFit-class` *Class "marrayFit", storing parameters and results of post-normalization cDNA microarray data*

Description

A simple list-based class for the storage of parameters and results of normalization of cDNA microarray data.

Creating Objects from the Class

Objects can be created by calls of the form `new('marrayFit', fit)` where `fit` is a list. Objects of `marrayFit` in the `StepNorm` package are typically created by functions `fitWithin` and `fit2DWithin`.

List Components

This class contains no slots, but objects should contain the following list components:

varfun : A character vector of names of predictor variables.

x : A numeric matrix of predictor variables.

y : A numeric matrix of responses.

residuals : A numeric matrix of normalized values (typically log ratios (M)).

fitted : A numeric matrix of the fitted values.

enp : The equivalent number of parameters; see `loess`.

df.residual : The residual degrees of freedom.

fun : A character string indicating the name of the function used for normalization.

Methods

This class inherits directly from class `list` so any operation appropriate for lists will work on objects of this class.

Author(s)

Yuanyuan Xiao, <yxiao@itsa.ucsf.edu>,
Jean Yee Hwa Yang, <jean@biostat.ucsf.edu>

See Also

`fitWithin`, `fit2DWithin`.

Examples

```
## load in swirl data
data(swirl)

## median normalization for the first slide of the swirl data
medWithin <- fitWithin(fun="medfit")
## medFit is an object of class marrayFit
medFit <- medWithin(swirl[,1])
## normalized ratios is stored in:
norm.M <- medFit$residuals
```

`seqWithinNorm` *Sequential within-slide normalization function*

Description

This function conducts cDNA microarray normalization in a sequential fashion. In a two-color cDNA array setting, within-slide normalization calibrates signals from the two channels to remove non-biological variation introduced by various processing steps.

Usage

```
seqWithinNorm(marraySet, y = "maM", subset = TRUE, loss.fun = square,
A = c("loess", "rlm", "median", "none"),
PT = c("median", "rlm", "loess", "none"),
PL = c("median", "rlm", "loess", "none"),
Spatial2D = c("none", "aov2D", "rlm2D", "loess2D", "spatialMedian"),
criterion = c("BIC", "AIC"))
```

Arguments

`marraySet`	Object of class `marrayRaw` or class `marrayNorm`, containing intensity data for the batch of arrays to be normalized.
`y`	Name of accessor method for spot statistics, usually the log-ratio `maM`.
`subset`	A "logical" or "numeric" vector indicating the subset of points used to compute the normalization values.
`loss.fun`	The loss function used in calculating deviance, the default uses squared sum of residuals; for absolute sum of residuals, use `abs`
`A`	A character string specifying the normalization model for the adjustment of intensity or A bias:

loess: global intensity or A-dependent robust nonlinear normalization using the `loess` function

rlm: global intensity or A-dependent robust linear normalization using the `rlm` function

median: global median location normalization

none: no normalization for the A bias

If not specified, `loess` normalization will be applied.

`PT` A character string specifying the normalization model for the adjustment of print-tip or PT bias:

median: within-print-tip-group median normalization

rlm: within-print-tip-group robust linear normalization using the `rlm` function

loess: within-print-tip-group robust nonlinear normalization using the `loess` function

none: no normalization for the PT bias

If not specified, `median` normalization within print-tip will be applied.

`PL` A character string specifying the normalization model for the adjustment of well-plate or PL bias:

median: within-well-plate median normalization

rlm: within-well-plate robust linear normalization using the `rlm` function

loess: within-well-plate robust nonlinear normalization using the `loess` function

none: no normalization for the PL bias

If not specified, `median` normalization within well-plate will be applied.

`Spatial2D` A character string specifying the normalization model for the adjustment of spatial 2D bias:

none: no normalization for the spatial 2D bias

aov2D: spatial bivariate location normalization using ANOVA

rlm2D: spatial bivariate location normalization using the `rlm` function

loess2D: spatial bivariate location normalization using the `loess` function

spatialMedian: spatial location normalization using a spatial median approach (see Wilson et al. (2003) in reference)

If not specified, no normalization will be carried out in this step.

