

Package ‘DenoIST’

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Title DenoIST: Denoising Image-based Spatial Transcriptomics data

Version 0.99.2

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Description DenoIST identifies and removes contamination in Image-based Spatial Transcriptomics data, using a transposed poisson mixture model with local neighbourhood offsets to infer genes that are likely to be due to neighbourhood contamination rather than endogenous expression.

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Encoding UTF-8

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URL <https://github.com/aaronkwc/DenoIST>

BugReports <https://github.com/aaronkwc/DenoIST/issues>

Imports flexmix, hexbin, pbapply, sparseMatrixStats,
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denoist	<i>DenoIST</i>
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Description

DenoIST (Denoising Image-based Spatial Transcriptomics) is a method for identifying and removing contamination artefacts in image-based single-cell transcriptomics (IST) data. It uses a transposed Poisson mixture model to identify contamination.

Usage

```
denoist(
  mat,
  tx,
  coords = NULL,
  tx_x = "x",
  tx_y = "y",
  feature_label = "gene",
  distance = 50,
  nbins = 200,
  posterior_cutoff = 0.6,
  cl = 1,
  out_dir = NULL,
  verbose = FALSE
)
```

Arguments

mat	A matrix of counts (genes x cells), or a SpatialExperiment object.
tx	A data frame of transcript with x, y and qv columns.
coords	A data frame of coordinates (n_cells x 2). Only used if not using a SpatialExperiment object as input.
tx_x	Column name for the x coordinates in the transcripts dataframe. Default is 'x'.
tx_y	Column name for the y coordinates in the transcripts dataframe. Default is 'y'.
feature_label	Column name for the gene of each transcript in the transcripts dataframe. Default is 'gene'.
distance	The maximum distance to consider for local background estimation.
nbins	The number of bins to use for hexagonal binning.

posterior_cutoff	The cutoff for posterior probability to determine contamination.
cl	The number of cores to use for parallel processing.
out_dir	The output directory to save the results.
verbose	Logical, if TRUE, print progress messages.

Details

The function calculates local background using hexagonal binning and applies a Poisson mixture model to identify contamination. It returns a matrix of memberships and adjusted counts for each gene in each cell.

Value

A list containing the following elements:

memberships	A matrix of memberships for each gene in each cell.
adjusted_counts	A matrix of adjusted counts for each gene in each cell.
params	A list of parameters for each gene.

Examples

```
# Load example data
set.seed(42)
mat <- matrix(rpois(1000, lambda = 10), nrow = 10, ncol = 100)
rownames(mat) <- paste0("gene", 1:10)
coords <- data.frame(x = rnorm(100), y = rnorm(100))
tx <- data.frame(x = c(rnorm(500), rnorm(500, 3)),
                 y = c(rnorm(500), rnorm(500, 3)),
                 qv = rep(30, 1000), gene = paste0('gene', 1:10))

# Run DenoIST
result <- denoist(mat, tx, coords, distance = 1, nbins = 50, cl = 1,
                  out_dir = NULL, verbose = TRUE)

# Check results
print(result$memberships[1:5, 1:5])
print(result$adjusted_counts[1:5, 1:5])
print(result$params[[1]])
```

local_offset_distance_with_background
Neighbourhood offset

Description

This function calculates the local neighbourhood offset together with ambient background for each cell in a count matrix.

Usage

```
local_offset_distance_with_background(
  mat,
  tx,
  coords,
  tx_x = "x",
  tx_y = "y",
  feature_label = "gene",
  distance = 50,
  nbins = 200,
  cl = 1,
  verbose = FALSE
)
```

Arguments

mat	A count matrix with genes as rows and cells as columns.
tx	A transcript dataframe with x, y coordinates and qv values.
coords	A dataframe with x, y coordinates of each cell as separate columns.
tx_x	Column name for the x coordinates in the transcripts dataframe.
tx_y	Column name for the y coordinates in the transcripts dataframe.
feature_label	Column name for the gene of each transcript in the transcripts dataframe.
distance	The maximum distance to consider for local background estimation.
nbins	The number of bins to use for hexagonal binning, used for calculating background transcript contamination.
cl	The number of cores to use for parallel processing.
verbose	Logical, if TRUE, print progress messages.

Details

The function calculates the offset used for each cell based on their local neighbourhoods. In most cases you do not need to use this as denoise already runs this internally but it is good for debugging if needed.

Value

A matrix of local background counts for each gene in each cell.

Examples

```
# Load example data
set.seed(42)
mat <- matrix(rpois(1000, lambda = 10), nrow = 10, ncol = 100)
rownames(mat) <- paste0("gene", 1:10)
coords <- data.frame(x = rnorm(100), y = rnorm(100))
tx <- data.frame(x = c(rnorm(500), rnorm(500, 3)),
                 y = c(rnorm(500), rnorm(500, 3)),
```

```

      qv = rep(30, 1000), gene = paste0('gene', 1:10))
# Run DenoIST
off_mat <- local_offset_distance_with_background(mat, tx, coords,
                                                distance = 1, nbins = 50,
                                                cl = 1, verbose = TRUE)

# Check results
print(off_mat[1:5, 1:5])

```

solve_poisson_mixture *Poisson mixture model solver*

Description

This function implements a 2-component Poisson mixture model using the EM algorithm. It estimates the parameters of the model and assigns memberships to each observation based on the posterior probabilities.

Usage

```

solve_poisson_mixture(
  x,
  s,
  max_iter = 5000,
  tol = 1e-06,
  pi_inits = runif(10, min = 0, max = 0.5),
  posterior_cutoff = 0.6,
  verbose = FALSE
)

```

Arguments

x	A vector of counts.
s	A vector of offsets.
max_iter	Maximum number of iterations for the EM algorithm.
tol	Tolerance for convergence.
pi_inits	Initial values for the mixing proportion.
posterior_cutoff	Cutoff for posterior probabilities to assign memberships.
verbose	Logical, if TRUE, print progress messages.

Details

The function takes a vector of counts and a vector of offsets as input. It uses the EM algorithm to iteratively update the parameters of the model until convergence is reached or the maximum number of iterations is exceeded. The function also allows for multiple initialisations of the mixing proportion to find the best solution.

Value

A list containing the following elements:

Examples

```
x <- rpois(100, lambda = 5)
s <- runif(100, min = 0, max = 1)
result <- solve_poisson_mixture(x, s)
print(result)
```

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