

Package ‘polyICSFlow’

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Type Package

Title Identifying the Frequency of Polyfunctional Antigen-Specific T cells in ICS Flow Cytometry Data

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Description polyICSFlow systematically identifies all cytokine combinations detected in an intracellular cytokine staining (ICS) flow cytometry assay and quantifies polyfunctional antigen-specific responses to single or multiple antigens. The package also offers versatile plotting options for efficient data exploration and visualization of cytokine co-expression profiles.

URL <https://github.com/SoegaardLab/polyICSFlow>

BugReports <https://github.com/SoegaardLab/polyICSFlow/issues>

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Contents

assignMarkerCombinations	2
calcPolyfunctionality	3
extractCellCounts	6
getMarkerPositivity	7
plotCellCountsAtLeast	8
plotCellCountsExactly	10
plotMarkerComb2DScatters	11
plotMarkerCombHeatmap	13
%>%	14

Index	15
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assignMarkerCombinations

Assing each cell to a combination of gated markers.

Description

Assing each cell to a combination of gated markers.

Usage

```
assignMarkerCombinations(data_markerPos, mode = c("simple", "exhaustive"))
```

Arguments

- | | |
|----------------|--|
| data_markerPos | A data.frame where each row is a cell and each column is a gated marker describing whether a cell is included in the gate, as constructed by the getMarkerPositivity function. |
| mode | Character string specifying the level of detail in the polyfunctionality assignment. If simple, the output includes only MarkerComb (exact marker combinations), MarkerCount (number of functions), and Functionality (mono/polyfunctional). If exhaustive, the output additionally contains all "atleast_n" columns, which are logical describing whether cells have at least 1, 2, 3, ... <i>n</i> markers in any combination. Default = simple. |

Value

The input data.frame with three extra columns describing

- MarkerComb: A factor describing which marker combination a specific cell has. Describes the exact combinations, which are mutually exclusive (e.g., 4 markers $\rightarrow 2^4 = 16$ combinations).
- MarkerCount: An integer describing the number of functions of the cell (count of positive markers).

- **Functionality:** A factor grouping MarkerCount) into three categories; No cytokines (MarkerCount == 0), Monofunctional (MarkerCount == 1), or Polyfunctional (MarkerCount > 1).

See Also

[getMarkerPositivity](#), [calcPolyfunctionality](#)

Examples

```
# Define path to data
path_data <- system.file("extdata", package = "polyICSFlow")

# Read fcs files as flowSet
fs <- flowCore::read.flowSet(path = path_data,
                             pattern = ".fcs",
                             truncate_max_range = FALSE,
                             transformation = FALSE)

# Create GatingSet
gs <- flowWorkspace::GatingSet(fs)

# Apply GatingTemplate
gt <- openCyto::gatingTemplate(file.path(path_data, "gatingTemp_cytokines.csv"))
openCyto::gt_gating(x = gt, y = gs)

# Get marker positivity per cell
df_markerPos <- getMarkerPositivity(x = gs,
                                    gate_names = c("IFN+", "TNFa+", "IL2+", "CD107a+"))

# Assign marker combinations to each cell based on marker positivity
# Simple mode
df_markerComb <- assignMarkerCombinations(data_markerPos = df_markerPos,
                                           mode = "simple")

# Exhaustive mode
df_markerComb <- assignMarkerCombinations(data_markerPos = df_markerPos,
                                           mode = "exhaustive")
```

calcPolyfunctionality *Calculate the frequency of polyfunctional cells*

Description

Calculate the frequency of cells positive for each marker combination within a defined population and subtract the corresponding background. Negative values are set to zero.