`criterion` Character string specifying the criterion:

AIC: the AIC criterion is used; see `calcAIC`.

BIC: the BIC criterion is used; see `calcBIC`.

If no specification, `BIC` is used. Note that here we don't use the criterion to choose normalization model in each step. Criterion is calculated solely for informaion purpose.

Details

Typical systematic non-biological variations of a two-color cDNA microarray include the dependence of ratio measurements (M) on intensity (A), print-tip IDs (PT), plate IDs (PL) and spatial heterogeneity of the slide (Spatial 2D). The sequential normalization procedure in `seqWithinNorm` normalizes a slide in a sequential fashion: A -> PT -> PL -> Spatial2D. In each step one kind of variation is targeted for correction, and the user chooses the normalization method as desired. We calculate the AIC/BIC criterion along the normalization steps, but they are not used for selection of models.

Value

An object of class "list":

normdata	an object of class marrayNorm, containing the normalized intensity data.
res	a list of the sequential normalization result for each slide within the marray dataset. Each list component is also a list containing the name of the biases, deviance, equivalent number of parameters, AIC/BIC value for a certain slide.

Author(s)

Yuanyuan Xiao, <yxiao@itsa.ucsf.edu>,
Jean Yee Hwa Yang, <jean@biostat.ucsf.edu>

References

Y. H. Yang, S. Dudoit, P. Luu, and T. P. Speed (2001). Normalization for cDNA microarray data. In M. L. Bittner, Y. Chen, A. N. Dorsel, and E. R. Dougherty (eds), *Microarrays: Optical Technologies and Informatics*, Vol. 4266 of *Proceedings of SPIE*.

D. L. Wilson, M. J. Buckley, C. A. Helliwell and I. W. Wilson (2003). New normalization methods for cDNA microarray data. *Bioinformatics*, Vol. 19, pp. 1325-1332.

See Also

stepWithinNorm, withinNorm, fitWithin, fit2DWithin, calcAIC, calcBIC.

Examples

```
# Examples use swirl dataset, for description type ? swirl
data(swirl)

# Apply sequential normalization for the first slide
# default: loess(A)->median(PT)->median(PL)-> none (Spatial2D)
## Not run:
res.swirl1 <- seqWithinNorm(swirl[,1])

# normalized data
norm.swirl <- res.swirl1[[1]]

# sequential normalization information
step.swirl <- res.swirl1[[2]]


## End(Not run)
# median(A)->median(PT)->median(PL)->none(Spatial2D)
res.swirl <- seqWithinNorm(swirl[,1], A="median",PT="median",PL="median",Spatial2D="none")
```

stepNorm-internal *Internal stepNorm functions*

Description

Internal stepNorm functions.

Usage

```
calcEnp(X)
noFit(y.fun="maM", x.fun="maA")
medfit(x, y, subset=TRUE)
loessfit(x, y, span=0.4, subset=TRUE, degree=1, family="symmetric",
 control=loess.control(trace.hat="approximate", iteration=5, surface="direct"),...)
rlmfit(x, y, subset=TRUE)
loess2Dfit(x1, x2, y, span=0.2, subset=TRUE, degree=1, family="symmetric",
        control=loess.control(trace.hat="approximate", iteration=5, surface="direct"),...)
aov2Dfit(x1, x2, y, subset=TRUE)
rlm2Dfit(x1, x2, y, subset=TRUE)
spatialMedfit(x1, x2, y, subset=TRUE, width = 11,  height = width)
MedianSmooth(m, width, height=width)
square(x)
```

Details

These are not to be called directly by the user.

stepWithinNorm *Stepwise within-slide normalization function*

Description

This function conducts cDNA microarray normalization in a stepwise fashion. In a two-color cDNA array setting, within-slide normalization calibrates signals from the two channels to remove non-biological variation introduced by various processing steps.

Usage

```
stepWithinNorm(marraySet, subset=TRUE, wf.loc, criterion = c("BIC", "AIC"), loss.fun = square)
```

Arguments

marraySet	Object of class marrayRaw or class marrayNorm, containing intensity data for the batch of arrays to be normalized.
subset	A "logical" or "numeric" vector indicating the subset of points used to compute the normalization values.
wf.loc	Object of class list, each component is a step for the removal of a particular systematic variation. Typically each step is also a list of several candidate models of different complexity, the best model will be chosen by the criterion specified. For a user friendly way of constructing such a list, consult the function makeStepList.If missing, the default procedure will be used, which we consider appropriate for most slides. See details for how to specify the list and how it is used.
criterion	Character string specifying the criterion used for the selection of the best normalization procedure in each step. This argument can be specified using the first letter of each method; if no specification is made, the default is BIC: **AIC:** the AIC criterion is used **BIC:** the BIC criterion is used.
loss.fun	loss function; default set at using residual sum of squares.

Details

Typical systematic non-biological variations of a two-color cDNA microarray include the dependence of ratio measurements (M) on intensity (A), print-tip IDs (PT), plate IDs (PL) and spatial heterogeneity of the slide (SP). The stepwise normalization procedure normalizes a slide in a stepwise fashion. In each step one kind of variation is targeted for correction. Within each step, various candidate models are assessed for their adequacy with respect to the observed data. The assessment is made based on a common model selection criterion, AIC (see calcAIC) or BIC (see calcBIC), and the best model is then chosen for the specified step.

The argument wf.loc is a list of steps. Each step is also a list of models. The user uses the function fitWithin or fit2DWithin to specify a model. Below is a table of how to do so:

systematic variation	model	function
intensity (A)	median	fitWithin(fun="medfit")
A	robust linear	fitWithin(fun="rlmfit")
A	robust nonlinear	fitWithin(fun="loessfit")
print-tip (PT)	median	fitWithin(z.fun="maPrintTip", fun="medfit")
PT	robust linear	fitWithin(z.fun="maPrintTip", fun="rlmfit")
PT	robust nonlinear	fitWithin(z.fun="maPrintTip",fun="loessfit")
plate (PL)	median	fitWithin(z.fun="maCompPlate", fun="medfit")
PL	robust linear	fitWithin(z.fun="maComplate", fun="rlmfit")
PL	robust nonlinear	fitWithin(z.fun="maCompPlate", fun="loessfit")
spatial (SP)	robust linear	fit2DWithin(fun="rlm2Dfit")
SP	robust nonlinear(span=0.2)	fit2DWithin(fun="loess2Dfit", span=0.2)
SP	anova	fit2DWithin(fun="aov2Dfit")
SP	spatial median (11X11)	fit2DWithin(fun="spatialMedfit", width=11)

If the wf.loc is not specified by the user, the default procedure conducts normalization in four steps: A -> PT -> PL -> SP and models are as described in the table above. The user can choose not to follow such a procedure by passing in a different list, however we advocate normalizing the intensity (A) variation first as it is usually the source of most variation in most slides. The list can be easier specified using the function makeStepList by inputing models as character strings, see makeStepList for details.

Value

An object of class "list":

normdata	an object of class marrayNorm, containing the normalized intensity data.
res	a dataframe of the stepwise normalization result, containing the name of the model chosen for each step, deviance, equivalent number of parameters, AIC/BIC value.

Author(s)

Yuanyuan Xiao, <yxiao@itsa.ucsf.edu>,
Jean Yee Hwa Yang, <jean@biostat.ucsf.edu>

References

Y. H. Yang, S. Dudoit, P. Luu, and T. P. Speed (2001). Normalization for cDNA microarray data. In M. L. Bittner, Y. Chen, A. N. Dorsel, and E. R. Dougherty (eds), *Microarrays: Optical Technologies and Informatics*, Vol. 4266 of *Proceedings of SPIE*.