Usage

```
calcPolyfunctionality(
  x,
  md_cols,
  pop_col = "cell_type",
  resolution = "MarkerComb",
  condition_col = "Stimulation",
  background_val = "unstimulated"
)
```

Arguments

<code>x</code>	A dataframe generated by the assignMarkerCombinations function, containing marker combination data along with associated metadata.
<code>md_cols</code>	A character vector specifying the names of metadata columns in <code>x</code> . These columns provide contextual information (e.g., participant ID, sample ID, time point).
<code>pop_col</code>	A string giving the name of the column in <code>x</code> that contains the population of interest. The function calculates the percentage of positive cells for each marker combination within this population. Can be obtained from any source, e.g., annotated metacluster IDs from FlowSOM clustering or cell labels from manual gating (see GetFlowJoLabels).
<code>resolution</code>	A string specifying the column in <code>x</code> that defines polyfunctionality, e.g., "MarkerComb", "MarkerCount", or "Functionality". Default is "MarkerComb". Can also refer to any of the logical columns generated if mode = "exhaustive" was used in assignMarkerCombinations , or to custom resolutions created by combining outputs from assignMarkerCombinations . For example, <code>atleast_IFN+TNFa+ == TRUE & MarkerCount == 3</code> refers to cells with three functions positive for at least IFN and TNFa. Users must first create the logical column in <code>x</code> (e.g., using <code>dplyr::mutate()</code>) before specifying it here.
<code>condition_col</code>	A string giving the name of the column in <code>x</code> containing the stimulation condition (e.g., antigen-stimulated, unstimulated). Default = "Stimulation".
<code>background_val</code>	A value from <code>condition_col</code> indicating the background condition (e.g., "unstimulated"). This is used as the reference for stimulation conditions. Default = "unstimulated".

Value

A dataframe similar to `x`, with the addition of four new columns:

- `n_cells_pos`: Number of positive cells
- `n_cells_tot`: Number of total cells
- `perc_pos`: Percentage positive cells, i.e. $(n_cells_pos/n_cells_tot)*100$
- `perc_pos_backgroundSubtracted`: Percentage positive cells with corresponding background subtracted. Negative values are set to zero.

See Also

[getMarkerPositivity](#), [assignMarkerCombinations](#)

Examples

```
# Define path to data
path_data <- system.file("extdata", package = "polyICSFlow")

# Read fcs files as flowSet
fs <- flowCore::read.flowSet(path = path_data,
                             pattern = ".fcs",
                             truncate_max_range = FALSE,
                             transformation = FALSE)

# Create GatingSet
```



```
# Calculate by logical Polyfunctional, at least marker
result <- calcPolyfunctionality(x = df,
                               md_cols = c("ID"),
                               pop_col = "cell_type",
                               resolution = "Polyfunctional_atleast_IFN+",
                               condition_col = "Stimulation",
                               background_val = "Unstim")

# Calculate by custom resolution
df2 <- df %>%
  dplyr::mutate(MarkerCountAbove3_atleast_IFN_TNFa = MarkerCount == 3 &
               .data[["atleast_2_IFN+TNFa+"]])

result <- calcPolyfunctionality(x = df2,
                               md_cols = c("ID"),
                               pop_col = "cell_type",
                               resolution = "MarkerCountAbove3_atleast_IFN_TNFa",
                               condition_col = "Stimulation",
                               background_val = "Unstim")
```

extractCellCounts	<i>Get number of cells positive for each distinct marker combination</i>
-------------------	--

Description

Get number of cells in your dataset positive for each distinct marker combination (MarkerComb), number of functions (MarkerCount), and Functionality.

Usage

```
extractCellCounts(
  data_markerComb,
  cell_subset_params = list(cellTypes = NULL, cellTypes_plot = NULL)
)
```

Arguments

data_markerComb
A dataframe generated by the [assignMarkerCombinations](#) function, containing marker combination data.

cell_subset_params
List with parameters describing what subset of cells (cellTypes_plot) should be counted, from a vector of cell types (cellTypes). Default = `list(cellTypes = NULL, cellTypes_plot = NULL)`, i.e. all cells in the dataset are included in the cell count.

Value

A grouped tibble with cell counts of the input data grouped by "MarkerComb", "MarkerCount", and "Functionality".