D. L. Wilson, M. J. Buckley, C. A. Helliwell and I. W. Wilson (2003). New normalization methods for cDNA microarray data. *Bioinformatics*, Vol. 19, pp. 1325-1332.

See Also

seqWithinNorm, withinNorm, fitWithin, fit2DWithin, calcAIC, calcBIC.

Examples

```
# Examples use swirl dataset, for description type ? swirl
data(swirl)

# Apply stepwise normalization for the first slide
res.swirl1 <- stepWithinNorm(swirl[,1])

# normalized data
norm.swirl <- res.swirl1[[1]]

# stepwise procedure
step.swirl <- res.swirl1[[2]]

# using a stepwise procedure different than the default
# corrects intensity (A) and print-tip (PT), this can be
# carried out in two ways:
# 1)
```

```
steps <- list(
     wholeChipA = list(med = fitWithin(fun="medfit"),
                               rlm = fitWithin(fun="rlmfit"),
                               loess = fitWithin(fun="loessfit")),
            printTipA = list(med = fitWithin(z.fun="maPrintTip", fun="medfit"),
                                rlm = fitWithin(z.fun="maPrintTip", fun="rlmfit"),
                                loess = fitWithin(z.fun="maPrintTip",fun="loessfit")))

#2)
steps <- makeStepList(PL=NULL, Spatial2D=NULL)
## Not run:
res.swirl <- stepWithinNorm(swirl[,1], wf.loc=steps)
## End(Not run)


# using AIC criterion for the first slide
## Not run:
res.swirl <- stepWithinNorm(swirl[,1], criterion="A")
## End(Not run)
```

`withinNorm` *Within-slide normalization function for cDNA spotted microarrays*

Description

This function is a wrapper function around `fitWtihin` and `fit2DWithin`. It allows the user to choose from a set of thirteen basic location normalization procedures. The function operates on an object of class `marrayRaw` or `marrayNorm` and returns an object of class `marrayNorm`.

Usage

```
withinNorm(marraySet,  y = "maM", subset = TRUE, norm = c("none",
    "median", "rlm", "loess", "medianPrintTip", "rlmPrintTip",
    "loessPrintTip", "medianPlate", "rlmPlate", "loessPlate",
    "aov2D", "rlm2D", "loess2D", "spatialMedian"), ...)
```

Arguments

`marraySet`	Object of class `marrayRaw` or class `marrayNorm`, containing intensity data for the batch of arrays to be normalized.
`y`	Name of accessor method for spot statistics, usually the log-ratio `maM`.
`subset`	A "logical" or "numeric" vector indicating the subset of points used to compute the normalization values.
`norm`	A character string specifying the normalization procedures: **none:** no normalization **median:** global median location normalization

rlm: global intensity or A-dependent robust linear normalization using the `rlm` function

loess: global intensity or A-dependent robust nonlinear normalization using the `loess` function

medianPrintTip: within-print-tip-group median normalization

rlmPrintTip: within-print-tip-group intensity or A-dependent robust linear normalization using the `rlm` function

loessPrintTip: within-print-tip-group intensity or A-dependent robust nonlinear normalization using the `loess` function

medianPlate: within-well-plate-group median normalization

rlmPlate: within-well-plate-group intensity or A-dependent robust linear normalization using the `rlm` function

loessPlate: within-well-plate-group intensity or A-dependent robust nonlinear normalization using the `loess` function

aov2D: spatial bivariate location normalization using ANOVA

rlm2D: spatial bivariate location normalization using the `rlm` function

loess2D: spatial bivariate location normalization using the `loess` function

spatialMedian: spatial location normalization using a spatial median approach (see Wilson et al. (2003) in reference)

... Misc arguments for the specified `norm` function

Details

The function `withinNorm` dispatches to the function `fitWithin` or `fit2DWithin` with specified arguments according to the choice of `norm`. For instance, when `norm="loess"` for global intensity dependent robust nonlinear normalization, `withinNorm` calls `fitWithin(fun="loess")` with the default span parameter set at 0.4. If a different span is preferred, it should be input by `span=0.2` through the argument `...` in the `withinNorm` function (see example below). For more details see `fitWithin`, `fit2DWithin` and individual fitting functions such as `loessfit`.