Examples

```
# Define path to data
path_data <- system.file("extdata", package = "polyICSFlow")

# Read fcs files as flowSet
fs <- flowCore::read.flowSet(path = path_data,
                             pattern = ".fcs",
                             truncate_max_range = FALSE,
                             transformation = FALSE)

# Create GatingSet
gs <- flowWorkspace::GatingSet(fs)

# Apply GatingTemplate
gt <- openCyto::gatingTemplate(file.path(path_data, "gatingTemp_cytokines.csv"))
openCyto::gt_gating(x = gt, y = gs)

# Get marker positivity per cell
df_markerPos <- getMarkerPositivity(x = gs,
                                    gate_names = c("IFN+", "TNFa+", "IL2+", "CD107a+"))

# Assign marker combinations to each cell based on marker positivity
df_markerComb <- assignMarkerCombinations(data_markerPos = df_markerPos,
                                          mode = "exhaustive")

# Get cell counts from all data
cell_counts <- extractCellCounts(data_markerComb = df_markerComb)

# Get cell counts from subset of data (metacluster 2 and 4)
metacluster_labels <- readRDS(file.path(path_data, "cell_labels.rds"))
cell_counts <- extractCellCounts(data_markerComb = df_markerComb,
                                cell_subset_params = list(cellTypes = metacluster_labels,
                                                           cellTypes_plot = c(2,4)))
```

getMarkerPositivity *Create dataframe of marker positivity*

Description

Create a dataframe describing whether each cell is included by the gates where each row is a cell and each column is a marker.

Usage

```
getMarkerPositivity(x, gate_names = NULL)
```

Arguments

x	An object of class GatingHierarchy or GatingSet with an applied gatingTemplate.
gate_names	The names of the cytokine gates. If NULL (default), all leaf nodes will be used.

Value

A data.frame where each row is a cell and each column is a gated marker which can be used as input for the [assignMarkerCombinations](#) function.

References

This code is strongly based on the [GetFlowJoLabels](#) function.

See Also

[GatingSet](#), [gatingTemplate](#), [gt_gating](#), [assignMarkerCombinations](#), [calcPolyfunctionality](#)

Examples

```
# Define path to data
path_data <- system.file("extdata", package = "polyICSFlow")

# Read fcs files as flowSet
fs <- flowCore::read.flowSet(path = path_data,
                             pattern = ".fcs",
                             truncate_max_range = FALSE,
                             transformation = FALSE)

# Create GatingSet
gs <- flowWorkspace::GatingSet(fs)

# Apply GatingTemplate
gt <- openCyto::gatingTemplate(file.path(path_data, "gatingTemp_cytokines.csv"))
openCyto::gt_gating(x = gt, y = gs)

# Get marker positivity per cell
df_markerPos <- getMarkerPositivity(x = gs,
                                    gate_names = c("IFN+", "TNFa+", "IL2+", "CD107a+"))
```

`plotCellCountsAtLeast` *Plot counts of cells expressing at least 1, 2, ..., n markers.*

Description

Plot a stacked bar plot of cell counts by marker combination, faceted by the minimum number of markers co-expressed ($\geq 1, \geq 2, \dots, \geq n$). Within each facet, the x-axis shows all marker or marker-combination categories corresponding to that level (e.g., single markers for ≥ 1 , two-marker combinations for ≥ 2 , etc.).

Usage

```
plotCellCountsAtLeast(
  data_markerComb,
  cell_subset_params = list(cellTypes = NULL, cellTypes_plot = NULL),
  palette_comb = NULL
)
```

Arguments

data_markerComb

A dataframe generated by the `assignMarkerCombinations` function, containing marker combination data.

cell_subset_params

List with parameters describing what subset of cells (`cellTypes_plot`) should be plotted, from a vector of cell types (`cellTypes`). Default = `list(cellTypes = NULL, cellTypes_plot = NULL)`, i.e. all cells in the dataset are plotted.

palette_comb

A named character vector mapping marker combination names (as produced by `assignMarkerCombinations`) to colors. If NULL (default), a qualitative palette is generated with `colorRampPalette` and `CATALYST:::cluster_cols`.

Value

A ggplot.

See Also

plotCellCountsExactly

Examples

[illegible]

`plotCellCountsExactly` *Plot counts of cells expressing exactly 1, 2, ..., n markers.*

Description

Plot a stacked bar plot of cell counts within each marker combination for cells positive for exactly 1, 2, ..., n markers.

Usage

```
plotCellCountsExactly(
  data_markerComb,
  cell_subset_params = list(cellTypes = NULL, cellTypes_plot = NULL),
  palette_comb = NULL
)
```

Arguments

`data_markerComb` A dataframe generated by the [assignMarkerCombinations](#) function, containing marker combination data.

`cell_subset_params` List with parameters describing what subset of cells (`cellTypes_plot`) should be plotted, from a vector of cell types (`cellTypes`). Default = `list(cellTypes = NULL, cellTypes_plot = NULL)`, i.e. all cells in the dataset are plotted.

`palette_comb` A named character vector mapping marker combination names (as produced by [assignMarkerCombinations](#)) to colors. If `NULL` (default), a qualitative palette is generated with [colorRampPalette](#) and `CATALYST:::cluster_cols`.

Value

A `ggplot`.

See Also

[plotCellCountsAtLeast](#)

Examples

```
# Define path to data
path_data <- system.file("extdata", package = "polyICSFlow")

# Read fcs files as flowSet
fs <- flowCore::read.flowSet(path = path_data,
                             pattern = ".fcs",
                             truncate_max_range = FALSE,
                             transformation = FALSE)

# Create GatingSet
gs <- flowWorkspace::GatingSet(fs)

# Apply GatingTemplate
```

```

gt <- openCyto::gatingTemplate(file.path(path_data, "gatingTemp_cytokines.csv"))
openCyto::gt_gating(x = gt, y = gs)

# Get marker positivity per cell
df_markerPos <- getMarkerPositivity(x = gs,
                                   gate_names = c("IFN+", "TNFa+", "IL2+", "CD107a+"))

# Assign marker combinations to each cell based on marker positivity
df_markerComb <- assignMarkerCombinations(data_markerPos = df_markerPos,
                                          mode = "simple")

# Plot all cells
plotCellCountsExactly(data_markerComb = df_markerComb)

# Plot only subset of data (metacluster 2 and 4)
metacluster_labels <- readRDS(file.path(path_data, "cell_labels.rds"))
plotCellCountsExactly(data_markerComb = df_markerComb,
                      cell_subset_params = list(cellTypes = metacluster_labels,
                                                cellTypes_plot = c(2,4)))

```

plotMarkerComb2DScatters

Plot 2D scatterplots of all marker combinations.

Description

Plots 2D scatter plots of all possible marker combinations, where each panel (facet) corresponds to one of the 2^n possible combinations of n markers. The top-left panel represents cells negative for all markers, while the bottom-right represents cells positive for all markers, with intermediate panels showing mixed combinations. Within each panel, the x-axis denotes the n markers and the y-axis shows the transformed MFI values. Each point corresponds to a single cell, with cells belonging to the combination represented in that facet highlighted in color, and all other cells displayed in grey background for reference.

Usage

```

plotMarkerComb2DScatters(
  input,
  assigned_MarkerCombs,
  cell_subset_params = list(cellTypes = NULL, cellTypes_plot = NULL),
  palette_comb = NULL,
  return_list = FALSE
)

```

Arguments

input	Either a matrix of marker expression or an object of class SingleCellExperiment , GatingSet , or flowSet , from which the expression data can be extracted. If input is either GatingSet or flowSet, an aggregate of the data is created with the AggregateFlowFrames function.
assigned_MarkerCombs	A vector of assigned marker combinations per cell in x, as created by assignMarkerCombinations .

cell_subset_params	List with parameters describing what subset of cells (cellTypes_plot) should be plotted, from a vector of cell types (cellTypes). Default = list(cellTypes = NULL, cellTypes_plot = NULL), i.e. all cells in the dataset are plotted.
palette_comb	A named character vector mapping marker combination names (as produced by assignMarkerCombinations) to colors. If NULL (default), a qualitative palette is generated with colorRampPalette and CATALYST:::cluster_cols.
return_list	Logical. If FALSE (default), the plots are combined by cowplot. If TRUE, a list of plots are returned to allow further customization.

Value

A [ggplot](#) object visualizing 2D scatterplots of all marker combinations.