Value

An object of class `marrayNorm`, containing the normalized intensity data.

Author(s)

Yuanyuan Xiao, <yxiao@itsa.ucsf.edu>,
Jean Yee Hwa Yang, <jean@biostat.ucsf.edu>

References

Y. H. Yang, S. Dudoit, P. Luu, and T. P. Speed (2001). Normalization for cDNA microarray data. In M. L. Bittner, Y. Chen, A. N. Dorsel, and E. R. Dougherty (eds), *Microarrays: Optical Technologies and Informatics*, Vol. 4266 of *Proceedings of SPIE*.

D. L. Wilson, M. J. Buckley, C. A. Helliwell and I. W. Wilson (2003). New normalization methods for cDNA microarray data. *Bioinformatics*, Vol. 19, pp. 1325-1332.

See Also

seqWithinNorm, stepWithinNorm, fitWithin, fit2DWithin, loessfit, rlmfit.

Examples

```
# Examples use swirl dataset, for description type ? swirl
data(swirl)

# Apply loess normalization for the first slide, span=0.4
## Not run:
res.swirl1 <- withinNorm(swirl[,1], norm="loess")
## End(Not run)

# Apply loess normalization for the first slide, span=0.2
## Not run:
res.swirl1 <- withinNorm(swirl[,1], norm="loess", span=0.2)
## End(Not run)
```

Index

∗ **classes**
  marrayFit-class, 13
∗ **internal**
  stepNorm-internal, 17
∗ **manip**
  calcAIC, 2
  calcBIC, 3
  maCompPlate2, 9
∗ **models**
  fit2DWithin, 5
  fitWithin, 7
  makeStepList, 11
  seqWithinNorm, 14
  stepWithinNorm, 17
  withinNorm, 20

abs, *2*, *4*, *14*
AIC, *3*, *4*
aov2Dfit *(*stepNorm-internal*)*, 17

calcAIC, 2, *4*, *15*, *16*, *18*, *19*
calcBIC, *3*, 3, *15*, *16*, *18*, *19*
calcEnp *(*stepNorm-internal*)*, 17

deviance, *3*, *4*

factor, *10*
fit2DWithin, 5, *9*, *13*, *16*, *18*, *19*, *21*, *22*
fitWithin, *6*, 7, *13*, *16*, *18*, *19*, *21*, *22*

list, *18*
lm, *5*
loess, *5*, *6*, *8*, *11*–*15*, *21*
loess2Dfit *(*stepNorm-internal*)*, 17
loessfit, *21*, *22*
loessfit *(*stepNorm-internal*)*, 17

maCompPlate, *8*–*10*
maCompPlate2, 9
makeStepList, 11, *18*, *19*
maPrintTip, *8*
marrayFit, *2*, *4*, *6*, *8*
marrayFit *(*marrayFit-class*)*, 13
marrayFit-class, 13
marrayLayout, *10*
marrayNorm, *5*–*8*, *10*, *14*, *16*, *18*–*21*
marrayRaw, *5*–*8*, *10*, *14*, *18*, *20*
medfit *(*stepNorm-internal*)*, 17
MedianSmooth *(*stepNorm-internal*)*, 17

noFit *(*stepNorm-internal*)*, 17

rlm, *5*, *8*, *11*, *12*, *14*, *15*, *21*
rlm2Dfit *(*stepNorm-internal*)*, 17
rlmfit, *22*
rlmfit *(*stepNorm-internal*)*, 17

seqWithinNorm, 14, *19*, *22*
spatialMedfit *(*stepNorm-internal*)*, 17
square *(*stepNorm-internal*)*, 17
step, *2*, *4*
stepNorm-internal, 17
stepWithinNorm, *11*, *12*, *16*, 17, *22*

withinNorm, *16*, *19*, 20