Examples

```
# Define path to data
path_data <- system.file("extdata", package = "polyICSFlow")

# Read fcs files as flowSet
fs <- flowCore::read.flowSet(path = path_data,
                             pattern = ".fcs",
                             truncate_max_range = FALSE,
                             transformation = FALSE)

# Create GatingSet
gs <- flowWorkspace::GatingSet(fs)

# Apply GatingTemplate
gt <- openCyto::gatingTemplate(file.path(path_data, "gatingTemp_cytokines.csv"))
openCyto::gt_gating(x = gt, y = gs)

# Get marker positivity per cell
df_markerPos <- getMarkerPositivity(x = gs,
                                    gate_names = c("IFN+", "TNFa+", "IL2+", "CD107a+"))

# Assign marker combinations to each cell based on marker positivity
df_markerComb <- assignMarkerCombinations(data_markerPos = df_markerPos)

# Plot 2D scatter of marker combinations in dataset (all data)
# From gatingSet
plotMarkerComb2DScatters(input = gs,
                          assigned_MarkerCombs = df_markerComb$MarkerComb)

# From flowSet
plotMarkerComb2DScatters(input = fs,
                          assigned_MarkerCombs = df_markerComb$MarkerComb)

# Plot only subset of data (metacluster 2 and 4)
metacluster_labels <- readRDS(file.path(path_data, "cell_labels.rds"))
plotMarkerComb2DScatters(input = fs,
                          assigned_MarkerCombs = df_markerComb$MarkerComb,
                          cell_subset_params = list(cellTypes = metacluster_labels,
                                                    cellTypes_plot = c(2,4)))

# As list
plot_list <- plotMarkerComb2DScatters(input = fs,
```

```
assigned_MarkerCombs = df_markerComb$MarkerComb,
return_list = TRUE)
```

plotMarkerCombHeatmap *Plot a heatmap of all possible marker combinations.*

Description

Plot a heatmap of all possible marker combinations.

Usage

```
plotMarkerCombHeatmap(
  marker_names,
  orientation = c("vertical", "horizontal"),
  palette_count = NULL,
  palette_comb = NULL,
  full_names = TRUE
)
```

Arguments

marker_names	A character vector specifying the marker names used to generate all possible combinations.
orientation	A string specifying the orientation of the heatmap, either vertical or horizontal. Default = "vertical".
palette_count	A named character vector mapping the number of markers per cell to colors. If NULL (default), a qualitative palette is generated with brewer_pal .
palette_comb	A named character vector mapping marker combination names to colors. If NULL (default), a qualitative palette is generated with <code>CATALYST:::cluster_cols</code> .
full_names	Logical. If TRUE (default) the marker combinations are displayed with their full names (i.e., negative markers are also present). If FALSE, only the positive markers are displayed.

Value

A [Heatmap](#) object visualizing all marker combinations. The heatmap includes two layers of annotation:

- MarkerCount: A marker count annotation (colored with `palette_count`) showing the number of markers in each combination.
- MarkerComb: A combination annotation (colored with `palette_comb`) showing the exact subset of markers present.

See Also

[Heatmap](#)

Examples

```
# Vertical heatmap
plotMarkerCombHeatmap(marker_names = c("IFN", "TNFa", "IL2", "CD107a"),
  orientation = "vertical")

# Horizontal heatmap
plotMarkerCombHeatmap(marker_names = c("IFN", "TNFa", "IL2", "CD107a"),
  orientation = "horizontal")
```

%>%	<i>Pipe operator</i>
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Description

See [%>%](#) for details.

Index

* **internal**

%>%, [14](#)

%>%, [14](#), [14](#)

AggregateFlowFrames, [11](#)

assignMarkerCombinations, [2](#), [4](#), [6](#), [8–12](#)

brewer_pal, [13](#)

calcPolyfunctionality, [3](#), [3](#), [8](#)

colorRampPalette, [9](#), [10](#), [12](#)

extractCellCounts, [6](#)

flowSet, [11](#)

GatingSet, [8](#), [11](#)

gatingTemplate, [8](#)

GetFlowJoLabels, [4](#), [8](#)

getMarkerPositivity, [2–4](#), [7](#)

ggplot, [12](#)

gt_gating, [8](#)

Heatmap, [13](#)

plotCellCountsAtLeast, [8](#), [10](#)

plotCellCountsExactly, [9](#), [10](#)

plotMarkerComb2DScatters, [11](#)

plotMarkerCombHeatmap, [13](#)

SingleCellExperiment, [11](#)